



UNIVERSIDAD NACIONAL AUTÓNOMA DE MÉXICO

**PROGRAMA DE MAESTRÍA Y DOCTORADO EN CIENCIAS MÉDICAS,
ODONTOLÓGICAS Y DE LA SALUD**

**INSTITUTO NACIONAL DE CARDIOLOGÍA IGNACIO CHÁVEZ
CAMPO DE CONOCIMIENTO DE CARDIOLOGÍA**

**THE EDN-1 GLU105GLU (RS5369) GENE POLYMORPHISM IN THE ENDOTHELIN
GENE (EDN1) IS ASSOCIATED WITH RISK OF DEVELOPING CORONARY
ARTERY DISEASE IN MEXICAN PATIENTS.**

**MODALIDAD DE GRADUACIÓN: PRODUCCION CIENTÍFICA
QUE PARA OPTAR POR EL GRADO DE MAESTRO EN CIENCIAS MEDICAS**

PRESENTA

OMAR GOMEZ MONTERROSAS

TUTOR

DR. MARCO ANTONIO MARTÍNEZ RIOS

**PROGRAMA DE MAESTRÍA Y DOCTORADO EN CIENCIAS MÉDICAS, ODONTOLÓGICAS Y DE
LA SALUD**

CIUDAD DE MEXICO, FEBRERO DEL 2017



Universidad Nacional
Autónoma de México

Dirección General de Bibliotecas de la UNAM

Biblioteca Central



UNAM – Dirección General de Bibliotecas
Tesis Digitales
Restricciones de uso

DERECHOS RESERVADOS ©
PROHIBIDA SU REPRODUCCIÓN TOTAL O PARCIAL

Todo el material contenido en esta tesis esta protegido por la Ley Federal del Derecho de Autor (LFDA) de los Estados Unidos Mexicanos (México).

El uso de imágenes, fragmentos de videos, y demás material que sea objeto de protección de los derechos de autor, será exclusivamente para fines educativos e informativos y deberá citar la fuente donde la obtuvo mencionando el autor o autores. Cualquier uso distinto como el lucro, reproducción, edición o modificación, será perseguido y sancionado por el respectivo titular de los Derechos de Autor.

“Noli foras ire, in te ipsum redi, in interiore homine habitat veritas”

Sant' Agostino

Index

1. INTRODUCTION

1.1 From endothelial dysfunction to atherosclerotic process

1.2 Endothelin 1

1.3 Endothelial nitric oxide

1.4 Single nucleotid polimorphisms

1.5 Aim of the study

2. METHODS

2.1 Study population

2.2 DNA extration

2.3 Determination of eNOS and EDN-1 genotypes

2.4 Statistical analysis

3. RESULTS

3.1 Study population

3.2 Genetic profile of study group

3.3 Predictors of ACS

4. DISCUSSION

5. LIMITATIONS OF THE STUDY

6. CONCLUSION

7. REFERENCES

8. TABLES and FIGURES

9. LEGENDS

1. INTRODUCTION

1.1 From the endothelial dysfunction to the atherosclerotic process

Atherosclerosis is a complex multifactorial and polygenic disorder resulting from endothelial dysfunction (ED) (1) and peculiar responses to various forms of injurious stimuli to the arterial wall (2).

The latest observations suggest that plaque rupture (PR) represent the most common substrate of coronary thrombosis (in nearly 50% of cases) (3,4), whereas coronary erosion is responsible for 25-30% of acute coronary syndromes (ACS) (5). A recent study showed that patients with PR as culprit lesion present a poor prognosis compared to patients with intact fibrous cap (6). Mechanisms of PR have been mainly focused on inflammation as plaque inflammation may predispose to PR by thinning of the fibrous cap due to release of metalloproteinase and to reduced collagen synthesis (7). This hypothesis is in keeping with data showing higher values of C-reactive protein (CRP) a sensitive marker of inflammation in patients with ACS as compared to patients with chronic stable angina (8). Yet, CRP levels are normal in about a half of ACS patients (9), suggesting that inflammation is probably involved in PR in a subset of patients.

On the other hand, previous studies discovered several molecules that have been associated with the vascular physiology and severity of coronary diseases (10), the most important of them are represented by Endothelin-1 (ET-1) and endothelial nitric oxide (eNOS).

1.2 Endothelin-1

Endothelin (ET-1) is encoded by the EDN1 gene located in chromosome 6p21–24, and acts as a potent vasoconstrictor and modulator of vasomotor tone and vascular remodelling (11,12). Moreover, ET-1 has the following properties: 1) it is expressed in several tissues, including endothelial cells and cardiomyocytes (13); 2) it has a pro-atherogenic activity and mediates smooth muscle cell proliferation via ET-1 receptors; 3) it also acts as a chemoattractant for monocytes, and induces platelet aggregation and expression of adhesion molecules (14-16); 4) finally, it stimulates superoxide production, NF- κ B expression, and atherosclerotic lesion formation, and inhibits endothelial nitric oxide (eNOS) (17).

1.3 Endothelial nitric oxide.

Another crucial actor of the atherosclerotic process is represented by eNOS. The gene encoding eNOS is located on chromosome 7q35–36. It is considered a protective enzyme, not only for its role in NO synthesis but also because its inhibition is associated with the production of superoxide (18,19). Yet, eNOS regulates a wide spectrum of functions in the cardiovascular system, like vasorelaxation and migration and proliferation of vascular smooth muscle cell (20). Of note, it inhibits T helper 1 type immune response and can activate directly endothelial NADPH oxidases, (21). For these reasons, several studies have assessed the correlation between endothelial expression levels of ET-1 and eNOS with the instability of the atherosclerotic plaque (22-24).

1.4 Single-nucleotide polymorphisms.

The human genome is rich in variation. These variants include single nucleotide substitutions (mutations or single nucleotide polymorphisms), insertions and deletions

(indels), copy number variants, and short tandem repeats. In many cases, the genetic variant detected is a marker for another genetic defect at a nearby locus rather than a cause of the phenotypic abnormality. Single nucleotide polymorphisms (SNPs; ie, substitutions of one nucleotide for another) represent the most abundant form of genetic variation and are responsible for much of the heritable phenotypic variation observed in human populations. Estimates based on whole genome sequencing efforts suggest that individuals carry between 2.8 and 3.9 million single-base pair variants, with higher estimates observed in individuals of African descent.

Estimates suggest that unrelated haploid genomes differ every 185 to 2000 nucleotides. This wide range is due in part to the variable rate of polymorphism across the genome, with some regions demonstrating high heterozygosity (such as the MHC cluster on chromosome 6), while others display more restricted variation.

By convention, single-base pair changes that achieve a population frequency of at least 1 percent are referred to as SNPs. Less-frequent substitutions that interrupt gene function, or that have clinical consequences, are labeled as mutations, even though many of these are observed at a fixed frequency (albeit below 1 percent) across diverse populations. Differences in the frequency distribution of SNPs result from the combination of effects of natural selection, genetic drift, and other population genetic effects. The functional impact of a variant is not determined by its frequency, but rather by its location within genes or non-genic regulatory regions. Single-base substitutions resulting in SNPs or mutations arise through shared mechanisms, primarily single-base slip mispairing during DNA replication and CpG-mediated cytosine deamination (25-27).

EDN-1 presents three important SNPs, two in the promoter region [-974CNA (rs3087459) and-1394TNG (rs1800541)] and one in the coding region [Glu105Glu (rs5369)]. The two SNPs in the promoter region have been identified in the 5'-flanking

region of the EDN-1 gene and both SNPs have been associated with risk of developing myocardial infarction, essential left ventricular hypertrophy, asthma, and rheumatoid arthritis (28-31). The SNP located in the coding region has been associated with asthma and high blood pressure (30,31). On the other hand, the eNOS gene presents two relevant SNPs in the promoter region [-786TNC (rs2070744) and-1474 ANT (rs1800783)] and one in the exon 7 of the coding region (G894T). The SNPs in the promoter region have been associated with a significant reduction in the eNOS gene promoter activity, whereas the SNP in the coding region leads to an amino acid change from Glu to Asp (Glu298Asp) and has been associated with reduced basal NO production. These eNOS variants have been associated with an increased risk for coronary artery disease, heart failure, atherosclerosis, and myocardial infarction (24, 32-36).

1.4 Aim of the study

Considering the prominent role of ET-1 and eNOS as regulators of endothelial function, inflammatory processes, and vascular physiology, they seem to have a measurable influence on the development of the atherosclerotic plaque and contribute to or increase the occurrence of ACS. In this context, the objective of this study was to establish the role of EDN-1 and eNOS gene polymorphisms in the risk of developing ACS in a group of Mexicans patients.

2. METHODS

2.1 Study population

The clinical characteristics of the study population are reported in the Table 1. We included 218 Mexican patients with ACS (148 males, mean age of 60.4 ± 11.7) referred to the National Institute of Cardiology Ignacio Chávez. Of them, EDN-1-974CNA

(rs3087459), EDN-1-1394TNG (rs1800541), and EDN-1 Glu105Glu (rs5369) single nucleotide polymorphisms were genotyped using 5' exonuclease TaqMan genotyping assays on an ABI Prism 7900 HT Fast Real time PCR System, according to manufacturer's instructions. Acute coronary syndromes were diagnosed on the basis of clinical history, physical examination with electrocardiography, chest radiography, echocardiography, and coronary angiography. ACS diagnosis was made according to the World Health Organization and the American Heart Association (AHA)/American College of Cardiology (ACC) criteria (37). As control, 204 healthy unrelated individuals (83 males, mean age of 59.4 ±9.8) with neither symptoms nor previous diagnosis of cardiovascular problems. All included subjects were ethnically matched, and we considered as Mexican Mestizos only those individuals who for three generations, including their own, had been born in Mexico. Clinical characteristics of study population are summarized in the Table 1.

2.3 DNA extraction

The DNA extraction method proposed by Lahiri and Numberger was used for DNA extraction (Lahiri and Numberger, 1991). Briefly, 500 µl of whole blood was mixed with 500 µl of TKM buffer (Tris-HCl pH 7.6 10 mM, KCl 10 mM, MgCl₂ 10 mM, EDTA 2 mM) in a microcentrifuge tube. To this, 1 ml of TKM + 2.5% Triton X-100 was added to disrupt the cell membrane without damaging nuclei. The tube was mixed by inversion (10 times) and then centrifuged at 1000 g for 8 min at 4 °C. The cytoplasmic supernatant was discarded and the nuclear pellet was resuspended in 1 ml of TKM by pipetting. Cells were again centrifuged at 1000 g for 8 min and washed with TKM once more. The final pellet was resuspended in 200 µl TKM, after which 15 µl of 10% SDS was added to lyse nuclei and mixed by inversion. Material was incubated for 5 min at 55 °C and 75 µl of

saturated NaCl solution was added. The tube was mixed by inversion and centrifuged at maximum speed (12,600 g) in an Eppendorf microcentrifuge for 5 min at room temperature. Supernatant was transferred to a fresh tube and 0.7 volume of isopropanol was added. The sample was mixed by inversion and centrifuged for 10 min at maximum speed in a microcentrifuge. Pellet was washed in cold 70% ethanol and permitted to air dry for 15 min. DNA material was resuspended in 100 µl TE at 65 °C for 1 h.

2.3 Determination of eNOS and EDN-1 genotypes

ENOS298, eNOS785, eNOS1474, EDN-1-974CNA (rs3087459), EDN-1-1394TNG (rs1800541), and EDN-1 Glu105Glu (rs5369) single nucleotide polymorphisms were genotyped using 5' exonuclease TaqMan genotyping assays on an ABI Prism 7900 HT Fast Real time PCR System, according to manufacturer's instructions.

2.4 Statistical analysis

The distribution of continuous variables was assessed by visual inspection of frequency histograms and with the use of the Shapiro–Wilk test. Continuous variables were expressed as mean \pm standard deviation (SD) or median with interquartile range, if they followed a normal or non-normal distribution, respectively. Continuous variables were compared with unpaired t-test or Mann–Whitney U-test, whereas categorical variables were compared using the chi square test or Fisher's exact test, as appropriate. Correlations between variables were performed using the Pearson test.

All variables were tested in an univariable model and then entered in the multivariable model if p-value was < 0.05 . The software SPSS 17.0 (SPSS Italy, Florence, Italy) was used for statistical analyses.

3. RESULTS

3.1 Study population

Results of the study population are summarized in the Table 1. Patients with ACS were older than healthy subjects (<0.001). Moreover, they had more frequently family history (<0.001), smoking habitus (<0.001), hypertension (<0.001) and diabetes mellitus (<0.001) compared to control group. Yet, patients with ACS presented an enhances risk of cardiovascular disease.

3.2 Genetic profile of study groups

Results of genetic profile of study population are summarized in the Table 2. An increased frequency of EDN-1 Glu105Glu (rs5369) polymorphism was found in ACS patients compared to healthy subjects ($p=0.004$) (Figure 1). Within patients with ACS, patients with hypertension showed more frequently eNOS 1474AA polymorphism compared to those without hypertension ($p=0.0022$) (figure 2, panel A). Yet, obese patients presented more frequently eNOS 1474AA ($p=0.001$), eNOS-1474T/A ($p=0.004$) and eNOS786CC ($p=0.011$) polymorphisms than those without obesity (figure 2, panel B).

3.2 Predictors of ACS

Predictors of ACS are reported in the Table 3. At univariate analysis, predictors of ACS are age, male sex, hypertension, family history of CAD, high CV risk and EDN-1 Glu105Glu (rs5369) polymorphism. At multivariable analysis, age, male sex, family history of CAD, high CV risk and EDN-1 Glu105Glu (rs5369) polymorphism were independent predictors of ACS.

4. DISCUSSION

Our study shows that, in a Mexican population, 1) compared to healthy subjects, patients with ACS exhibit an increased frequency of EDN-1 Glu105Glu (rs5369) polymorphism; 2) within patients with ACS, patients with hypertension showed more frequently eNOS 1474AA polymorphism compared to those without hypertension; 3) obese patients presented more frequently eNOS 1474AA, eNOS-1474T/A and eNOS786CC polymorphisms than those without obesity; 4) age, male sex, family history of CAD, high CV risk and EDN-1 Glu105Glu (rs5369) polymorphism were independent predictors of ACS.

Susceptibility to CAD is claimed to be 40% to 60% inherited, but until recently genetic risk factors predisposing to CAD have been elusive. Routine genetic screening is unlikely until management is improved by genetic testing. Risk variants should provide pathophysiological insights and targets for novel therapy. While risk variants are less potent predictors of CAD, compared with biomarkers, they have the advantage of not changing in one's lifetime and are unaffected by diet, sex, age, or medication. The most important genes responsible for vascular physiology and severity of coronary diseases, are represented by ET-1 and eNOS. (10).

ET-1 as a potent vasoconstrictor and modulator of vasomotortone and vascular remodelling (11,12). Yet, ET-1 has a pro-atherogenic activity and mediates smooth muscle cell proliferation via ET-1 receptors. Experimental model showed as ET-1 and ET receptors are upregulated in both human and experimental animal atherosclerotic lesions. This notion has been further supported by a recent finding that administration of ET receptor antagonists resulted in a significant reduction of atherosclerosis in apoE-KO mice. Moreover, it acts as a chemoattractant for monocytes, and induces platelet

aggregation and expression of adhesion molecules (14-16); 4) finally, it stimulates superoxide production, NF- κ B expression, and atherosclerotic lesion formation, and inhibits eNOS (17). On the other hand, eNOS is considered a protective enzyme, not only for its role in NO synthesis but also because its inhibition is associated with the production of superoxide (18,19). Vascular NO \cdot dilates all types of blood vessels by stimulating soluble guanylyl cyclase and increasing cyclic guanosine monophosphate (cGMP) in smooth muscle cells. NO \cdot released toward the vascular lumen is a potent inhibitor of platelet aggregation and adhesion. NO \cdot also can inhibit leukocyte adhesion to the vessel wall either by interfering with the ability of the leukocyte adhesion molecule CD11/CD18 to form an adhesive bond with the endothelial cell surface or by suppressing CD11/CD18 expression on leukocytes. White cell adherence is an early event in the development of atherosclerosis; therefore, NO \cdot may protect against the onset of atherogenesis. Furthermore, NO \cdot has been shown to inhibit DNA synthesis, mitogenesis, and proliferation of vascular smooth muscle cells. The inhibition of platelet aggregation and adhesion protects smooth muscle from exposure to platelet-derived growth factor(s). Therefore, NO \cdot also prevents a later step in atherogenesis, fibrous plaque formation. Based on the combination of those effects, endothelial NO \cdot probably represents the most important antiatherogenic defense principle in the vasculature (21). For these reasons, several studies have assessed the correlation between endothelial expression levels of ET-1 and eNOS with the instability of the atherosclerotic plaque. These genetic factors could act with the classical risk factors (e.g. hypertension, obesity, male sex, age, etc.) in order to cause a cardiovascular event.

5. LIMITATIONS OF THE STUDY

The mayor limitation of the study is the small sample size. Indeed, further larger studies are required to confirm these results. Moreover, the lack of a follow-up does not allow to get informations about the prognostic implication of these polymorphisms.

6. CONCLUSION

Resulting data suggest that EDN-1 Glu105Glu (rs5369) polymorphism could be involved in the risk of developing ACS in Mexican in patients. Risk variants should provide pathophysiological insights and targets for novel therapy.

7. REFERENCES

1. Garcia-Moll, Xavier, 2005. Inflammatory and anti-Inflammatory markers in acute coronary syndromes. Ready for use in the clinical setting? *Rev. Esp. Cardiol.* 58, 615–617
2. Crea F, Liuzzo G. Pathogenesis of acute coronary syndromes. *J Am Coll Cardiol.* 2013;61:1-11.
3. Cheruvu PK, Finn AV, Gardner C, Caplan J, Goldstein J, Stone GW, Virmani R, Muller JE. Frequency and distribution of thin-cap fibroatheroma and ruptured plaques in human coronary arteries: a pathologic study. *J Am Coll Cardiol* 2007;50:940-949.
4. Virmani R, Kolodgie FD, Burke AP, Farb A, Schwartz SM. Lessons from sudden coronary death: a comprehensive morphological classification scheme for atherosclerotic lesions. *Arterioscler Thromb Vasc Biol* 2000;20:1262–1275-
5. Arbustini E, Dal Bello B, Morbini P, Burke AP, Bocciarelli M, Specchia G, Virmani R. Plaque erosion is a major substrate for coronary thrombosis in acute myocardial infarction. *Heart* 1999;82:269–272.
6. Niccoli G, Montone RA, Di Vito L, Gramegna M, Refaat H, Scalone G, Leone AM, Trani C, Burzotta F, Porto I, Aurigemma C, Prati F, Crea F. Plaque rupture and intact fibrous cap assessed by optical coherence tomography portend different outcomes in patients with acute coronary syndrome. *Eur Heart J.* 2015;36:1377-84.
7. Moreno PR, Falk E, Palacios IF, Newell JB, Fuster V, Fallon JT. Macrophage infiltration in acute coronary syndromes. Implications for plaque rupture. *Circulation* 1994; 90:775–778
8. Cristell N, Cianflone D, Durante A, Ammirati E, Vanuzzo D, Banfi M, et al; FAMI Study Investigators. High-sensitivity C-reactive protein is within normal levels at the very onset of first ST-segment elevation acute myocardial infarction in 41% of cases: a multiethnic case-control study. *J Am Coll Cardiol* 2011; 58:2654-61.

9. Liuzzo G, Biasucci LM, Gallimore JR, Grillo RL, Rebuffi AG, Pepys MB, et al. The prognostic value of C-reactive protein and serum amyloid a protein in severe unstable angina. *N Engl J Med* 1994; 331:417-24.
10. Topol, E.J., Smith, J., Plow, E.F., Wang, Q.K., 2006. Genetic susceptibility to myocardial infarction and coronary artery diseases. *Hum. Mol. Genet.* 15, R117–R123.
11. Rankinen, T., et al., 2007. Effect of endothelin 1 genotype on blood pressure is dependent on physical activity or fitness levels. *Hypertension* 50, 1120–1125.
12. Hickey, K.A., Rubanyi, G., Paul, R.J., Highsmith, R.F., 1985. Characterization of a coronary vasoconstrictor produced by cultured endothelial cells. *Am. J. Physiol.* 248, C550–C556.
13. Brunner, F., Bras-Silva, C., Cerdeira, A.S., Leite-Moreira, A.F., 2006. Cardiovascular endothelins: essential regulators of cardiovascular homeostasis. *Pharmacol. Ther.* 111, 508–531.
14. Barton, M., Huadenschild, C.C., d'Uscio, L.V., Shaw, S., Munter, K., Luscher, T.F., 1998. Endothelin ETA receptor blockade restores NO-mediated endothelial function and inhibits atherosclerosis in apolipoprotein E-deficient mice. *Proc. Natl. Acad. Sci. U. S. A.* 95, 14367–14372.
15. Best, P.J.M., Lerma, A., 1998. Endothelin in cardiovascular disease: from atherosclerosis to heart failure. *J. Cardiovasc. Pharmacol.* 31, S61–S63.
16. Ivey, M.E., Osman, N., Little, P.J., 2008. Endothelin 1 signalling in vascular smooth muscle: pathways controlling cellular functions associated with atherosclerosis. *Atherosclerosis* 199, 237–247.
17. Bohm, F., Pernow, J., 2007. The importance of endothelin-1 for vascular dysfunction in cardiovascular disease. *Cardiovasc. Res.* 76, 8–18.
18. Simionescu, M., 2007. Implications of early structural–functional changes in the endothelium for vascular disease. *Arterioscler. Thromb. Vasc. Biol.* 27, 266–274.
19. Yang, Z., Xiu-Fen, M., 2006. Recent advances in understanding endothelial dysfunction in atherosclerosis. *Clin. Med. Res.* 4, 53–65.
20. Yang, Z., Luscher, T.F., 2002. Vascular endothelium. In: Lanzer, P., Topol, E.J. (Eds.), *Panvascular medicine*. Springer, Berlin - Heidelberg – New York, pp. 190–204.
21. Rabelink, T.J., Lusher, T.F., 2006. Endothelial nitric oxide synthase. Host defense enzyme of the endothelium? *Arterioscler. Thromb. Vasc. Biol.* 26, 267–271.

22. Duerschmidt, N., Wippich, N., Goettsch, W., Broemme, H.J., Morawietz, H., 2000. Endothelin-1 induces NAD(P)H oxidase in human endothelial cells. *Biochem. Biophys. Res. Commun.* 269, 713–717.
23. Fukuchi, M., Giaid, A., 1999. Endothelial expression of endothelial nitric oxide synthase and endothelin-1 in human coronary artery disease. Specific reference to underlying lesion. *Lab. Invest.* 79, 659–670.
24. Wilcox, J.N., et al., 1997. Expression of multiple isoforms of nitric oxide synthase in normal and atherosclerotic vessels. *Arterioscler. Thromb. Vasc. Biol.* 17, 2479–2488.
25. 1000 Genomes Project Consortium, Abecasis GR, Altshuler D, et al. A map of human genome variation from population-scale sequencing. *Nature* 2010; 467:1061.
26. Chakravarti A. It's raining SNPs, hallelujah? *Nat Genet* 1998; 19:216.
27. Li WH, Sadler LA. Low nucleotide diversity in man. *Genetics* 1991; 129:513.
28. Castro, M.G., et al., 2007. Screening of the endothelin 1 gene (END1) in a cohort of patients with essential left ventricular hypertrophy. *Ann. Hum. Genet.* 71, 601–610.
29. Palacin, M., et al., 2009. Lack of association between endothelin-1 gene variants myocardial infarction. *J. Atheroscler. Thromb.* 16, 388–395.
30. Zhu, G., et al., 2008. Polymorphisms in the endothelin-1 (EDN1) are associated with asthma in two populations. *Genes Immun.* 9, 23–29.
31. Tobe, S.W., et al., 2011. The impact of endothelin-1 genetic analysis and job strain on ambulatory blood pressure. *J. Psychosom. Res.* 71, 97–101.
32. Colombo, M.G., et al., 2003. Endothelial nitric synthase gene polymorphisms and risk of coronary artery disease. *Clin. Chem.* 49, 389–395.
33. Kusmanic-Samija, R., et al., 2011. Association of NOS3 tag polymorphisms with hypoxicischemic encephalopathy. *Croat. Med. J.* 52, 396–402.
34. McKnight, A.J., et al., 2010. Genetic polymorphisms in nitric oxide synthase 3 gene and implications for kidney diseases: a meta-analysis. *Am. J. Nephrol.* 32, 476–481.
35. Prescluttini, S., et al., 2009. Promoter polymorphisms of the NOS3 gene are associated with hypnotizability-dependent vascular response to nociceptive stimulation. *Neurosci. Lett.* 467, 252–255.

36. Shentil, D., et al., 2005. Genotype-dependent expression of endothelial nitric oxide synthase (eNOS) and its regulatory proteins in cultured endothelial cells. *DNA Cell Biol.* 24, 218–224.

37. Richardson, P., et al., 1996. Report of the 1995 World Health Organization/International Society and Federation of Cardiology Task force on the definition and classification of cardiomyopathies. *Circulation* 93, 841–842

8. TABLES AND FIGURES

Table 1 Clinical characteristics of study groups

	Case (n 218)	Control (n 204)	p
Age, yrs, mean±SD	60.39±11.07	58.9±9.85	<0.001
Male , n (%)	148 (67)	83 (40)	<0.001
Smoking, n (%)	130 (59)	27 (13)	<0.001
Diabetes Mellitus, n (%)	81 (37)	26 (12)	<0.001
Hypertension, n (%)	131 (60)	85 (41)	<0.001
Dislipidemia, n (%)	133 (61)	111 (54)	0.102
Family history of CAD, n (%)	118 (25)	14 (6)	<0.001
Obesity, n (%)	47 (21)	33 (16)	0.277
CV Risk grade, n(%)			<0.001
Mild	38 (17)	86 ()	
Moderate	56 (25)	61 (30)	
high	124 (56)	57 (28)	
TC, mg/dl, median (IR)	169.0 (138.0- 197.0)	175.0 (150-195)	0.220
HDL, mg/dl, median (IR)	40.0 (34.0-47.65)	41.8 (34.0-48.0)	0.360
LDL, mg/dl, median (IR)	93.0 (72.0-117.0)	98.0 (75.2-118.5)	0.269
TG, mg/dl, median (IR)	144.5 (108.6- 204.5)	140.5 (104.0-201.5)	0.441

Coronary artery disease, CAD; CV, cardiovascular risk factors; interquartile range, IR; standard deviation, SD; total cholesterol, TC; LDL, HD

Table 2 Genetic profile of study groups

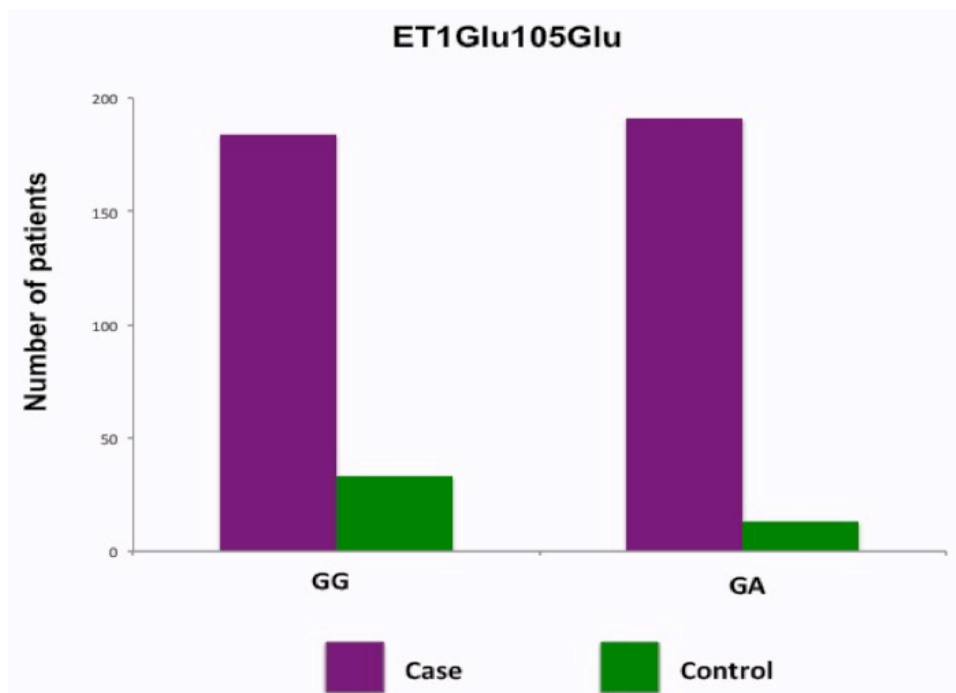
	Case (n 218)	Control (n 204)	p
eNOS2980, n (%)			0.441
GG	138	140	
GT	66	55	
TT	14	9	
eNOS785, n (%)			0.121
TT	123	135	
TC	81	59	
CC	14	10	
eNOS1474, n (%)			0.058
TT	119	130	
TA	88	60	
AA	11	14	
ET1974, n (%)			0.551
AC	41	34	
AA	176	170	
ET1394, n (%)			0.432
TT	171	170	
TG	46	33	
GG	1	1	
ET1Glu105Glu, n (%)			0.004
GG	184	191	
GA	33	13	

Table 3 Independent predictors of ACS.

	Univariable regression model		Multivariable regression model	
	HR (CI 95%)	p	OR (CI 95%)	p
Age	0.010 (0.005-0.014)	<0.001	0.127.(-0.001-0.008)	0.127
Male	0.274 (0.182-0.367)	<0.001	0.171 (0.086-0.257)	<0.001
Smoking	-0.065 (-0.132-0.001)	0.055		
Hypertension	0.018 (0.085-0.275)	<0.001	0.080 (-0.002-0.163)	0.055
Diabetes Mellitus	0.322 (0.216-0.428)	0.054		
Family history of CAD	0.549 (0.460-0.638)	<0.001	0.459 (0.410-0.580)	<0.001
CV risk grade	0.190 (0.136-0.244)	<0.001	0.121 (0.011-0.132)	0.020
Dislipidemia	0.068.(-0.029-0.164)	0.171		
ET1Glu105Glu	0.227.(0.074-0.379)	0.004	0.129 (0.082-0.329)	0.001

Coronary artery disease, CAD; confidence of interval, CI; CV, cardiovascular risk factors; Hazard ratio, HR. Odds ratio, OR.

Figure 1



9. LEGENDS

Figure 1.

Distribution of ETGlu105Glu polymorphism in case and control groups.