



**UNIVERSIDAD NACIONAL AUTÓNOMA DE MÉXICO**

**POSGRADO EN CIENCIAS BIOLÓGICAS**

**INSTITUTO DE ECOLOGÍA**

**BIOLOGÍA EVOLUTIVA**

**Caracterización evolutiva de *Pinus pinceana* (Gordon & Glend.)  
a escalas filogenética, poblacional y funcional con  
aproximaciones genómicas**

**TESIS**

**QUE PARA OPTAR POR EL GRADO DE:**

**DOCTORA EN CIENCIAS**

**PRESENTA:**

**LAURA ALICIA FIGUEROA CORONA**

**TUTOR PRINCIPAL DE TESIS: DR. DANIEL PIÑERO DALMAU**

**INSTITUTO DE ECOLOGÍA, UNAM**

**COMITÉ TUTOR: DR. LUIS E. EGUIARTE FRUNS**

**INSTITUTO DE ECOLOGÍA, UNAM**

**DR. DAVID SEBASTIAN GERNANDT**

**INSTITUTO DE BIOLOGÍA, UNAM**

**Ciudad Universitaria, CD. MX.**

**Enero, 2022.**



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**COORDINACIÓN DEL POSGRADO EN CIENCIAS BIOLÓGICAS**

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**M. en C. Ivonne Ramírez Wence**  
**Directora General de Administración Escolar, UNAM**  
**Presente**

Me permito informar a usted, que el Subcomité de Biología Evolutiva y Ecología, del Posgrado en Ciencias Biológicas, en su **reunión ordinaria del día 16 de agosto de 2021** se aprobó el siguiente jurado para el examen de grado de **DOCTORA EN CIENCIAS** de la estudiante **FIGUEROA CORONA LAURA ALICIA**, con número de cuenta **304289071**, con la tesis titulada: **“CARACTERIZACIÓN EVOLUTIVA DE PINUS PINCEANA (GORDON & GLEND.) A ESCALAS FILOGENÉTICA, POBLACIONAL Y FUNCIONAL CON APROXIMACIONES GENÓMICAS**, Tutor Principal, quedando integrado de la siguiente manera:

Presidente: DRA. ANA ELENA ESCALANTE HERNÁNDEZ  
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Secretario: DR. LUIS ENRIQUE EGUIARTE FRUNS

Sin otro particular, me es grato enviarle un cordial saludo.

**ATENTAMENTE**  
**“POR MI RAZA HABLARÁ EL ESPÍRITU”**  
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**DR. ADOLFO GERARDO NAVARRO SIGÜENZA**



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## Abreviaciones

ABA	Ácido abscísico
bp	Base pair
CDS	Coding DNA sequence
ChD	Desierto Chihuahuense
cpDNA	Chloroplast DNA
Gbp	Gigabase pairs; mil millones de nucleótidos
ILS	Incomplete lineage sorting
IUCN	International Union for Conservation of Nature
IR	Repetición invertida
JA	Ácido jasmónico
kya	Thousand years ago
LIG	Last Interglacial
LRR	Leucine-rich repeat
LTR-RT	Long terminal repeat (LTR) retrotransposons
LSC	Large single-copy region
Ma	Millón de años
mRNA	<i>Messenger RNA</i>
ROS	Reactive oxygen species
rRNA	ribosomal RNA
SA	Ácido salicílico
SMO	Sierra Madre Oriental
SNP	Single nucleotide polymorphisms
SSC	Small single-copy region
SSRs	Microsatellites o Single Sequence Repeats
tRNA	transfer RNA

## Resumen

La respuesta a la aridez constituye un rasgo poligénico que es resultado de interacciones del genotipo - fenotipo con el ambiente; además genera mecanismos ecofisiológicos, y genéticos que están interrelacionados, y que son codependientes y complementarios, para producir una respuesta plástica y/o adaptativa. En este trabajo busco identificar los cambios en la respuesta transcripcional a la aridez en *Pinus pinceana* (Gordon & Glend.) y el efecto de las fuerzas evolutivas que han modificado su arquitectura genómica. En el Primer Capítulo de esta tesis describo los estudios de los niveles de expresión del mRNA, como aproximación de la respuesta fisiológica a los factores bióticos y abióticos en su ambiente. Para esto analicé el transcriptoma, buscando resolver la relación entre la divergencia ambiental y las diferencias en los perfiles de expresión. El estudio del transcriptoma con un ensamble *de novo* caracterizó 45,431 anotaciones del tejido diploide de acículas de siete árboles maduros, y el análisis de expresión diferencial identificó reguladores fitohormonales de ABA y JA y la regulación de ROS posiblemente involucrados en la respuesta al estrés abiótico. Los perfiles de expresión genética resaltaron la expresión diferencial de 26 CDS involucrados en la regulación de respuesta al estrés abiótico, y proteínas con receptores LRR relacionadas con el reconocimiento de patógenos. En el Segundo Capítulo analizo la diversidad genética en la especie, así como su historia demográfica y patrón filogeográfico. Encontré dos grupos genéticos generados por la dinámica interglacial durante los cambios climáticos del Pleistoceno (~627 kya), seguidos de la subfragmentación de ambas regiones (~127.7 - ~539.2 kya), el Desierto Chihuahuense entre su región este y oeste, y la Sierra Madre Oriental en su porción sur y norte, mientras la aridificación aumentaba en el Desierto Chihuahuense. En el Tercer Capítulo estudio los patrones de divergencia dentro y entre los dos clústers genéticos y sus especies hermanas, reconstruyendo las relaciones filogenéticas y la divergencia de los pinos piñoneros de conos grandes a partir de datos del genoma plastídico y la concatenación de SNPs de genes nucleares altamente conservados. Los resultados apoyan la estructuración de *P. pinceana* en dos grupos, descartando para cualquiera de las especies del clado eventos de introgresión o eventos de migración con *P. maximartinezii*.

## Abstract

The response to aridity constitutes a polygenic trait that is the result of genotype-phenotype interactions with the environment; it also generates ecophysiological and genetic mechanisms that are interrelated, and that are codependent and complementary, to produce a plastic and/or adaptative response. In this work, I seek to identify the changes in the transcriptional response to aridity in *Pinus pinceana* (Gordon & Glend.) And the effect of evolutionary forces that have modified its genomic architecture. In the First Chapter of this thesis, I describe the studies of mRNA transcription, as an approximation of physiological response to biotic and abiotic factors in their environment. For this, I analyzed the transcriptome, seeking to resolve the relationship between environmental divergence and differences in expression profiles. The study of the transcriptome with a *de novo* assembly characterized 45,431 annotations of diploid tissue of needles from seven mature trees, and the differential expression analysis identified phytohormonal regulators of ABA and JA and the regulation of ROS possibly involved in the response to abiotic stress. The gene expression profiles highlighted the differential expression of 26 CDS involved in the regulation of the response to abiotic stress and proteins with LRR receptors related to the recognition of pathogens. In the Second Chapter, I analyze the genetic diversity in the species, as well as its demographic history and phylogeographic pattern. I found two genetic groups generated by interglacial dynamics during the Pleistocene climate changes (~627 kya), followed by the subfragmentation of both regions (~ 127.7 - ~ 539.2 kya), the Chihuahuan Desert between its eastern and western regions, and the Sierra Madre Oriental in its southern and northern portions, while aridification increased in the Chihuahuan Desert. In the Third Chapter, I study the patterns of divergence within and between the two genetic clusters and their sister species, reconstructing the phylogenetic relationships and divergence of large cone stone pines from plastidic genome data and the concatenation of nuclear gene SNPs highly preserved. The results support the structuring of *P. pinceana* into two groups, ruling out introgression events or migration events with *P. maximartinezii* for any of the species of the clade.

## **Introducción**

### **La adaptación, la plasticidad fenotípica y la historia demográfica, y su efecto en la evolución**

La respuesta a factores abióticos y bióticos influye en el proceso evolutivo y repercute en los patrones de variación genómica (Hand et al., 2015). El reconocimiento de las rutas de respuesta a estos factores es importante porque permite entender la tolerancia al estrés, la adaptación y la interacción entre el genotipo, el fenotipo y el ambiente. Los mecanismos evolutivos afectan la distribución de la diversidad genética de las poblaciones, y para detectar estos mecanismos se analizan las desviaciones en los modelos neutros (Leigh, 2007). Los estudios de asociación genotipo - fenotipo - ambiente buscan ubicar los caracteres genéticos que determinan la varianza fenotípica y establecen correlaciones con la divergencia del ambiente. Un ejemplo de estos estudios es el análisis de patrones de expresión, que bajo determinadas condiciones, edades o tejidos, en un panorama comparativo y multilocus, describen la respuesta fisiológica (Nosil et al., 2009).

#### **1. La adaptación y la plasticidad en la respuesta a la aridez**

Evolutivamente, la regulación de la humedad representa una estrategia clave para el establecimiento de las plantas en ambientes terrestres. La aridez, entendida como el déficit de recursos hídricos en el suelo y la falta de humedad en el aire (Pompa-García, 2017), representa una de las presiones más agresivas para el crecimiento y desarrollo de los organismos sésiles (Le & McQueen-Mason, 2006). Ante fluctuaciones en las condiciones hídricas, los organismos tienen tres escenarios posibles: 1) evadir la exposición a los factores y condiciones estresantes; 2) reducir los niveles de deshidratación con latencia o de reducción de biomasa; o 3) tolerar los cambios ambientales implementando mecanismos de resistencia que contrarresten los daños (Farooq et al., 2012).

Estos escenarios de respuesta al estrés abiótico están interrelacionados, son co-dependientes y complementarios, y es evolutivamente posible que generen respuestas adaptativas de un organismo para establecerse en un ambiente árido. Estas respuestas se clasifican en respuestas proximales, donde la respuesta fisiológica es plástica; son importantes en ambientes heterogéneos pues confieren varianza fenotípica que puede

proporcionar una mayor tolerancia ambiental (Via et al., 1995); y respuestas evolutivas, producidas por cambios y polimorfismos genéticos. Los cambios plásticos en el fenotipo pueden ser adaptativos o no; pero, si existe variabilidad genética apropiada, las respuestas plásticas son potencialmente adaptativas (Via et al., 1995, Schlichting & Levin, 1986).

Los efectos de la aridez repercuten en todas las respuestas fisiológicas y metabólicas de un organismo (Zhu et al., 2002). Hernández et al. (2012) han descrito una conservación de estrategias comunes a la respuesta al congelamiento, la aridez y la salinidad en los linajes de plantas. Los efectos netos en la respuesta a la aridez ocurren a nivel morfológico, fisiológico y molecular, e involucran tres puntos: 1) optimizar el consumo y almacenamiento de agua; 2) reducir la pérdida de agua; y, 3) prevenir el daño celular (De Micco et al., 2012; Huang et al., 2012).

Los cambios morfológicos y de desarrollo involucrados en la tolerancia a la desecación y la regulación del balance del contenido de agua se presentan en todas las plantas terrestres. Estos cambios pueden ser la evolución de la cobertura de esporopolenina en polen y esporas, un politerpeno impermeable, o la aparición de la cutícula; y fisiológicamente a una escala temporal relativamente larga en la vida de una planta ocurren cambios en la arquitectura de la raíz y depósitos de cera en hojas, mientras que a corto plazo tenemos procesos como el cierre de los estomas y el marchitamiento (Vilagrosa et al., 2012; Le & McQueen-Mason, 2006; De Micco et al., 2012). Los mecanismos fisiológicos comprenden las vías de regulación y abastecimiento de factores fitohormonales, osmoprotectores y la regulación de agentes oxidantes (McDowell et al., 2008; Moran et al., 2017).

Tardieu et al. (1998) clasifican la regulación de la evapotranspiración en dos tipos: 1) la regulación isohídrica, en la que se produce el cierre de los estomas implicando la vía de señalización de ácido abscísico (ABA), se presenta en organismos con traqueidas gruesas en coníferas; este tipo de regulación se presenta en la familia Pinaceae; 2) la regulación anisohídrica, que permite la pérdida de recursos hídricos y genera cambios del potencial hídrico, como señal para el cierre de estomas. Este último tipo de regulación es característica de las familias Cupressaceae y Taxaceae, y se ha descrito que el tejido vascular de organismos de regulación anisohídrica tiende a ser más resistente y con menor tendencia a la cavitación (McDowell et al., 2008; Brodribb et al., 2014).

Las estrategias isohídricas son frecuentemente asociadas a plantas con mayor sensibilidad a déficits hídricos. En las coníferas, se presenta la combinación de estrategias para el transporte de agua, y la eficacia de su regulación ha sido asociada al ambiente y la altura del tronco (Brodribb et al., 2014). Hochberg et al. (2017) han señalado que la clasificación de los mecanismos de regulación hídricos no es tajante y es más un continuo, pues 1) la misma especie o especies dentro de la misma familia pueden presentar diferencias en su control hídrico (*p.e. Vitis vinifera*; Cupressaceae y Callitroideae; *Populus alba*; Hochberg et al., 2017; Pittermann et al., 2012; Zhang et al., 2020), y 2) al ser una respuesta gradual, los mecanismos no son excluyentes.

La relevancia de la regulación por ABA ha formado parte de los argumentos para explicar la conquista del medio terrestre de las plantas, ya que su participación en la regulación del estrés hídrico y como promotor del desarrollo ha sido documentado en los clados más basales de plantas (Le & McQueen-Mason, 2006; Sun et al., 2020). Pittermann et al. (2012) describen basados en paleoclimas que la diversificación dentro de Cupressaceae partió de ancestros intolerantes a la sequía en hábitats húmedos durante el Mesozoico, y que los cambios climáticos durante los últimos 150 Ma, especialmente el aumento de la aridez desde el Oligoceno, han producido modificaciones en la regulación hídrica, variaciones estructurales del xilema, una menor eficiencia de transporte del xilema y, a nivel de las hojas, una menor capacidad fotosintética entre Cupressaceae y Callitroideae.

### 1.1 Regulación fitohormonal

Las fitohormonas son los reguladores fundamentales del desarrollo, el crecimiento, la diferenciación y la senescencia de las plantas (Davies, 2010); son necesarias en concentraciones mínimas para dirigir vías de señalización (Farooq et al., 2009a). Las fitohormonas que están involucradas directamente en la respuesta al déficit hídrico son el ácido abscísico (ABA) y el ácido jasmónico (JA); sin embargo, el aumento de estos dos compuestos desencadena la disminución de auxinas, giberelinas, y citoquininas, principales promotores del crecimiento de plantas (Taiz & Zeiger, 2010).

El ABA promueve la latencia del embrión, genera la regulación de la respuesta al estrés osmótico provocado por la sequía, el aumento de salinidad y el congelamiento. Inicialmente induce el cierre de estomas mediante el cambio de turgencia en las células

guardas, activando los canales de transporte de  $\text{Ca}^{2+}$  y de  $\text{K}^{+}$  (Corcuera et al., 2012a, b). Sin embargo, se estima que en cambios ambientales repentinos y a medida que la altura de los organismos aumenta la señalización de regulación estomática por ABA se vuelve ineficiente (Moran et al., 2017).

El JA participa en los procesos de maduración de frutos en angiospermas, el desarrollo del polen y el crecimiento de la raíz. Además, actúa como regulador celular de la respuesta de defensa al daño por patógenos, por exposición a ozono y durante el déficit hídrico (Creelman & Mullet, 1995). Durante periodos de estrés, el JA promueve la síntesis de fosfatasa e inhibidores de proteinasas, y enzimas involucradas en la biosíntesis de flavonoides como la chalcona sintasa y la biosíntesis de terpenoides. Estos factores generan la reducción de agentes oxidantes y favorecen la estabilización de la membrana (Creelman & Mullet, 1995).

## 1.2 Regulación del estrés oxidativo

Debido al suministro limitado de agua y el cierre de estomas, el estrés oxidativo aumenta (Kiani et al., 2007) y produce una disminución de  $\text{CO}_2$ , que a su vez genera alteraciones en la actividad enzimática y conduce a la producción y acumulación de especies reactivas de oxígeno (ROS) como  $^1\text{O}_2$ ,  $\text{O}_2^-$ ,  $\text{H}_2\text{O}_2$ ,  $\text{OH}^-$ ,  $\text{RO}^*$ ,  $\text{ROO}^*$  y  $\text{ROOH}^*$  (Flexas-Medrano, 2002; Lawlor & Cornic, 2002). Las propiedades oxidantes de las ROS producen daño a lípidos, proteínas y otras macromoléculas (Rout & Shaw, 2001). Al aumentar su concentración, se activan dos estrategias: una involucra la reducción de ROS y la segunda promueve la protección de enzimas, DNA y membranas.

Los intermediarios osmoreguladores y osmoprotectores más importantes son salicilatos y flavonoides (Scandalios et al., 2005; Li et al., 2008; Ozkur et al., 2009). Los flavonoides son compuestos polifenólicos, solubles en agua, como el  $\alpha$ -tocoferol, el glutatión, y los  $\beta$ -carotenos, que están involucrados en la filtración UV, y en la fijación simbiótica de nitrógeno. Los flavonoides actúan también como reguladores del ciclo celular, y adicionalmente en la raíz facilitan la formación de nódulos fijadores de Nitrógeno e interacciones micorrícicas.

### 1.3 Producción de carbohidratos osmoprotectores

Cuando hay déficit de agua, la tasa fotosintética disminuye, se dispara el uso de las reservas de carbono al reducir el almidón en sacarosa y tetralosa, metabolitos que contribuyen al mantenimiento del turgor y la prevención de la desnaturalización de proteínas y la fusión de membranas (Le & McQueen-Mason, 2006). Estos osmoprotectores están relacionados con la protección y reparación del DNA, la estabilización y la reconstrucción de la membrana. Cambios abruptos en salinidad disparan la formación residual de alcoholes polihídricos como manitol, sorbitol y pinitol que regulan la osmolaridad, por lo que se reduce el flujo extracelular y se favorece la retención de agua (Bacelar et al., 2012). Al mismo tiempo que la producción de prolina y compuestos monoméricos de glicina (betaína o trimetilglicina) y las cetonas (chalconas) es promovida por el incremento de aridez, salinidad y temperatura, se logra la estabilización de la estructura cuaternaria de enzimas, complejos proteicos, y la membrana del tilacoide (Xing-Rajashekar, 1999).

## 2. Enfoques transcripcionales aplicados al estudio de la respuesta a la aridez

Actualmente, la respuesta al déficit hídrico se puede estudiar a partir de análisis ecofisiológicos, que conjuntan estudios genómicos, mapeos genéticos (Mackay & Dean, 2011) y la descripción de perfiles transcripcionales que han facilitado el entendimiento de la interacción genotipo-fenotipo con el ambiente, además de favorecer la identificación de genes responsables de la respuesta a la desecación (Cañas et al., 2015; Kramer & Boyer, 2015).

El ensamble de un transcriptoma constituye la medición y caracterización de la actividad transcripcional en una condición determinada en un momento específico (Brown, 2002). Ayuda a la comprensión de las redes de regulación en las estrategias de resistencia y susceptibilidad al estrés abiótico (Mackay et al., 2012). En esta tesis en el Primer Capítulo describo la respuesta al estrés biótico y abiótico en *Pinus pinceana* Gordon & Glend. en su ambiente natural, el cual es heterogéneo en cuanto a la precipitación y humedad.

Estos enfoques genómicos han hecho posible distinguir las respuestas plásticas de los mecanismos evolutivos adaptativos. Para que exista adaptación en términos evolutivos, es necesario que haya selección y que existan bases genéticas; en otras palabras, que sea



heredable. Este proceso selectivo implica cambios y frecuencias alélicas en genes que afectan el fenotipo, relacionado con diferencias en la adecuación de los individuos entre los distintos hábitats. Estas diferencias adaptativas pueden estar dadas por (o relacionadas con) distintas tasas de crecimiento, capacidades reproductivas y de supervivencia (Kuparinen et al., 2010). Las variaciones adaptativas pueden aparecer a diferentes escalas, desde una nueva morfología hasta un cambio en los niveles de expresión (Romero et al., 2012). Los estudios que abordan la adaptación local de las poblaciones forestales cubren escalas espaciales amplias entre ambientes heterogéneos y evalúan la varianza genética en rasgos cuantitativos y poligénicos (Morgenstern, 1996; Eckert et al., 2010).

Sin embargo, es importante señalar que los efectos de la selección natural dependen de la arquitectura genética del rasgo fenotípico, su relación con la adecuación, la historia demográfica de las poblaciones, el tipo y fuerza de selección, además del momento del inicio de la selección (Linnen & Hoekstra, 2009). Sin desestimar la detección de genes candidatos bajo selección natural, es importante considerar que el efecto y las interacciones de otras fuerzas evolutivas como la migración y la historia demográfica modifican la diversidad genética de manera similar (Mitton et al., 1989).

Los métodos que buscan identificar adaptación en poblaciones detectan valores atípicos en la frecuencia de alelos entre las poblaciones, comparando la diferenciación poblacional o buscan estructuras de varianza-covarianza entre poblaciones con los factores ambientales (Nielsen, 2005; Gunther & Coop, 2013; Rellstab et al., 2015).

### **3. Los efectos de la historia demográfica en la diversidad genética**

Al analizar la variación genética, es difícil distinguir los efectos selectivos de los efectos histórico-demográficos, debido a que los cambios en el tamaño de la población dejan huella en la distribución de frecuencias de la diversidad genética (Johri et al., 2021). Considerando la variación neutral en un solo locus, las predicciones del modelo de un cuello de botella, y los efectos de un barrido selectivo generan el mismo patrón de variación genética (Tajima 1989; Depaulis et al., 2003, 2009). Por lo tanto, es importante reconocer que, para distinguir estos procesos, se deben analizar polimorfismos en varios loci, ya que los eventos demográficos influyen en todo el genoma. En contraste, la selección afecta solo a regiones específicas del genoma (Rellstab et al., 2015).

Para la identificación del efecto de la demografía histórica en la distribución de las frecuencias alélicas y la diversidad genética se utilizan estadísticos descriptivos como la  $D$  de Tajima o la  $H$  de Fay y Wu y el espectro de frecuencias de alelos (Tajima, 1989; Fay & Wu, 2000; Przeworki, 2002; Depaulis et al., 2003). Los valores negativos de  $D$  indican poblaciones con expansiones recientes, efectos de selección purificadora o un barrido selectivo. Estos valores negativos de  $D$  indican una distribución de las frecuencias de los alelos que difiere de la expectativa neutral con un exceso de alelos raros o derivados. Valores de  $D$  positivos evidencian que las poblaciones han pasado por selección balanceadora o cuellos de botella, y han reducido sus niveles de diversidad (Nei et al., 1975, Nielsen & Slatkin, 2013).

En el Segundo Capítulo de esta tesis analizo la demografía histórica de *P. pinceana* y la distribución geográfica de la diversidad genética en un contexto geológico, y de los cambios climáticos que han ocurrido en el Desierto Chihuahuense en el Pleistoceno utilizando polimorfismos del genoma completo.

#### 4. Enfoques genéticos y genómicos a la ecología evolutiva en pinos

Con centro de origen en Asia, el género *Pinus* está actualmente distribuido en el hemisferio norte (Saladin et al., 2017; Wei-Tao et al., 2021). Las especies extendieron el rango de distribución a partir de la migración a través de los sistemas montañosos y se adaptaron a diferentes gradientes de elevación (Perry, 1991; Richardson, 1998; Farjon & Styles, 2010). En México, con una orografía y amplia heterogeneidad ambiental, el género *Pinus* presenta una gran diversidad; de hecho, es el país con mayor diversidad de pinos del mundo, donde se han descrito entre 47 y 51 de las ca. de 111 especies existentes (Richardson et al., 1998; Perry, 1991; Perry et al., 2000).

En *Pinus* se han reportado genomas extremadamente grandes y complejos, que varían de 16 a 35 Gbp (De la Torre et al., 2014; Leitch et al., 2012). A pesar de los costos decrecientes de la secuenciación, la resecuenciación del genoma completo en múltiples individuos aún no es factible. Actualmente, existen dos genomas de referencia para el género: uno de *P. taeda* (22.1 Gbp, con 50,172 genes que codifican a proteínas) y otro de *P. lambertiana* (30 Gbp, 38,518 CDS; Wegrzyn et al., 2014; Zimin et al., 2014; Scott et al., 2020). De la Torre et al., (2014) atribuyen el gran tamaño del genoma en *Pinus* a la

acumulación masiva y eliminación deficiente de elementos transponibles (retrotransposones) con secuencias terminales de repetidos largos (LTR-RT). Los elementos transponibles constituyen el 74% de los genomas de *P. taeda*, y el 79% en *P. lambertiana* (Neale et al., 2014; et al., 2017; Wan et al., 2018; Wegrzyn et al., 2014).

Para el genoma plastídico en las coníferas se hereda paternalmente y, por lo tanto, rastrea la historia evolutiva de la dispersión del polen, independiente del genoma nuclear, y con frecuencia es independiente del genoma mitocondrial (Neale & Sederoff, 1989; Whittall et al., 2010). Se puede obtener fácilmente una historia genealógica única e independiente para la comparación con el genoma nuclear que separen las contribuciones parentales (polen vs semilla). Los plastomas de coníferas también se caracterizan por extensos reordenamientos genómicos y estudios recientes describen cambios principalmente en las dos repeticiones invertidas (IR) de 20 a 30 Kb (IRA e IRB), que generalmente contienen cuatro RNA ribosomales que en Pinaceae aparecen con reordenamientos y deleciones en la región IRB, además de la recombinación de IR cortos (Wu et al., 2011a,b; Hsu et al., 2016).

Tras analizar el genoma de las coníferas, Chen et al. (2013) describen que existe una tendencia por conservar los genes del metabolismo general sin duplicaciones, mientras que las secuencias de proteínas involucradas en el metabolismo especializado y secundario tienden a ser redundantes. Además, se ha reportado un número elevado de genes que codifican para las cinasas con repeticiones ricas en leucina, citocromo P450, y los factores de transcripción MYB (De la Torre et al., 2015; Neale et al., 2014; Pavy et al., 2013; Porth et al., 2011; Warren et al., 2015). Debido al gran tamaño de sus genomas, estrategias de secuenciación de RNA (RNA-seq) y RAD-seq, que se basan en un muestreo fragmentado del genoma representan las mejores alternativas para el genotipado de miles de loci (Schlötterer et al., 2014; Hoban et al., 2016; Jones et al., 2016; Oney-Birol et al., 2018).

La regulación genética de la respuesta a la aridez en los pinos ha sido poco explorada, y, aunque se han propuesto algunas etapas de las vías de señalización de la respuesta al estrés abiótico, para ninguna de ellas se elucidaron intermediarios o sustratos y productos (Balderas-Hernández et al., 2013). En general, el análisis de perfiles transcripcionales en plantas bajo condiciones de estrés sugiere la conservación de respuestas a la aridez, el congelamiento y la salinidad entre angiospermas y gimnospermas (Díaz-Sala, 2013; Moran et al., 2017). Los diseños experimentales que se han explotado con mayor

detalle para reconocer la respuesta al déficit hídrico se han elaborado en plántulas de *P. taeda* y *P. pinaster* (Lorenz, 2006; Watkinson et al., 2003; Paiva et al., 2008) con el objetivo de reconocer la regulación de las vías de respuesta a la humedad, los patrones de crecimiento y la modificación en los procesos de lignificación.

A nivel genético, los estudios adaptativos en los pinos han reportado niveles altos de  $d_N / d_S$  (proporción de sustituciones no sinónimas y sinónimas) en comparación con angiospermas modelo. Esto ha sido interpretado como el efecto de presiones de selección fuertes, probablemente debido a tamaños de población efectivos grandes (De la Torre et al., 2017), mientras que la diferenciación poblacional es alta debido a los niveles de diversidad genética dentro de las poblaciones, y el flujo genético a larga distancia por la dispersión del polen entre poblaciones conduce a un rápido decaimiento del desequilibrio de ligamiento (De la Torre et al., 2014; et al., 2017; Porth & El-Kassaby, 2014).

En general, para las coníferas, los estudios adaptativos se han centrado en preguntas relacionadas con la resistencia al estrés térmico y a la formación de madera (Garnier-Géré & Ades, 2001; Aitken & Hannerz, 2001), elaborados a partir de una selección previa de genes o polimorfismos candidatos identificados por medio de estudios funcionales o de expresión (Neale & Savolainen, 2004) con la hipótesis de que estos loci candidatos presentarán valores de diversidad y diferenciación inusual entre las poblaciones por efecto de la selección natural (Luikart et al., 2003; Kelley et al., 2006). Mosca et al. (2016) describen para *Abies alba*, *Larix decidua*, *P. cembra* y *P. mugo*, coníferas de distribución alpina en Europa de uno a siete polimorfismos, dependiendo del género, asociados a la resistencia al congelamiento. En respuesta al estrés térmico por congelamiento o aridez en *Pseudotsuga menziesii* Eckert et al. (2009) reportaron 10 marcadores asociados a la respuesta de resistencia y adaptación al congelamiento. Otros estudios han detectado cambios correlacionados con diferencias fenológicas en *Picea mariana* (Prunier et al., 2011), y cambios ligados a diferencias en el crecimiento en la madera de *Picea sitchensis* (Holliday et al., 2010).

En México, los estudios de adaptación local en coníferas no han encontrado correlaciones entre los mecanismos genéticos de respuesta al estrés abiótico y los cambios ambientales. Pero se ha podido correlacionar la diversidad genética en *Pinus* con los procesos climáticos a nivel histórico para ubicar patrones filogeográficos (Cuenca et al., 2003; Jardón-Barbolla et al., 2011; Rebolledo-Camacho et al., 2018; Moreno-Letelier et al., 2008). Eventos

como los ciclos interglaciares del Pleistoceno y el surgimiento de la Faja Volcánica Transmexicana generaron cambios en los patrones de distribución de la diversidad genética en *Abies* (Giles-Pérez, 2017; Méndez-González et al., 2017), *Juniperus* (Moreno-Letelier et al., 2014; Reyes-Galindo, 2016), *Picea* (Jaramillo-Correa et al., 2006; Quiñones-Pérez et al., 2014), *Taxodium* (Hernández-Leal, 2009) y *Pinus* (Cuenca et al., 2003; Jardón-Barbolla et al., 2011; Rebolledo-Camacho et al., 2018; Moreno-Letelier et al., 2008).

Entre los estudios evolutivos y de historia demográfica en *Pinus* distribuidos en México debemos mencionar el de Cuenca et al. (2003), que describe en *P. nelsonii* una colonización a larga distancia entre 125,000 y 309,000 años y una expansión de 425 veces su tamaño poblacional entre 59,000 y 146,000 años atrás. Jardón-Barbolla et al. (2011) y Rebolledo-Camacho et al. (2018) describen en *P. caribaea* una reducción drástica en el tamaño demográfico, por lo que sugirieron que la endogamia ha jugado un papel importante en la disminución de la variación genética, con una dinámica demográfica de eventos de contracción y expansión generados por cambios climáticos de sequía e inundaciones extremas ocurridos durante los últimos 10,000 años. Moreno-Letelier et al. (2008) encontraron en *P. strobiformis* una diferenciación genética en dos grupos separados por el Desierto Chihuahuense, que han mantenido flujo genético durante las etapas glaciales y ciclos de expansiones poblacionales, favoreciendo la conectividad entre grupos.

### 5. Los pinos piñoneros (subsección *Cembroides*)

Distribuidas en ambientes áridos de América del Norte, las 13 especies de pinos piñoneros (subsección *Cembroides*) tienen semillas agrandadas con un megagametofito rico en lípidos, con un ala vestigial o sin ella. Características como el número y la rigidez de las acículas, la longitud del cono y el tamaño de la semilla son indicadores de la divergencia morfológica (Malusa, 1992; Farjon, 1996; Montes et al., 2019). Estudios sugieren que las semillas agrandadas sin alas evolucionaron en dos o tres eventos independientes de un ancestro común con una semilla pequeña y alada (Gernandt, 2005).

En especies de conos pequeños de la subsección *Cembroides* se han detectado eventos de introgresión y flujo genético interespecífico. Lanner & Phillips (1992) analizaron la introgresión de *P. edulis* y *P. monophylla* a partir de la variación en caracteres morfológicos de poblaciones simpátricas. Little (1968) sugirió la posibilidad de eventos de hibridación

entre *P. remota*, y *P. edulis*. Por su parte Montes et al., (2019) han descartado la hibridación entre *P. quadrifolia* y *P. monophylla* propuesta por Lanner (1974a,b) para explicar la variación en el número de acículas por fascículo de *P. quadrifolia*. Los eventos en el clado de pinos piñoneros de conos pequeños han sido favorecidos por la superposición de fenología en poblaciones simpátricas, la dispersión aerófila del polen y las débiles barreras precigóticas dada su reciente divergencia.

Ortiz-Medrano et al. (2016) describen que, en ambientes donde se incrementa la temperatura y disminuye la humedad se incrementa la cubierta de la semilla, las dimensiones del cono y la altura del árbol, y disminuye el número de acículas por fascículo. Las especies de linajes con divergencias más ancestrales de los pinos piñoneros de México son también las especies que habitan áreas con condiciones ambientales más húmedas, lo que sugiere que los caracteres ancestrales del grupo están relacionados con ambientes húmedos y fríos, a partir de los cuales el resto de las especies han divergido y colonizado ambientes más áridos (Ortiz-Medrano et al., 2016).

En el Tercer Capítulo, analizo las divergencias y cambios en los linajes de pinos piñoneros de conos grandes, para reconstruir las relaciones filogenéticas comparando polimorfismos en regiones de DNA nuclear y el genoma plastídico a fin de resolver la hipótesis propuesta por Ledig et al., (2009), que sugieren que la divergencia de los pinos piñoneros de conos grandes ha presentado eventos de introgresión.

### 5.1. *Pinus pinceana* Gordon & Glend.

*Pinus pinceana* es un pino piñonero endémico de México que se distribuye en suelos calcáreos y rocosos (Farjon & Styles, 1997). Se conocen pocas poblaciones, las cuales son pequeñas, dispersas y discontinuas, y están distribuidas a lo largo de la Sierra Madre Oriental y el Desierto Chihuahuense (1480 - 3000 msnm). Esta especie forma parte de las especies de pinos que han sido descritas como más tolerante a condiciones de sequía (Luna-Cavazos et al., 2010; Ortiz-Medrano et al., 2016). La especie está amenazada por su distribución limitada, el aumento en la erosión del suelo, el sobrepastoreo y los incendios inducidos (Martíñón-Martínez et al., 2010; Figueroa-Corona, 2016). Está catalogada como especie en peligro de extinción en la lista de especies amenazadas de la legislación nacional y en el índice internacional de la Unión Internacional para la Conservación de la Naturaleza (IUCN,

2010; NOM 059-SEMARNAT-2010; <https://www.biodiversidad.gob.mx/especies/catRiesMexico>).

Estudios genéticos a partir de diferentes marcadores en *P. pinceana* han mostrado que existe una alta diferenciación genética entre sus poblaciones (Escalante, 2001; Ledig et al., 2001; Molina-Freaner et al., 2001) en comparación con otras especies de pinos (*e.g.*, Delgado et al., 1999; Cuenca, 2003; Karhu et al., 2006). Escalante (2001) determinó con evidencia de microsatelites plastídicos que existen dos grupos genéticos, los cuales se han mantenido sin flujo genético.

A nivel morfológico, Martiñón-Martínez et al. (2010) han descrito que las poblaciones de *P. pinceana* presentan cambios en la longitud de su raíz, correlacionados con la localidad geográfica. Ramírez-Herrera et al. (2008) encontraron una relación positiva entre la longitud de las raíces y la abundancia de cera en las hojas con la ubicación geográfica de los organismos; por tanto, sugirien una adaptación diferencial al estrés hídrico y las altas temperaturas; siendo que, en las poblaciones del Desierto Chihuahuense hay un mejor desempeño de los individuos en condiciones de estrés hídrico y de temperatura elevada.

El texto está estructurado para reconocer en *Pinus pinceana* la respuesta transcripcional a factores bióticos y abiótico en su particularidad ambiental actual, para ir desarrollando el efecto de la historia demográfica y la filogeografía en la diversidad genética, y, finalmente, reconocer los patrones de divergencia de *P. pinceana* con sus especies hermanas.

## Objetivos

- Ensamblar *de novo* el transcriptoma de acículas de árboles maduros de *Pinus pinceana* (Gordon & Glend.).
- Identificar los perfiles de expresión diferencial de *Pinus pinceana* y correlacionar las diferencias a la regulación de respuesta a factores abióticos y bióticos presentes en la distribución.
- Inferir cómo los procesos geológicos y climáticos durante el Pleistoceno han influido en la variación genética y la historia demográfica de *P. pinceana* utilizando polimorfismos en todo el genoma.
- Reconocer la variación intraespecífica del genoma plastídico y polimorfismos en genes nucleares en los pinos piñoneros de conos grandes.
- Analizar la divergencia entre *P. maximartinezii* (Rzed.) y *P. pinceana* e identificar posibles eventos de reticulación entre los linajes de los pinos piñoneros de conos grandes.



## Capítulo 1

### **Transcriptome of weeping pinyon pine, *Pinus pinceana*, shows differences across heterogeneous habitats**

Laura Figueroa-Corona, Patricia Delgado Valerio, Jill Wegrzyn & Daniel Piñero

**Trees structure and function.**

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El siguiente capítulo está constituido por el artículo de requisito, publicado en la revista *Trees structure and function* el 19 abril de 2021. Los fundamentos de la publicación parten de la hipótesis de que la plasticidad fenotípica incluye rasgos fisiológicos afectados por cambios sutiles en los niveles de expresión de mRNA. Con esta consideración evaluamos la respuesta diferencial dentro de la heterogeneidad climática de *Pinus pinceana* (G. Gordon & Glend). Este pino piñonero presenta una distribución geográfica fragmentada que sugiere una adaptación a las condiciones áridas y extremas de las laderas orientales de las montañas del Desierto Chihuahuense, con variaciones significativas en la temperatura y precipitación anual. Evalué si la divergencia fenotípica está respaldada por diferencias en los perfiles de expresión de árboles adultos a lo largo de su distribución natural y si se correlaciona a los cambios en la sequía y otras presiones ambientales.

Los resultados principales incluyen la reconstrucción de un transcriptoma completo con 45,431 anotaciones de genes codificantes a proteínas del tejido de acículas de siete individuos adultos. Para la comparación entre hábitats se encontró que los árboles de las regiones más secas compartieron respuestas activas relacionadas con respuestas metabólicas correlacionadas a factores abióticos. El análisis de expresión diferencial identificó intermediarios y reguladores de la respuesta al estrés abiótico para 26 genes, destacando las familias ortólogas involucradas en la respuesta ambiental al estrés abiótico, como la serina / treonina-proteína quinasa LRR similar al receptor y L -receptores quinasas de lectina de tipo (LecRK), directamente relacionados con la regulación de la respuesta de defensa a patógenos. Cuando se compararon muestras del Desierto Chihuahuense con muestras externas a la distribución natural, en donde encontramos la expresión diferencial de un regulador de la morfogénesis de la raíz, la proteína quinasa tipo receptor rico en cisteína (CRK28), que podría relacionarse con la absorción diferencial en condiciones de sequía.



# Transcriptome of weeping pinyon pine, *Pinus pinceana*, shows differences across heterogeneous habitats

Laura Figueroa-Corona<sup>1,2</sup> · Patricia Delgado Valerio<sup>3</sup> · Jill Wegrzyn<sup>4</sup> · Daniel Piñero<sup>2</sup>

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## Abstract

**Key message** We reconstructed the needle tissue Transcriptome of *P. pinceana* for individuals from distinct biogeographic regions across a temperature and precipitation gradient that represents its natural distribution. Gene expression analysis via RNA-Seq identified differential response to biotic stress.

**Abstract** Phenotypic plasticity includes physiological traits affected by subtle changes in mRNA expression levels, causing different molecular phenotypes. We assessed the differential response to climate heterogeneity across the geographic range of *Pinus pinceana* G. Gordon & Glendinning. This pinyon pine shows a fragmented geographic distribution suggesting adaptation to the arid and extreme conditions of the eastern mountain slopes of the Chihuahuan desert. The geographic distribution of *P. pinceana* spans regions with significant variation in annual temperature and precipitation. We tested whether phenotypic divergence is supported by differences in expression profiles in *P. pinceana* mature trees along its natural distribution and corresponds to the changes in drought and other environmental stress. The reconstructed Transcriptome included 45,431 high-quality annotations derived from the needles of seven individuals across contrasting biogeographic and climatic localities. Trees from the driest regions shared active responses related to abiotic factors. The differential expression analysis identified intermediates and regulators of abiotic stress response for 26 genes, highlighting families involved in the environmental response to abiotic stress, and proteins linked to up-regulated responses, such as LRR receptor-like serine/threonine-protein kinase and L-type lectin receptor kinases (LecRK), directly related to pathogens. When Chihuahuan desert samples were compared with arboretum samples, we found differential expression of a regulator of root morphogenesis, cysteine-rich receptor-like protein kinase (CRK28), which could relate to differential absorption in drought conditions.

**Keywords** *Pinus pinceana* · Transcriptome · Drought conifer response · Weeping pinyon

## Introduction

Response to changing environments is frequently controlled through modification of mRNA expression and polymorphic variation, which enable phenotypic plasticity and adaptation (Schlichting 1986; Pfennig et al. 2010). Phenotypic plasticity includes physiological traits influenced by changes in mRNA expression levels and causes differential performance in response to environmental conditions (Ackermann et al. 2013; Schlichting 1986). When this plasticity occurs as a consequence of differential expression of the same genotype, directly or indirectly correlated with environmental factors, it could imply selective pressures (Henriques et al. 2018). Over several generations, this phenotype could become fixed in the population and recognized as a local adaptation.

Population studies have been conducted to understand response to drought through the genotype–phenotype

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✉ Daniel Piñero  
pinero@unam.mx

<sup>1</sup> Posgrado en Ciencias Biológicas, Universidad Nacional Autónoma de México, Ciudad de México, Mexico

<sup>2</sup> Departamento de Ecología Evolutiva, Instituto de Ecología, Universidad Nacional Autónoma de México, Ciudad de México, Mexico

<sup>3</sup> Facultad de Agrobiología, Universidad Michoacana de San Nicolás de Hidalgo, Uruapan, Mexico

<sup>4</sup> Department of Ecology and Evolutionary Biology, University of Connecticut, CT 06269 Storrs, USA

environment relationship. This has been approached via ecophysiological studies (Noctor et al. 2014; Álvarez-Yépez et al. 2014), provenance trials (Correia et al. 2006), and quantitative trait locus mapping (González-Martínez et al. 2006; Aitken et al. 2008; Neale and Kremer 2011). Transcriptional profiles are also frequently studied to examine response to abiotic stress, such as drought. Transcriptomic approaches are much more accessible than genome-scale studies in conifers due to their large genome sizes and significant complexity (repeat content and pseudogenes) (Biol et al. 2013; Neale et al. 2014; Gonzalez-Ibeas et al. 2016; Perera et al. 2018).

Species of conifers that grow along environmental gradients have developed plastic and adaptive mechanisms to respond to fluctuations in water availability to limit exposure and damage. Some conifers distributed in boreal regions, including *Larix decidua*, lose their needles seasonally to avoid freezing, which can result from the storage of air in the vascular tissue (Roden et al. 2015). Others reduce exposure by allowing the maintenance of osmotic balance through stomata closure. This is exhibited by members of the Cupressaceae and Taxaceae (McDowell et al. 2008; Brodribb et al. 2014). Finally, conifers demonstrate drought resistance and restorative mechanisms such as phytohormonal regulation; in the case of *Pinus pinaster*, this is associated with the accumulation of proline, abscisic acid (ABA), and jasmonic acid in the needles (Corcuera et al. 2012).

Distributed across arid environments of North America, the 13 species of pinyon pines (subsection *Cembroides*) have enlarged seeds with lipid-rich endosperms and the absence of a vestigial wing. Their morphological divergence, such as needle number, needle stiffness, length of cone, and seed size (Malusa 1992; Farjon 1996; Montes et al. 2019), is associated with different gradients of aridity (Ortiz-Medrano et al. 2016). Among the pinyon pines of North America, *P. pinceana* can withstand conditions of extreme aridity. It is an endemic species restricted to rocky soils in the Chihuahuan desert (ChD) and the Sierra Madre Oriental (SMO; 1480–3000 masl) (Farjon et al. 1997; González-Elizondo et al. 2012; Martínez-Ávalos et al. 2015) and is listed as endangered in the Mexican Official Law (NOM 059-SEMARNAT-2010; <https://www.biodiversidad.gob.mx/especies/catRiesMexico>).

Morphological differences in *P. pinceana* have been reported across biogeographic provinces. Martiñón-Martínez et al. (2010) detected significant differences in needle wax cover, which was thicker over needles in juvenile individuals from ChD compared to juveniles from SMO. They also reported that thermal increments produced faster growth, likely due to higher water consumption. Córdoba-Rodríguez et al. (2011) described larger primary roots and greater volume on secondary roots in ChD localities. Both studies described distinct physiological differences in

contrasting drought conditions. Martínez-Martiñón et al. (2010) reported that in a common garden experiment controlling temperature and humidity, ChD juveniles had less biomass but assigned more resources to root growth than SMO individuals. Population genetic approaches detected that *P. pinceana* exhibited greater diversity and differentiation across its fragmented geographic distribution than would be explained by isolation due to distance (Escalante 2001; Ledig et al. 2001; Molina-Freaner et al. 2001).

Assuming there are associations between environmental variables and expression profiles, we examined mature *P. pinceana* trees and report on three aspects: (1) characterization of the climatic differences along the natural range of *P. pinceana*, (2) a de novo assembled and annotated needle Transcriptome for the species, and (3) differential expression analysis in relation to environmental conditions.

## Materials and methods

### Climatic characterization

From 57 registered localities recorded in the National Herbarium of Mexico (MEXU), we retained 22 records verified in the previous literature (Escalante 2001; Molina-Freaner et al. 2001), at least 800 m from each other. To construct a potential niche model using the default parameters, we used Maxent v3.3.3e (Phillips and Dudík 2008). Climatic information for each record was extracted from 19 bioclimatic layers (World Clim; Hijmans et al. 2005). To quantify the annual drought changes within the sampled populations, we used four layers from the Global Aridity Index (Trabucco and Zomer 2009). The latter is constructed with the relationship of precipitation and water content in the soil with data from 1970 to 2000 and a 30-arcsecond resolution.

To avoid redundancy in environmental information, we performed pairwise correlations among all variables. Among the highly correlated pairs ( $r^2 > 0.75$ ), we chose variables that best explained the geographic distribution for *P. pinceana* using a threshold of the minimum training presence of 0.5. A restricted niche model was also built in Maxent using annual mean precipitation, isothermality, temperature changes per season, annual mean temperature, and soil water content in the warmest quarter of the year (Fig. S1; Table S1). The information was extracted at each sampled location using the GARP utility v1.0 (Stockwell 1999) on ArcGIS (ESRI Environmental Systems Research 2011). Subsequently, a principal components analysis (PCA) was conducted with *devtools* and *ggbiplot* packages in R (Wickham 2016) to quantify the distribution's environmental variance.

## Transcriptome characterization

### Plant material, cDNA library construction, and sequencing

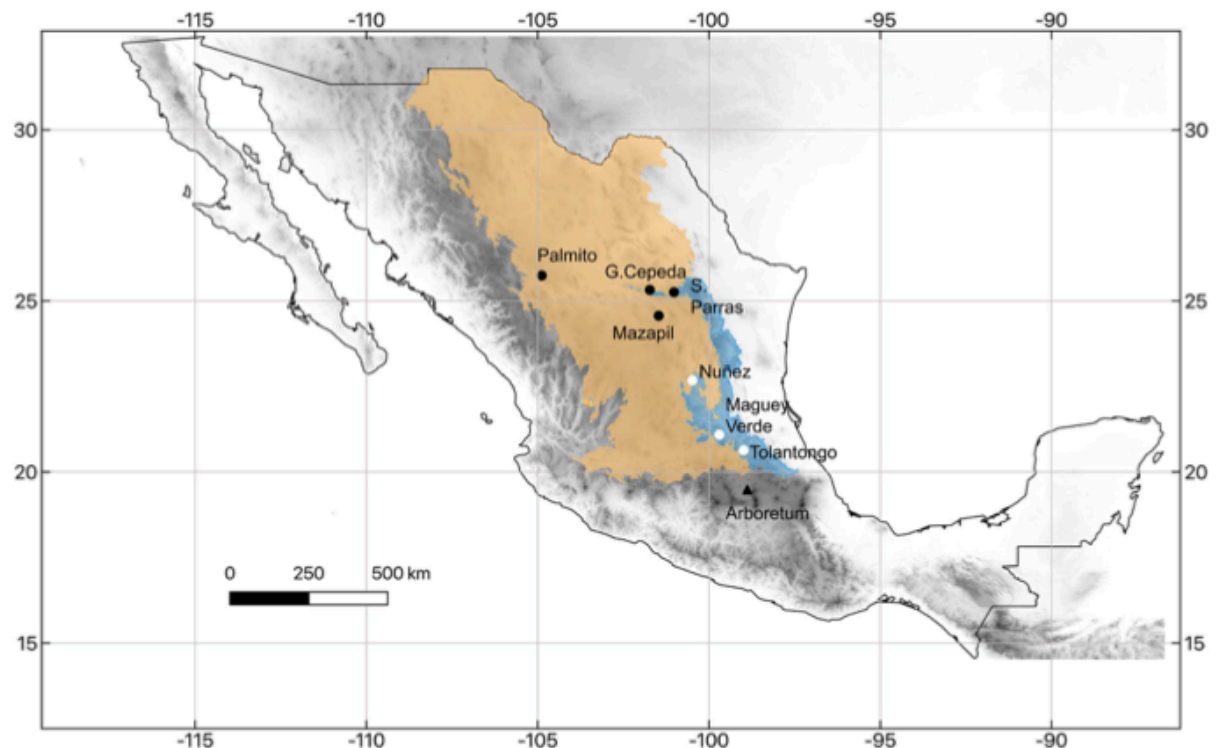
Foliar tissue was collected in the dry season (11–21 April 2018) from seven healthy adult individuals selected at random in seven localities of the natural distribution, located in two different biogeographic provinces (Morrone et al. 2017): localities General Cepeda, Sierra de Parras, Palmito, and Mazapil in ChD and localities Nuñez, Maguey Verde, and Tolantongo in SMO. We also sampled two individuals from the arboretum of the Universidad Autónoma de Chapingo in Texcoco City, Mexico, located outside the altitudinal range and the natural distribution (Fig. 1; Table S1).

RNA was extracted using the Spectrum Sigma® protocol (Sigma Aldrich, St. Louis, MO, USA), which uses  $\beta$ -mercaptoethanol to make the lysate, in accordance with the supplier's recommendations. We used the protocol adapted for conifer needles, or leaves with normal content of water and starch storage organs, to bind the RNA using a column. Wash, dry, and elution steps followed standard practice. The cleaned product was visualized on a 1% agarose gel stained with ethidium bromide. To evaluate the occurrence of salts,

we quantified all samples with a concentration greater than 100 ng/ml and a 260:280 ratio between 1.8 and 2 samples were further processed in the Genomics Sequencing Laboratory in the Institute of Quantitative Biosciences (QB3) at the University of California, Berkeley. The RNA integrity was first evaluated for values greater than 7.0 in an Agilent 2100 Bioanalyzer (Agilent Technologies, Santa Clara, CA, USA). cDNA libraries were constructed for each individual following Illumina TruSeq RNA standard protocol (Illumina, San Diego, CA, USA). Nine libraries were multiplexed and sequenced on a single run of an Illumina HiSeq 4000 (150-bp paired-end reads).

### Transcriptome assembly

The adapters were removed, and quality filtering was performed with TRIMMOMATIC v0.36 (Bolger et al. 2014) to retain sequences which were at least 80 bp in length and had a minimum Phred-scaled quality score of 35. The procedure was visualized and confirmed with FASTQC/MULTIQC (Ewels et al. 2016). A de novo assembly for each locality was executed for each paired-end read from each library. This was carried out with Trinity RNA-Seq v2.0.2



**Fig. 1** Localities of *Pinus pincea* sampled. Black dots correspond to natural distribution, and white dots correspond to pines collected in the arboretum of the Universidad Autónoma de Chapingo, outside its

natural distribution. The Chihuahuan desert (ChD) region is shown in orange and Sierra Madre Oriental (SMO) in blue

(Grabherr et al. 2011), requiring paired reads and a minimum contig length of 300 bp, and independently using rnaSPAdes v3.9 CPU following an assembly for paired reads (Bankevich et al. 2012). The assemblies were processed via USEARCH v9.0.2132 to reduce the redundancy of transcripts within each assembly (0.9 similarity index; Edgar 2010). Local clustered assemblies were frame-selected using Transdecoder v3.0.1 (Haas et al. 2013) to identify the longest and most likely open reading frames. To further refine the reference Transcriptome, we built a global reference with the 18 local assemblies provided by Trinity and rnaSPAdes (following clustering and frame-selection) via EvidentialGene v20190101 (Gilbert 2013). The *trformat* script reduced redundancies on the sequence identifiers, the *tr2aacds* script selected transcripts from the highly similar sequences, and *evgmr-na2tsa* formatted the selected sequences.

### Ortholog annotation and function

Each reduced local library and the global reference contigs were processed and annotated with EnTAP v 0.9.0-beta (Hart et al. 2020). EnTAP utilizes Diamond v0.9.19 for sequence similarity searches and EggNOG v0.99.1 (Huerta-Cepas et al. 2018) to provide gene family assignments. Functional annotation was carried out against NCBI Refseq Complete-Protein, Refseq Plant-Protein, and Swissprot UniProt using a threshold of 70% similarity target and an e-value of  $10^{-6}$  using DIAMOND (O'Leary et al. 2015; UniProt Consortium 2018). In addition, we selected and separated the matches that corresponded to potential contaminants, such as insects, fungi, and bacteria. The remaining sequences were annotated with Gene Ontology (GO) terms using Blast2GO (Conesa et al. 2005) and InterProScan v5.25 (Jones et al. 2014) for the identification of conserved domains using the Pfam-A v31.0 database. The integrity and quality were reviewed with the single-copy ortholog benchmarking tool BUSCO (v3.0; Waterhouse et al. 2018) and the Embryophyta database (Obd10).

Finally, using the annotated genes from the seven localities as independent proteomes, we composed putative gene families (orthogroups). By considering the classification within orthogroups, we could discriminate a set of genes that descended from a single gene of the considered localities. We did this to minimize the heterogeneity among samples (partial transcripts and isoforms characteristic of de novo assemblies) and maintain the gene families in all localities. Using OrthoFinder v2.4, we compared the translated proteins from the different localities and generated sets of single-copy genes as well as shared families (Emms and Kelly 2015).

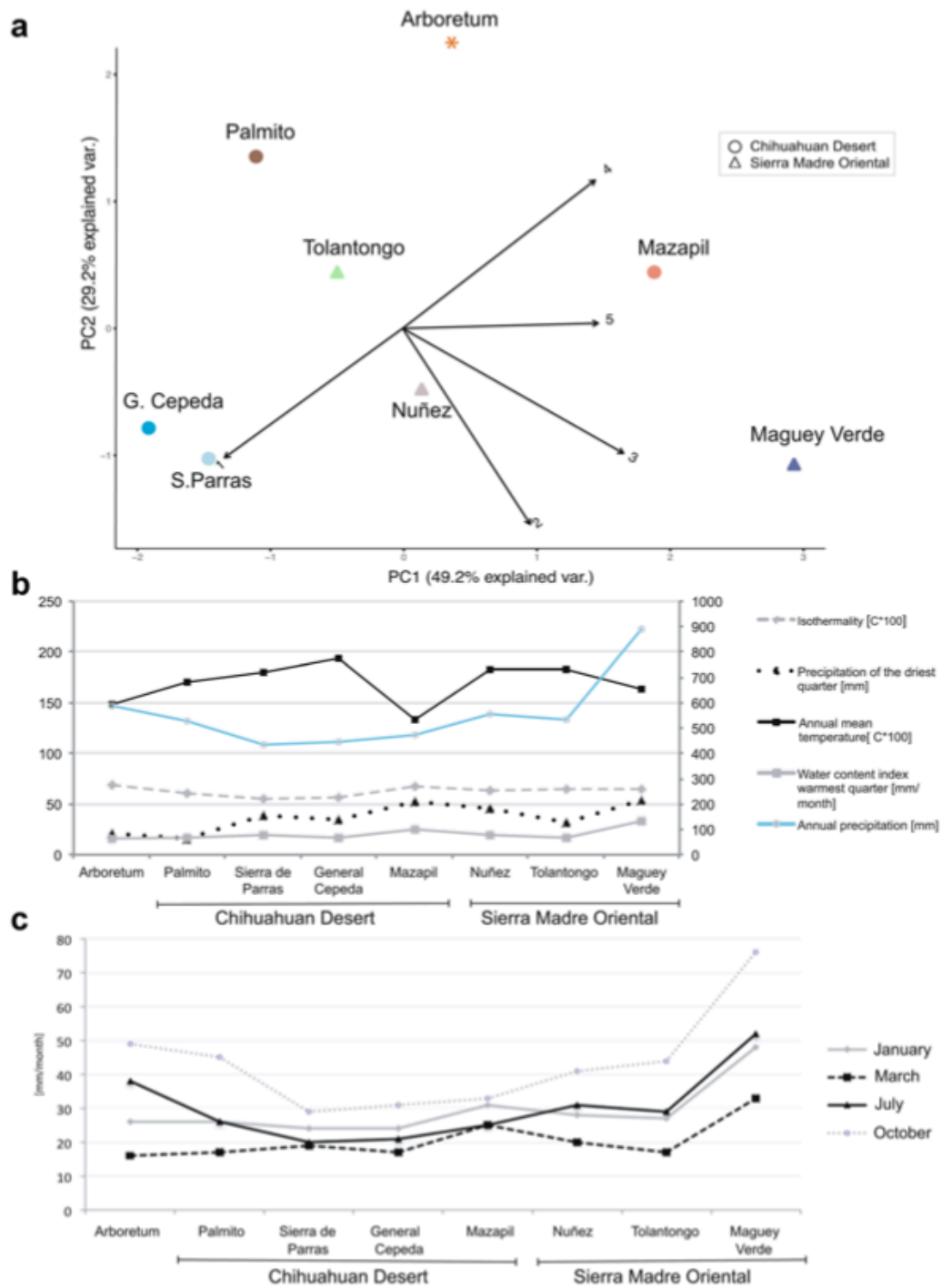
### Differentially expressed transcripts

The trimmed paired-end reads from each of the nine libraries were mapped to the global reference of *P. pinceana* to calculate transcript abundance via Kallisto v0.44.0 with a pseudo alignment using 100 bootstraps (Bray et al. 2016). To construct and analyze the expression profile, we used the R package for differential expression, DESeq2 v1.26, available in Bioconductor (Love et al. 2014), and all mapped genes were normalized by DESeq2 median ratio (reads/Kb/mapped). Significantly expressed transcripts were defined at a *p*-adjusted value  $< 0.1$  to report the less differences in samples very heterogeneous among them. Fold change values were reported for samples clustered by region: four libraries from ChD localities and three libraries from SMO localities. We compared samples from the two biogeographic provinces using the localities as biological replicates to understand the transcriptional changes across the natural distribution using the  $\log_4$ -fold change to obtain a conservative criterion for differences across regions. To control for phenotypic plasticity variation, we conducted a second comparison between each biogeographic province and the two arboretum samples.

## Results

### Climatic differentiation across the distribution range

The niche-modeled distribution was assigned to zones whose climatic characteristics were at least 50% similar to those of the used localities, defining the specific conditions in which *P. pinceana* grows (Table S2). Variables with the most significant effect on niche modeling were the annual mean temperature, precipitation of the driest quarter, water content index in the warmest quarter of the year, and isothermality (Table S3). PCA showed that the most considerable portion of the variance was explained by PC1 (49.2%) associated with the water content index in the warmest quarter of the year and isothermality. In comparison, PC2 (29.2%) showed the largest loadings of mean annual temperature and precipitation in the driest quarter of the year, demonstrating the environmental heterogeneity between ChD and SMO localities (Fig. 2a). Using locality values, we evaluated the seasonal changes in soil water content. We found that the most arid conditions in ChD occurred during the final quarter of the year, when SMO localities Maguey Verde and Tolantongo had the highest precipitation (Fig. 2b, c).



**Fig. 2** Climatic characterization of *Pinus pincea* geographic distribution. **a** Principal components analysis for the sampled localities (triangles show the Sierra Madre Oriental populations; circles, the Chihuahuan desert localities) and the climatic conditions: 1, annual average mean temperature; 2, precipitation of the driest quarter of the year; 3, water content index in the warmest quarter of the year;

4, isothermality; 5, annual precipitation. **b** isothermality, precipitation of the driest quarter of the year, water content index in the warmest quarter, and annual precipitation across localities. **c** Seasonal changes in water content on soil index for each of the studied sampled localities

## Transcriptome assembly and annotation

The paired-end RNA sequencing produced 37.3 million reads on average for each library, which was reduced to 30.7 million after quality control. From the de novo assemblies, the average total transcripts for each locality were 109,336 with an average length of 964 bp (N50 1459 bp) and BUSCO completeness of 79% (Table 1). We used USEARCH to reduce the redundancy in each assembly. This produced an average of 90,755 sequences for each library (Table S4). Processing with Transdecoder to determine open reading frames produced an average of 62,219 coding sequences per library.

Through Evgene, we built the global reference of 149,408 sequences from 18 original assemblies. A total of 27,952 sequences aligned to the databases selected in EnTAP. Of these, 16,276 were considered informative (based on the description provided by the target sequence). The other 11,676 were defined as uninformative (with descriptions focused on terms such as hypothetical or unknown). Alignments to sequences of the model moss, *Physcomitrella patens* (461; 1.66%) were prevalent, as well as more basal angiosperms, including *Amborella trichopoda* (2,838; 10.15%) and *Nelumbo nucifera* (631; 2.26%). In addition, other angiosperms, *Arabidopsis thaliana* (1,510; 5.4%), *Cucurbita pepo* (1,156; 4.14%), *Quercus suber* (2,303; 8.24%), and *Elaeis guineensis* (536; 1.9%), were prevalent. This phylogenetic bias can be understood as an artifact consequence of the limited genomic resources available for gymnosperms, where among genera, only 349 sequences were aligned (1.25%). Gene family assignment via EggNOG identified 45,135 transcripts that could be assigned. In addition, a subset of 34,232 sequences was assigned to at least one GO term. In total, 45,431 (54.2%) transcripts were assigned to either an EggNOG gene family or aligned (with confidence) to a sequence in our selected databases.

At this stage, we removed 1753 sequences that aligned to contaminant categories, corresponding to bacteria (25; 1.43%), insects (348; 19.85%), and fungi (1380; 78.76%) in the global reference Transcriptome. The description and abundance of the contaminants in the local and the global assemblies are shown in Table S5. Overall, a rich representation of parasitic yeasts was present, including *Saccharomyces cerevisiae*; *S. pombe*; yeasts associated with wood-ingesting insects, including *Aureobasidium namibiae*, *Candida albicans*, and *Sugiyamaella lignohabitans*; and pathogens of termites, like *Cryptotermes secundus*. The identified contaminants were consistent across all assemblies, including the arboretum samples. We detected an abundant presence of pathogenic fungi in SMO localities (Maguey Verde and Nuñez), including *Diplodia corticola*, *Bipolaris maydis*, *B. oryzae*, *Sclerotinia sclerotiorum*, *Cercospora beticola*, and the endophytic fungus *Xylona*

**Table 1** Sequencing summary statistics sequencing and de novo assembly

Sample	Condition	Total number of reads sequenced (M)	Total number of reads trimmed (M)	Trinity			rnaSPAdes				
				Total transcripts	Transcript N50	Median length	BUSCO completeness	Total transcripts	Transcript N50	Median length	BUSCO completeness
Arboretum		36.9	29.1	85,286	1721	708	82.9%	15,645	2085	473	76.8%
Arboretum		46.7	39.4	117,534	1565	602	87.0%	21,284	1355	591	87.8%
Tolantongo	SMO	26.5	21.7	86,111	1569	607	80.2%	17,023	300	546	80.4%
Maguey Verde	SMO	45.5	40.1	148,791	1396	559	83.1%	23,887	294	570	83.7%
Nuñez	SMO	40.6	33.1	82,559	1156	501	61.5%	13,327	722	502	64.8%
Mazapil	ChD	37.7	30.5	106,542	1500	589	84.6%	19,311	247	582	85.5%
Sierra de Parras	ChD	28.4	23.8	110,243	1416	565	79.5%	19,259	235	567	80.7%
General Cepeda	ChD	41.6	34.2	118,247	1524	598	84.6%	20,632	259	577	84.9%
Palmito	ChD	32.2	25	128,715	1286	541	77.6%	21,037	2763	523	77.8%



*heveae*. In ChD localities, we detected more abundance and diversity of pathogenic insects, including *Dendroctonus ponderosae* in Palmito and *Cryptoterms secundus* in Mazapil and General Cepeda. We also found across localities ant species such as *Vollenhovia emeryi*, *Trachymyrmex septentrionalis*, and *T. zeteki*. The natural growth conditions can likely explain their presence as many of the species listed as contaminants are known to be pathogenic, commensal, or mutualistic to *P. pinceana*.

Following removal of the contaminants, the final Transcriptome represented 147,531 sequences. The estimated completeness of the 45,431 annotated genes (a subset of the full reference) was 89.3%, as estimated by the BUSCO Embryophyta database. Sequences retained in each annotation stage are described for the local assemblies in Table 2.

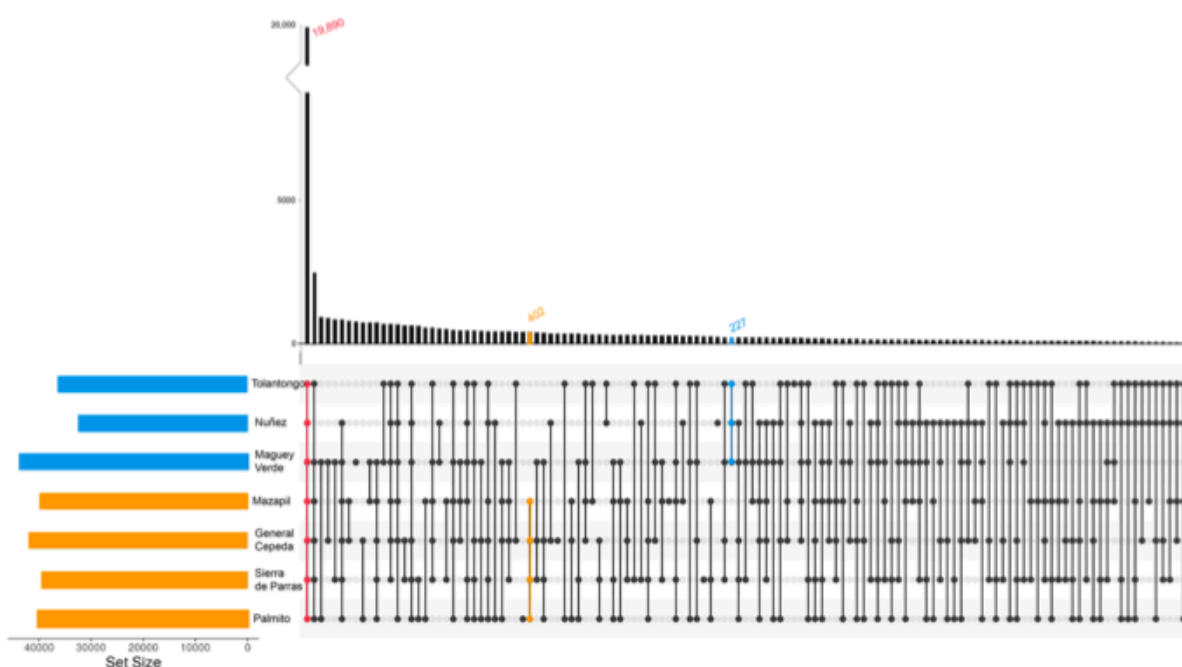
### Gene family analysis

The final location-specific transcriptomes revealed high-quality annotations: 29,731 in Tolantongo (SMO), 42,590 in Maguey Verde (SMO), 27,217 in Nuñez (SMO), 32,842 in Mazapil (ChD), 36,279 in General Cepeda (ChD), 33,012 in Sierra de Parras (ChD), and 33,556 in Palmito (ChD; Table 2). Using each locality as an identity unit in an OrthoFinder analysis, 58,757 orthogroups representing 403,614 transcripts were generated (Table S6). The expressed core present in all localities of *P. pinceana* was composed of 19,890 orthogroups representing 223,273 genes (33.8% of the total) (Fig. 3). These orthogroups contained at least 2 and as many as 115 genes. The locality-specific orthogroups ranged from 773 families represented by 1,827 genes (1.31% of the total) identified in Maguey Verde to 240 families represented by 514 genes (0.4%) in Tolantongo. Between biogeographic provinces, more shared gene families were found between Maguey Verde (SMO) and Palmito (ChD; 933) than between Maguey Verde and General Cepeda (ChD; 908; Table S6).

Present exclusively in ChD localities, we detected 1,992 genes representing 402 (0.68%) of the orthogroups. We identified gene families associated with abiotic stress response, like RNA polymerase II promoter in response to salt stress (GO:0036251); cell wall organization; and biogenesis of cell wax, such as MAP kinase kinase activity (GO: 0038068). Restricted to SMO localities, we found 885 function enriched genes grouped in 227 orthogroups (0.38%), including deacetoxycephalosporin-C hydroxylase activity (GO:0045442). Finally, orthogroups GO:0045414, GO:0045430, GO:0045442, GO:0045493, GO:0045522, GO:0045525, and GO:0045526 were related to interleukin production and regulation responsible for plant disease resistance and response to pathogens (Meyers et al. 2002).

**Table 2** Sequences processed at sequential annotation stages

	Arboretum 1	Arboretum 2	Tolantongo	Maguey Verde	Nuñez	Mazapil	General Cepeda	Sierra de Parras	Palmito	Global reference
1. Similarity search	24,603	27,017	22,670	30,406	19,211	24,905	27,124	24,417	24,911	27,952
2. EggNOG	32,301	35,335	29,590	42,367	27,094	32,692	36,124	32,852	33,384	45,135
3. Gene ontology	26,746	29,042	24,490	35,158	22,751	26,863	29,751	27,079	27,689	34,232
4. HQ annotations	32,429	35,522	29,731	42,590	27,217	32,842	36,279	33,012	33,556	45,431



**Fig. 3** Orthogroups shared among localities. Tiling beneath the histogram indicates the membership of genes to each orthogroup in the set. Set size is displayed as height on histogram. Horizontal histogram

indicates the number of orthogroups found in each locality. Orange indicates data from Chihuahuan desert; blue, Sierra Madre Oriental localities

### Differential expression

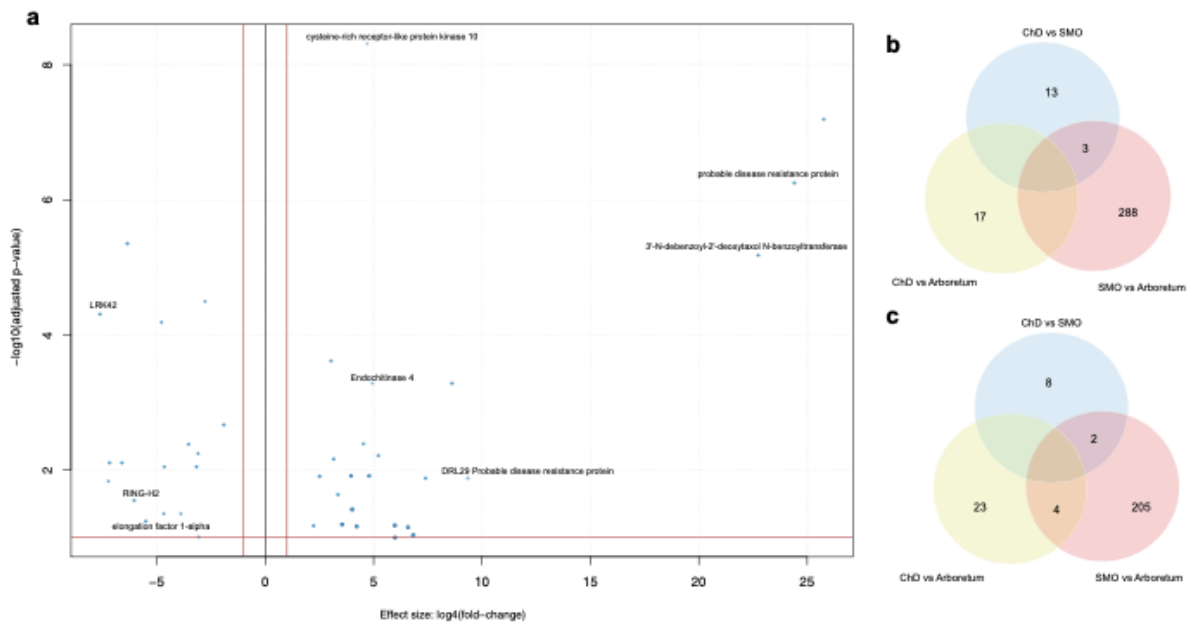
An average of 58% of the reads, per sample, aligned to the global Transcriptome reference; this shows the high diversity of transcriptional expression across localities (Table S7). The expression profiles were constructed by dividing the localities according to their biogeographic provenance. This was done to compare the climatic differences across a gradient from more humid SMO localities to drier ChD localities.

Using the Transcriptome reference of 147,531 genes, and after normalization, we removed 94,153 transcripts with low read counts and 1,216 outliers. The reference Transcriptome of 45,431 sequences was indexed for differential expression analysis. Three pairwise comparisons were conducted: SMO vs. ChD, arboretum vs. ChD, and arboretum vs. SMO.

After applying a threshold of fourfold change, we found 26 differentially expressed genes, 10 down-regulated and 16 up-regulated (Fig. 4; Fig. S2; Table S8). These genes were primarily associated with abiotic stress response and pathogen interaction. Those expressed in SMO localities included L-type lectin-domain containing receptor kinase IV.2 (LRK42) and membrane-spanning receptor-like kinases. These have putative roles in biotic and abiotic stress response and in plant development. A study in *A. thaliana* by Wang et al. (2014) reported overexpression of LRK42 upon infection of *Pseudomonas syringae* and even

suppressed pathogen resistance. In addition, involved in the response to biotic stress, we detected increased expression of finger protein ATL3 (RING-H2) related to disease resistance against tobacco mosaic virus (TMV) and *P. syringae*. This up-regulation has also been related to increased oxidative stress tolerance and drought tolerance in seed germination in rice (Liu et al. 2008). Disease-resistance proteins, such as DRL29, were significantly up-regulated in ChD individuals (samples from Sierra de Parras and General Cepeda) compared to SMO individuals. Finally, the disease-resistance protein At4g27220, with a LRR domain linked to plant immunity, was also up-regulated in ChD individuals.

The comparison between the two arboretum samples and ChD localities identified 44 differentially expressed genes, 17 up-regulated and 27 down-regulated (at least fourfold change) (Fig. S3a). In the arboretum samples, we identified up-regulated genes that are associated with pathogen recognition and the subsequent activation of plant defense mechanisms, including LRR receptor-like serine/threonine-protein kinase (At1g67720), universal stress family protein, and oxalate-CoA ligases, and related to plant defense against fungal pathogens, like phenylcoumaran benzylic ether reductase PT1 (PCBER), an intermediate in lignan and isoflavonoid production responsible for durability, longevity, and resistance of heartwoods in *Pinus taeda* and *Populus trichocarpa* (Gang et al. 1999). A pathogenesis-related



**Fig. 4** Comparison of levels of gene expression. **a** Volcano plot with the dispersion at  $\log_4$ -fold change on the comparison between Sierra Madre Oriental and Chihuahuan desert. **b** Genes up-regulated in dif-

ferent comparisons between localities and the two arboretum individuals. **c** Genes down-regulated across different comparisons between localities and the two arboretum individuals

thaumatin family protein, linked to early antifungal activity in various plant species, was significantly up-regulated in the arboretum samples (Zhang et al. 2018). The pathogenesis-related protein 1 A (PR1) is known to play an important role in stress-related infection of *Fusarium oxysporum* but also is an indicator of tolerance in tomato varieties under drought stress (Akbudak et al. 2020). In contrast, in ChD localities, we identified a threefold increase in the expression of disease-resistance protein (TAO1) compared to our arboretum samples. Gene ontology links probable disease-resistance protein (At4g33300) to *P. syringae* infection resistance in sweet pepper (Cheng et al. 2020). We also found overexpression of shikimate O-hydroxycinnamoyl transferase-like that has been identified as related to heat stress (Sun et al. 2018). Remarkably, we found overexpression of cysteine-rich receptor-like protein kinase (CRK28), regulated by ABA, which has been shown to slow down root growth and lateral root formation in *A. thaliana* (Pelagio-Flores et al. 2020).

The comparison between the two arboretum samples and SMO localities identified 502 differentially expressed genes, 291 up-regulated and 211 down-regulated (fourfold change; Fig. S3b). In SMO localities, we identified genes related to biotic response, including LRR receptor-like serine/threonine-protein kinases (At1g56140, At3g47570) associated with innate immunity in plants. In addition, proteins related to disease resistance (At5g63020) and

monooxygenase 2 (MO2) overexpressed in our samples from SMO have been linked to LRR proteins as a response to oomycete infections (Hok et al. 2011). In addition, we found overexpressed in SMO two different TMV-resistance proteins N that, with LRR proteins, have been shown to activate innate immunity regulating infections mediated by pathogen recognition in *Zea mays* and *A. thaliana* (Afzal et al. 2008). Linked to abiotic stress, we detected overexpression in SMO of UDP-glycosyltransferase 72B1 related to monolignol biosynthesis, lignin polymerization, and cell wall-related transcription, factors that produce secondarily thickened cells, making them rigid, which was shown by Lin et al. (2016). In addition, we found sodium/hydrogen exchanger 7, which has been linked to improving salt tolerance (Shi et al. 2003). The SMO samples show an overexpression of MYB4 transcription repressor, which encodes the control of flavonoid and shikimate biosynthesis that, under drought conditions, increases hormone signals such as ABA, jasmonate, and brassinosteroids (Wang et al. 2019). SMO samples also showed overexpression of aryl-alcohol dehydrogenase (AAD4), which participates in the degradation of lignin when carbon or nitrogen is limited (Reiser et al. 1994). In addition, we found overexpression of class II heat shock protein-like, which becomes highly abundant during heat stress but is also implicated in response to stomata regulation and response to viral or fungal disease (Lin

et al. 2018). SMO localities show overexpressed transcripts related to abiotic and biotic stress, like peroxidase 12-like, involved in removing  $H_2O_2$ , oxidation of toxic reductants, defense against insect attack, symbiosis, and normal cell growth (Coso and Dunand 2009). Linked to abiotic and biotic response, we detected three different heat shock protein-like and trans-cinnamate 4-monooxygenase involved in lignification and protection from UV-induced DNA damage, predators, and pathogens (Schillmiller et al. 2009). Proteins that show an up-regulation in the arboretum samples when compared to SMO localities were related to secondary metabolite production involved in the defense response, like taxadien-5- $\alpha$ -ol O-acetyltransferase (T5AT) involved in taxol and other alkaloid biosynthesis, and dihydroflavonol 4-reductase, linked to the biosynthesis of anthocyanins and condensed tannins involved in reproduction, growth, and protection against pathogens (Petit et al. 2007). In the arboretum samples, we also found overexpression of proteins linked to the immune response to invasive biotrophic pathogens and defense mechanisms, like four different LRR receptor-like serine/threonine-protein kinase (At3g47570, At5g59680, At5g04720, and At1g56140). Furthermore, we found overexpressed proteins that participate in the activity of two LRR-like genes with TMV protein resistance N-like activity, one gene with toll/interleukin-1 receptor-like protein function, three genes with lipase-like (PAD4) activity that induce the biosynthesis of salicylic acid (Jirage et al. 1999), and one gene with rust resistance kinase (Lr10) activity (Feuillet 1997). Mainly related to the response against pathogens, we detected that overexpression of patatin-like protein 2 promotes cell death and affects resistance to *Botrytis cinerea* and *P. syringae* in *Arabidopsis* sp. This overexpression contributes to resistance to cucumber mosaic virus (La Camera et al. 2009). We also found antifungal protein ginkbilobin-like (GNKL) linked to the resistance to infections of *Fusarium oxysporum*, *Trichoderma reesei*, and *C. albicans*. In addition, we detected nine sequences up-regulated in the SMO localities that are related to both stress (GO:0006950) and defense response (GO:0006952). Five sequences were down-regulated in the SMO samples characterized by orthologous proteins related to response to stress (GO:0006950), defense response (GO:0006952), response to reactive oxygen species (GO:0000302), response to osmotic stress (GO:0006970), or response to oxidative stress (GO:0006979).

In summary, these comparisons show that only two genes are up-regulated in ChD but down-regulated in SMO localities when compared to arboretum samples; one lacks an annotation and the other, 7-deoxyloganetin glucosyltransferase, is involved in the biosynthesis of alkaloids. We were not able to detect up-regulated genes in the arboretum samples compared to the ChD localities. In contrast, for down-regulated genes, ChD and the arboretum samples

shared four genes, but only one was annotated, a catechol O-methyltransferase. This gene is likely involved in flavonoid biosynthesis. Finally, two genes showed overexpression in SMO samples when compared with ChD and the arboretum samples, one associated with the response to ABA (GO:0009737; Fig. 4b, c) and the other not yet characterized.

## Discussion

### Transcriptome characterization

Phenotypic plasticity allows organisms to react to changes in their environment by adjusting their metabolism through the regulation of gene expression (Ackermann et al. 2013). The broad ranges of transcriptional phenotypes in the natural localities of *P. pinceana* in different environments are controlled by complex regulatory networks in response to heterogeneous environmental stimuli (abiotic and biotic).

Precipitation and temperature are highly divergent between ChD and SMO; ChD localities are typically much drier than SMO (Fig. 2; Ruiz-Sanchez et al. 2012). The climatic limits used in niche modeling and the low abundance in present day localities have contributed to *P. pinceana*'s geographic restriction. Overall, the climate characterization indicates that the index of water content at ChD localities shows less soil water accumulation during the year (Fig. 2b). Bioclimatic and physiological differences could produce ecological differentiation, reproductive isolation, and genetic divergence, as reported in events of parapatric speciation in other pines, including *P. contorta* (Eckert et al. 2012) and *P. hwangshanensis* (Zhou et al. 2017).

Between 27,217 and 42,590 genes were characterized in each of seven localities sampled for *P. pinceana* (Table 2). These numbers are largely in agreement with previous studies of conifer needle transcriptomes, as well as the estimated number of genes in the conifer genomes annotated to date. For example, 34,389 unique genes were reported in *P. lambertiana* (Gonzalez-Ibeas et al. 2016), 50,172 for *P. taeda* (Perera et al. 2018), and 23,932 in the needle Transcriptome of *P. monticola* (Baker et al. 2018).

Transcript heterogeneity among samples could be explained by variation between regions but is also related to variation in weather and time of day for the collections. Samples collected from the landscape are not subject to the controls of a greenhouse and are likely responding to both abiotic and biotic stimuli that could not be detected (Cho et al. 2018). The sampling design used in this study with only one individual for each locality has the advantage that several localities were sampled in two different biogeographic regions but also has the limitation that we cannot make inferences within a given locality.

The presence of 1753 contaminant sequences can be explained by the nature of the samples during collection and their biological interactions rather than subsequent contamination. We found insect and fungal pathogens in all localities, including the arboretum, suggesting that trees are responding to different levels of biotic stresses. In particular, the gene family term for thigmotropism (GO:0009652) is exclusively expressed in Maguey Verde and corresponds to the invasive growth of hyphae of *C. albicans* and/or *Sugiyamaella lignohabitans* (Davies et al. 1999). A phytopathological analysis in Maguey Verde is necessary since we found a relatively abundant representation of parasitic fungus in this locality. The most well-represented organisms reflect infections of *S. cerevisiae* in different localities. This organism has been reported to produce a delay in growth and even cause plantlets to die (Gognies et al. 2001). We also recognized direct pathogenic infections of wood-ingesting insects and indirect evidence from the yeast associated with them. An important pathogenic species is the mountain pine beetle (*Dendroctonus ponderosae*) since larvae feed on the phloem tissue. This does not appreciably harm the living trees, until pupae eventually mature and adults emerge from, and kill, the host tree (Amman 1982). Infections are rampant across North America, where around 8.5 million ha of forests are perturbed (Hicke et al. 2006). Recognizing the rate of insect infections in Palmito, which has also been impacted by new human settlements in the last 15 years, it will be important to evaluate and mitigate the potential damage.

The characterization of differential water absorption performance or morphological differentiation has been described in juvenile ChD individuals (Martinez-Martiñón et al. 2010; Córdoba-Rodríguez et al. 2011). This study identified candidate genes (Table S8) that can be further characterized in larger populations during both dry and rainy seasons. This can expand our understanding of local adaptation in particular localities in the same biogeographical provinces. The regional similarities that we found describe a plastic variation that corresponds to climatic heterogeneity (Fig. 2).

### Climatic differences related to changes in gene families

The 19,890 core gene families shared among sampled regions (Fig. 3) are likely related to constitutive metabolic components, such as photosynthesis elements, transcription factors, and regulatory factors of secondary metabolites. The differential abundance of exclusive orthogroups corresponds directly to the number of genes specific to different localities. The drought response-related gene family (GO:0009819, drought recovery) is present in all localities, with more representation in General Cepeda. On the other hand, genes associated with water deprivation

(GO:0042631) are present in all ChD localities (Palmito, General Cepeda, and Mazapil). The primary differences between regions included sequences related to response to stress such as herbivory. This produces short-term physiological changes and decreases nutrient assimilation, modifying the photosynthetic machinery. We found 227 orthogroups with functions associated with disease resistance and pathogen response restricted to SMO localities. In particular, we noted differential expression of genes supporting the regulation of interleukins (Bayless et al. 2020; Bréda et al. 2006).

### Differential expression in different regions reveals variations in stress regulators

Expression differences included genes related to biotic stress response. Several of these are involved in innate immunity through LRR proteins that are known to mediate pathogen recognition (Meyers et al. 2002; Eitias et al. 2008; Afzal et al. 2008). In addition, we identified pathogen response genes such as L-type lectin receptor kinases (LecRK), overexpressed in SMO in comparison to ChD localities, and disease-resistance protein (TAO1), overexpressed in ChD localities compared to arboretum samples. These proteins, detected in abundance in Nuñez, have been reported by Wang et al. (2014) and Eitas et al. (2008) as responsible for resistance to *Pseudomonas* infections of wood-ingesting nematodes and insects. We also identified patatin protein 2 (PLP2) linked to *Botrytis cinerea* and the antifungal protein ginkbilobin-like (GNKL), reported in *C. albicans* infections (La Camera et al. 2009; Gao et al. 2016).

We used samples from different localities in two biogeographic regions as biological replicates assuming environmental homogeneity across regions. In spite of this, neither climate (Fig. 2) nor gene expression (Fig. S4) showed a clear pattern among localities within each region. Likely, environmental conditions are more important for *P. pincheana* individuals than are the abiotic conditions measured here. Nonetheless, we found intermediates in the synthesis and regulation of salicylic acid, jasmonic acid, brassinosteroid-like *pr-1* genes, and lipase-like protein (PAD4) which are implicated in abiotic stress responses, such as freezing, salinity, and osmotic stress (Akbudak et al. 2020; Jirage et al. 1999).

Furthermore, the occurrence of genes that regulate oxidative stress tolerance and increase drought tolerance was more evident in the independent comparisons between arboretum samples and the natural localities. These include MYB4 transcription factors and several heat shock proteins. Both are important for heat stress, leaf water balance, and response to viral or fungal pathogens (Wang et al. 2019).

In the comparisons between the arboretum and ChD localities, the overexpression in ChD samples includes

cysteine-rich receptor-like protein kinase (CRK28), previously characterized as a regulator of abiotic stress response in relation to ABA signaling mechanisms (Lopez-Molina et al. 2001). Pelagio-Flores et al. (2020) have related it to the development of roots in drought conditions in *Arabidopsis*. This could contribute to the phenotypic differences described in *P. pinceana* by Córdoba-Rodríguez et al. (2011). The absence of this expression pattern in the comparison between the SMO remains unclear. The overlaps between arboretum and ChD, and between SMO and arboretum, are largely related to secondary metabolite production, a key aspect of conifer plant defense (Shalev et al. 2018; Tang et al. 2005).

## Conclusions

This study represents a comparative analysis of a de novo Transcriptome assembly in a pinyon pine that grows in contrasting climate localities and in a geographically fragmented distribution. We characterized transcripts across their natural distribution and examined the response of metabolic and resistance pathways impacted by abiotic stress. The differences in transcript abundance in the sampled localities were consistent with the aridity gradient. Determining whether differences among the regions are due to local adaptation or phenotypic plasticity based on RNA sequencing expression profiles will rely on future studies that include more tissues, regional replicates, and seasonal time points.

**Author contribution statement** All the authors contributed to the study conception and design. Material preparation, data collection, and analyses were performed by LFC. The first draft of the manuscript was written by LFC, JW, and DP. All the authors commented on previous versions of the manuscript and read and approved the final manuscript.

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**Availability of data and material** The RNA reads and the de novo Transcriptome assemblies are available in NCBI via Bioproject PRJNA63711. The code used is available at [https://gitlab.com/lcorona/transcriptome\\_of\\_weeping\\_pinyon\\_pine.git](https://gitlab.com/lcorona/transcriptome_of_weeping_pinyon_pine.git).

## Declarations

**Conflict of interest** The authors declare no conflict of interest.

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## Supplementary materials

**Table S1. Geography and biogeography of sampled *P. pinceana* locations.**

Location	Condition	State	Latitude (N)	Longitude (W)	Elevation (masl)
Arboretum		Edo. México	19.493	98.848	2318
Tolantongo	SMO	Hidalgo	20.635	98.990	1900
Maguay Verde	SMO	Querétaro	21.090	99.695	2300
Nuñez	SMO	San Luis Potosí	22.671	100.483	1539
Mazapil	ChD	Zacatecas	24.599	101.472	2823
Sierra de Parras	ChD	Coahuila	25.255	101.020	2450
General Cepeda	ChD	Coahuila	25.301	102.607	1789
Palmito	ChD	Durango	25.741	104.881	2000

**Table S2. Niche modeling predictions**

Cumulative threshold	Logistic threshold	Description	Fractional predicted area	Training omission rate
1.000	0.088	Fixed cumulative value 1	0.807	0.000
5.000	0.192	Fixed cumulative value 5	0.664	0.000
10.000	0.256	Fixed cumulative value 10	0.560	0.000
48.463	0.501	Minimum training presence	0.179	0.000
48.463	0.501	10 percentile training presence	0.179	0.000
53.664	0.526	Equal training sensitivity and specificity	0.149	0.111
48.463	0.501	Maximum training sensitivity plus specificity	0.179	0.000
5.194	0.195	Balance training omission, predicted area and threshold value	0.659	0.000
7.507	0.230	Equate entropy of thresholded and original distributions	0.607	0.000

**Table S3.** Climatic characteristics by location

<b>Location</b>	<b>Isothermality</b> [°C]	<b>Annual Precipitation</b> [mm]	<b>Precipitation of the driest quarter of the year</b> [mm]	<b>Annual mean temperature</b> [°C]	<b>Water content index in the warmest quarter of the year</b> [mm/month]
Arboretum	0.69	589	21	14.9	17
Maguey Verde	0.65	891	54	16.3	33
Tolantongo	0.65	532	32	18.3	17
Nuñez	0.64	557	46	18.3	20
Mazapil	0.68	471	52	13.3	25
Sierra de Parras	0.55	432	39	18.0	19
General Cepeda	0.56	446	34	19.4	17
Palmito	0.61	526	16	17.0	17

**Table S4.** Quality statistics and sequences processed

	<b>USEARCH</b>	<b>BUSCO completeness</b>	<b>Frame selected Transdecoder</b>	<b>BUSCO completeness</b>	<b>Genes annotated</b>	<b>BUSCO completeness</b>
<b>Arboretum</b>	70446	81.3%	56340	80.8%	32429	80.8%
<b>Arboretum</b>	96923	85.4%	66398	85%	35522	85%
<b>Tolantongo</b>	71792	79.1%	53449	78.8%	29731	78.8%
<b>Maguey Verde</b>	123042	80.5%	81495	80.1%	42590	80.1%
<b>Nuñez</b>	69189	60.4%	47864	60%	27217	60%
<b>Mazapil</b>	88813	82.9%	60729	82.4%	32842	82.4%
<b>Sierra de Parras</b>	92212	78.4%	61408	77.6%	33012	77.6%
<b>General Cepeda</b>	98173	82.9%	67174	82.4%	36279	82.4%
<b>Palmito</b>	106205	77.3%	65117	75.8%	33556	75.8%

**Table S5.** Most abundant contaminants

<b>Specie</b>	<b>Tolantongo</b>	<b>Maguey Verde</b>	<b>Nuñez</b>	<b>Mazapil</b>	<b>General Cepeda</b>	<b>Sierra de Parras</b>	<b>Palmito</b>	<b>Global reference</b>
<i>Acyrtosiphon pisum</i>				24				
<i>Alternaria alternata</i> :			8					
<i>Anoplophora glabripenni</i>							4	
<i>Arthrobotrys oligospora</i>			3					
<i>Aureobasidium nanibiae</i>	14	172	25		17	38	6	334
<i>Aquifex aeolicus</i>								
<i>Bacillus subtilis</i>	46	65	33	55	57	49	51	50
<i>Baudouinia panamericana</i>		73			8	9		145
<i>Bipolaris maydis</i>		46	4					61
<i>Bipolaris oryzae</i>		39	6					89
<i>Botrytis cinerea</i>		24				11		
<i>Candida albicans</i>	5	3	4	5				
<i>Cercospora beticola</i>		38						
<i>Clavispora lusitanae</i>	9	29	13	5	19	13		163
<i>Coniosporium apollinis</i>		140						
<i>Cryptotermes secundus</i>	6			7	7			
<i>Cyphomyrmex costatus</i>						7		
<i>Dendroctonus ponderosae</i>							4	
<i>Diplodia corticola</i>		47						68
<i>Drosophila melanogaster</i>	9			21		12	8	
<i>Emericella nidulans</i>	12	63	8	14	14	20	10	101
<i>Escherichia coli</i> (strain K12):	9		6	10	14	12	12	
<i>Gibberella zeae</i>		21						
<i>Glarea lozoyensis</i>		73			13	40		143
<i>Halobacterium salinarum</i>					12	9		
<i>Halymorphia halys</i>							4	
<i>Kluyveromyces marxianus</i>	3							

<i>Metarhizium robertsii</i>																			
<i>Mycobacterium tuberculosis</i>	8					8	11			6								9	
<i>Myzus persicae</i>							24												
<i>Neosartorya fumigata</i>		30																	50
<i>Neurospora crassa</i>	9	155				22				22								8	335
<i>Nostoc sp.</i>						6				8									
<i>Onthophagus taurus</i>	10	24				4			9	15								16	
<i>Papilio polytes</i>																		5	
<i>Phaeosphaeria nodorum</i>		17																	
<i>Phialocephala scopiformis</i>																		15	
<i>Protochlamydia amoebophila</i>	9									8								7	
<i>Pseudocercospora fijiensis</i>																			
<i>Pseudocercospora ferruginea</i>									7										
<i>Rasamsonia emersonii</i>		23																	
<i>Saccharomyces cerevisiae</i>	45	201				33			38	53								43	291
<i>Schizosaccharomyces pombe</i>	80	331				70			89	105								90	418
<i>Sclerotinia sclerotiorum</i>		39								7								30	89
<i>Sugiyamaella lignohabitans</i>	12	25				11			12	15								16	
<i>Synechococcus elongatus</i>									8									8	
<i>Synechocystis sp.</i>	39	38				28			37	33								37	
<i>Trachymyrmex septentrionalis</i>	7	6								7								8	
<i>Trachymyrmex zeteki</i>	2																	4	
<i>Vollenhovia emeryi</i>	7	16				13			12									29	60
<i>Xanthobacter autotrophicus</i>							6												
<i>Xylona heveae</i>		44																	
<i>Yarrowia lipolytica</i>		20																8	58
<i>Zootermopsis nevadensis</i>						4			6										
<b>Overall</b>	<b>126</b>	<b>1012</b>				<b>153</b>	<b>157</b>		<b>225</b>	<b>315</b>				<b>178</b>	<b>1,753</b>				

**Table S7.** Statistics number of reads per sample and aligned to the reference transcriptoma for each sample

	<b>TOTAL READS</b>	<b># R1 Reads</b>	<b># R2 Reads</b>	<b>MAPPED READS</b>	<b>%</b>
<b>Arboretum 1</b>	58,224,292	29,112,146	29,112,146	50.4%	
<b>Arboretum 2</b>	78,869,060	39,434,530	39,434,530	62.36%	
<b>Nuñez</b>	66,220,808	33,110,404	33,110,404	51.16%	
<b>Mazapil</b>	60,991,584	30,495,792	30,495,792	61.42%	
<b>Maguey Verde</b>	80,176,240	40,088,120	40,088,120	60.6%	
<b>Sierra de Parras</b>	47,661,070	23,830,535	23,830,535	59.95%	
<b>General Cepeda</b>	68,413,772	34,206,886	34,206,886	59.44%	
<b>Tolantongo</b>	43,366,814	21,683,407	21,683,407	59.98%	
<b>Palmito</b>	50,070,730	25,035,365	25,035,365	56.67%	

To consult Table S6 and S8 please see: <https://doi.org/10.1007/s00468-021-02125-8>.

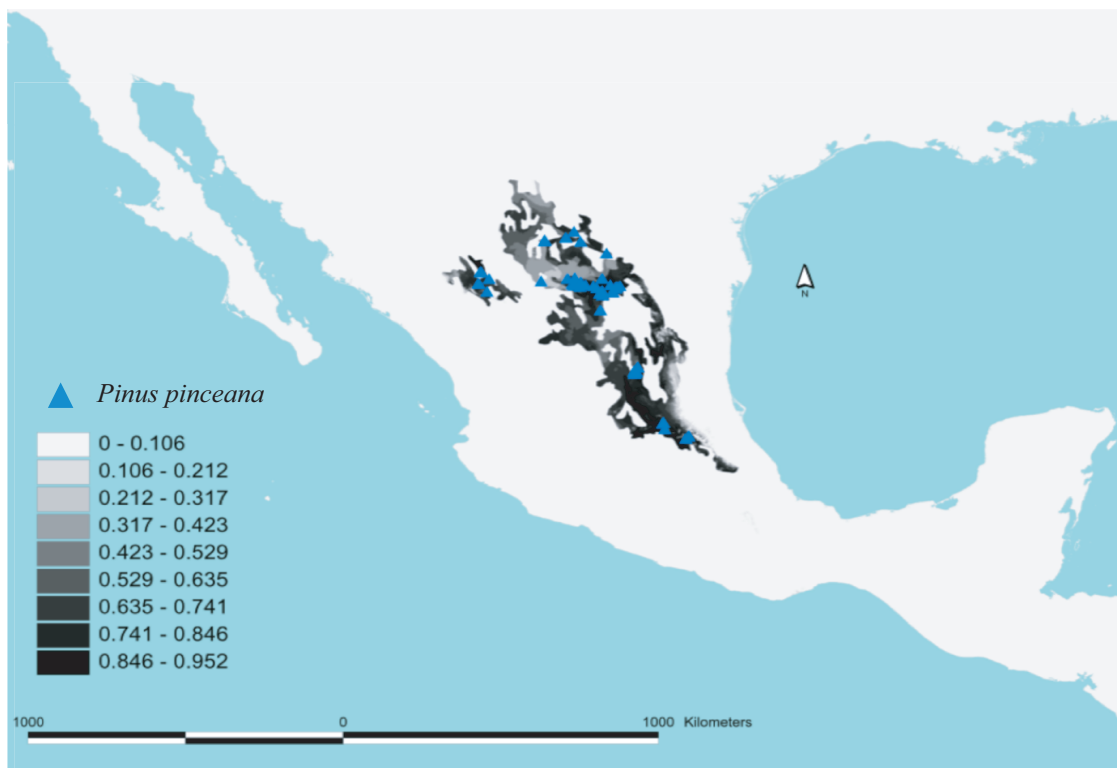


Figure S1. Potential distribution based on climatic conditions. In blue are shown the registered samples from the National Herbarium (MEXU) Black and greys show the probability of *P. pinceana* presence with respect to climatic conditions

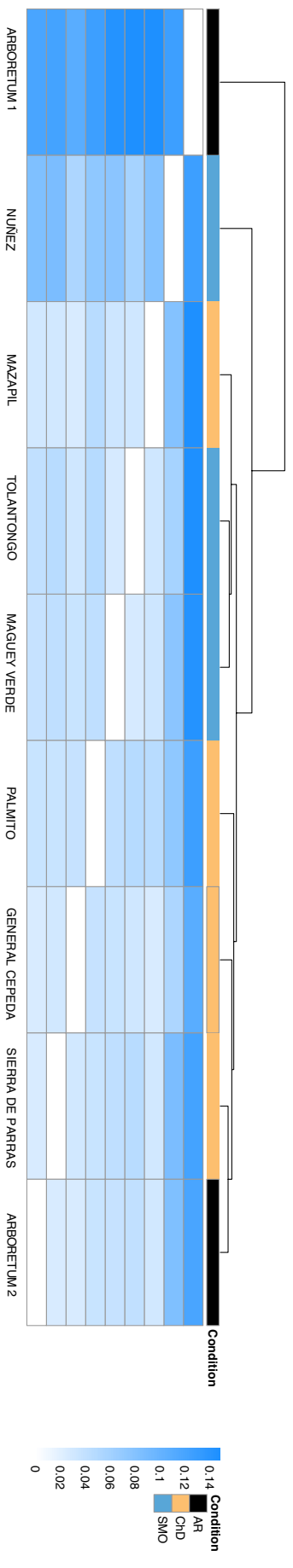


Figura S2. Sample heatmap with the comparison among levels of gene expression, between SMO and ChD.

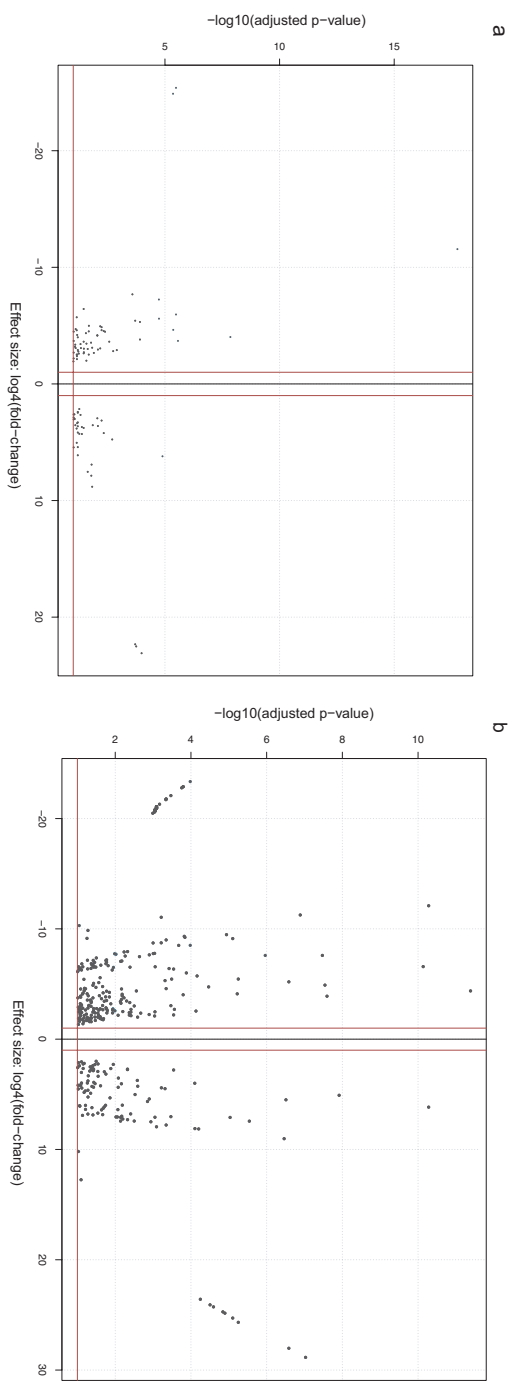


Figure S3 Volcano plot with the dispersion on the expression pattern against natural locations and arboretum samples. a. Chihuahuan Desert vs Arboretum. b. Sierra Madre Oriental vs Arboretum

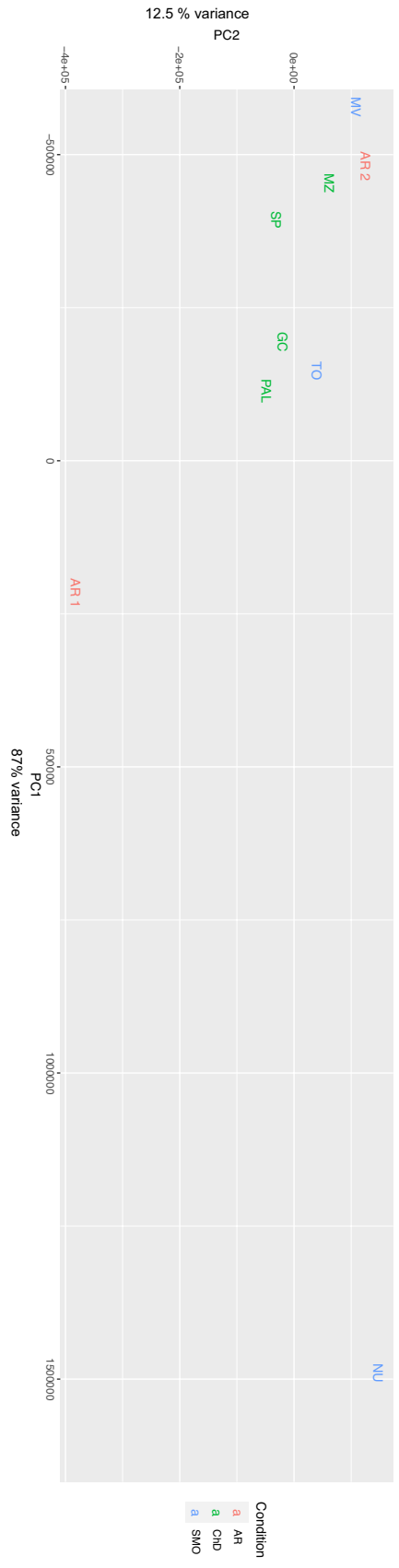


Figure S4. PCA analysis clustering of samples according to the expression pattern.



## Capítulo 2

### **Changes in demography and geographic distribution in the weeping pinyon pine (*Pinus pinceana*) during the Pleistocene**

Laura Figueroa-Corona, Alejandra Moreno-Letelier, Diego Ortega Del-Vecchyo, Pablo Peláez, David S. Gernandt, Luis E. Eguiarte, Jill Wegrzyn & Daniel Piñero

El siguiente capítulo está constituido por el artículo: Changes in demography and geographic distribution in the weeping pinyon pine (*Pinus pinceana*) during the Pleistocene. Los objetivos buscan identificar el efecto de cambios geológicos y climáticos durante el Pleistoceno a la historia demográfica y la diversidad genética de *Pinus pinceana* (Gordon & Glend.). Utilizando 4,141 SNPs obtenidos por Genotyping by sequencing (GBS) para 90 individuos adultos de diez poblaciones naturales en la distribución geográfica para inferir la demografía histórica usando un método basado en el espectro de frecuencias por sitio (fastsimcoal v2.6; fsc26) con el objetivo de contrastar posibles escenarios filogeográficos y reconstruir la divergencia de linajes. Además se reconstruyó la distribución histórica de *P. pinceana* utilizando reconstrucciones climáticas para el Holoceno medio (MH; 6 kya), Último Máximo Glaciar (LGM, 22 kya) y Último Interglaciar (LIG; 120-140 kya). Los resultados principales señalan que *Pinus pinceana* divergió en dos linajes ~ 627 kya (95% CI: 584.52 - 633.37), colonizando dos regiones: la Sierra Madre Oriental (SMO) y el Desierto Chihuahuense (ChD). En consecuencia, el aislamiento tuvo un impacto en cómo se distribuye la diversidad genética en estos dos grupos genéticos, mientras los ciclos interglaciares ocurrieron y la aridificación aumentó en el Desierto Chihuahuense con la reducción de los paleolagos. Los modelos de nicho ecológicos mostraron una mayor estabilidad del hábitat en Mazapil, Zacatecas (Central ChD), una de las poblaciones con mayor diversidad genética sugiriendo que los ciclos interglaciares, mantuvieron esta región como un refugio seguido de la progresiva aridificación del Desierto Chihuahuense.

## **Changes in demography and geographic distribution in the weeping pinyon pine (*Pinus pinceana*) during the Pleistocene**

Laura Figueroa-Corona<sup>✉1 2</sup>, Alejandra Moreno-Letelier<sup>3</sup>, Diego Ortega-Del Vecchyo<sup>4</sup>, Pablo Peláez<sup>5</sup>, David S. Gernandt<sup>6</sup>, Luis E. Eguiarte<sup>2</sup>, Jill Wegrzyn<sup>7</sup>, Daniel Piñero<sup>2</sup>

<sup>1</sup>Posgrado en Ciencias Biológicas, Universidad Nacional Autónoma de México, Ciudad de México, Mexico

<sup>2</sup>Departamento de Ecología Evolutiva, Instituto de Ecología, Universidad Nacional Autónoma de México, Ciudad de México, Mexico

<sup>3</sup>Jardín Botánico, Instituto de Biología, Universidad Nacional Autónoma de México, México, Ciudad de México, Mexico

<sup>4</sup>Laboratorio Internacional de Investigación sobre el Genoma Humano, Universidad Nacional Autónoma de México, Juriquilla, Mexico

<sup>5</sup>Centro de Ciencias Genómicas, Universidad Nacional Autónoma de México, Cuernavaca, Morelos, Mexico

<sup>6</sup>Departamento de Botánica, Instituto de Biología, Universidad Nacional Autónoma de México, Ciudad de México, Mexico

<sup>7</sup>Department of Ecology and Evolutionary Biology, University of Connecticut, Storrs, CT 06269, USA

L.F.C. laurafc25@gmail.com

A.M.L. amletelier@ib.unam.mx

D.O.D. dortega@liigh.unam.mx

P.P. ppelaez@ccg.unam.mx

D.S.G. dgernandt@ib.unam.mx

L.E.F. fruns@unam.mx

J.W. jill.wegrzyn@uconn.edu

D.P. pinero@unam.mx

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## **Conflict of Interest Statement**

The authors declare no conflict of interest

## **Abstract**

### **Aim**

To identify the processes underlying the demographic history and the genetic diversity of *Pinus pinceana* (Gordon & Glend.), distributed in the Chihuahuan Desert and the Sierra Madre Oriental, in the context of geological and climatic changes during the Pleistocene.

### **Location**

The Chihuahuan Desert and Sierra Madre Oriental, Mexico.

### **Methods**

We generated 4 141 SNPs from genotyping by sequencing on 90 adult individuals from ten natural populations covering the entire geographic distribution of *P. pinceana*. This resource was used to infer its population history using a site frequency spectrum-based method (fastsimcoal v2.6; fsc26) to contrast possible phylogeographic scenarios of lineage divergence. The past distribution for *P. pinceana* was studied using climate reconstructions

for the Mid-Holocene (MH; 6 kya), Last Glacial Maximum (LGM, 22 kya), and Last Interglacial (LIG; 120-140 kya).

## **Results**

*Pinus pinceana* diverged into two lineages ~627 kya (95% CI: 584.52 - 633.37), colonizing two regions: the Sierra Madre Oriental (SMO) and the Chihuahuan Desert (ChD). Consequently, isolation had an impact on how genetic diversity is distributed in these two lineages, while the interglacial cycles occurred and aridification increased in the Chihuahuan Desert while paleolakes were reduced in the area. Ecological niche models showed major habitat stability in Mazapil, Zacatecas (Central ChD), one of the most genetically diverse populations.

## **Main conclusion**

The population genomic analysis supports the divergence of two separate *P. pinceana* lineages during the climatic changes of the Pleistocene. These changes were related to the glacial-interglacial cycles, followed by the gradual aridification of the Chihuahuan desert, resulting in population isolation.

## **Keywords**

Chihuahuan Desert, Pleistocene refugia, *Pinus pinceana*, demographic expansion.

## **Introduction**

Chihuahuan Desert (ChD) communities have a long history of high variability in their spatial dimensions driven primarily by water input, which is scarce and has fluctuated over time (Zavala-Hurtado & Jiménez, 2020; Noy-Meir, 1973). Thus, the geological and climatic events of the Miocene–Pliocene, as well as the climatic cyclical changes of the Pleistocene, affected the diversity patterns of Chihuahuan Desert communities (Scheinvar et al., 2020). Indeed, several phylogeographic studies have reported dynamic scenarios that populations migrated and expanded over the Central Mexican Plateau from north to south and during the interglacial (15-20 glacial cycles in the Pleistocene) recolonized the northern Chihuahuan Desert (Van Devender & Burgess, 1985). See, for instance, the cases of *Ephedra compacta* or of *Agave lechuguilla* (Loera et al., 2017; Scheinvar et al., 2017), and others like *Pinus remota* which reduced to Pleistocene refugia in the Bolsón de Mapimí (Lanner & Van Devender, 1981).

The ChD is a high-elevation desert ranging from 600 to 1 675 masl and receives more rain than other deserts (235 mm of mean annual precipitation; National Park Service, <https://www.nps.gov/im/chdn/ecoregion.htm>). ChD is the third-largest desert in the American continent, and it is considered the most diverse in the Western Hemisphere, and one of the most diverse arid regions in the world (Beck & Gibbens, 1999). It also has both a high species diversity and richness of endemism (Rzedowski & Calderón de Rzedowski, 1993). ChD includes different endorheic basins, including those of the Nazas, Aguanaval, and Casas Grandes Rivers. It also possesses bolsons or endorheic desertic valleys, like the Bolsón de Mapimí and El Salado. The ChD landscape maintains different vegetation types, such as grass steppes, xeric shrubs in the intermountain plains and valleys, and open woodlands at higher elevations (Van Devender & Burgess, 1985; Czaja et al., 2014).

The Sierra Madre Oriental (SMO) is a high mountain range (1 480 - 3 720 masl) formed by a mountain chain dated to the Late Cretaceous and early Tertiary Laramide (80 – 40 Ma) formed by the deformation of Mesozoic rocks (Eguiluz de Antuñano et al., 2000). With a high climatic diversity due to its complex physiographic heterogeneity and meteorological phenomena (Hernández-Cerda & Carrasco-Anaya, 2004), SMO includes the Sabinas and Pánuco Basins, and receives moisture from the Gulf of Mexico; while the eastern slopes have tropical and temperate forests, the western slope is much drier with xeric scrubs and pine-oak-juniper woodlands (Rzedowski, 1978; Morrone et al., 2017).

The ChD biota has experienced climate fluctuations throughout geologic time. Some of these fluctuations occurred during the Late Quaternary. Paleoecological reconstructions suggest that a woodland corridor covered the area between the Sierra Madre Oriental and the Sierra Madre Occidental in Mexico (Lanner & Van Devender, 1981; Metcalfe, 2006). The Late Wisconsin vegetation that covered most of the ChD during the Pleistocene was composed of pinyon pines, junipers, and oaks, and started to decrease 11 kya (Lanner & Van Devender, 1981; Van Devender & Burgess, 1985; Elias & Van Devender, 1990). One of several reductions in pinyon-juniper and oak woodland occurred during the last 2.5M years (Pleistocene), which included intense environmental and climatic changes. During the Pleistocene, 11 climatic cycles of growth and reduction of the polar ice cap occurred in North America, affecting the global climatic conditions and consequently the composition of species associated with this woodland corridor (Lanner & Van Devender, 1981). A drastic

climate change during the Holocene turned this corridor into a warmer and drier area, starting with the reduction of the paleolakes formed during the last glaciation (~115-117 kya) and culminating with the present-day aridification, resulting in a shift to xeric vegetation (Ortega-Ramirez et al., 2004; Castiglia & Fawcett, 2006; Czaja et al., 2014).

*Pinus pinceana* is an endangered conifer (SEMARNAT 059-2010; [https://www.profepa.gob.mx/innovaportal/file/435/1/nom\\_059\\_semarnat\\_2010.pdf](https://www.profepa.gob.mx/innovaportal/file/435/1/nom_059_semarnat_2010.pdf)) that inhabits rocky soils with conditions of extreme aridity. It is locally restricted to slopes of the ChD and the western slopes of the SMO (1 480 - 3 000 masl; Farjon et al., 1997). Previous population genetic studies have shown that *P. pinceana* has a high diversity within populations and high differentiation between populations (Escalante, 2001; Ledig et al., 2001; Molina-Freaner et al., 2001).

This study focuses on how phylogeography, historical demography, and historical migration have influenced the current genetic variation and its geographic distribution of *P. pinceana*. We used genome-wide single nucleotide polymorphisms (SNPs) data to apply a coalescent-based simulation framework to infer its past demographic history during the expansion over the ChD and the SMO. We also tested the hypothesis that intraspecific gene flow may have occurred due to increased geographical contact between the two lineages during the last 2 Ma.

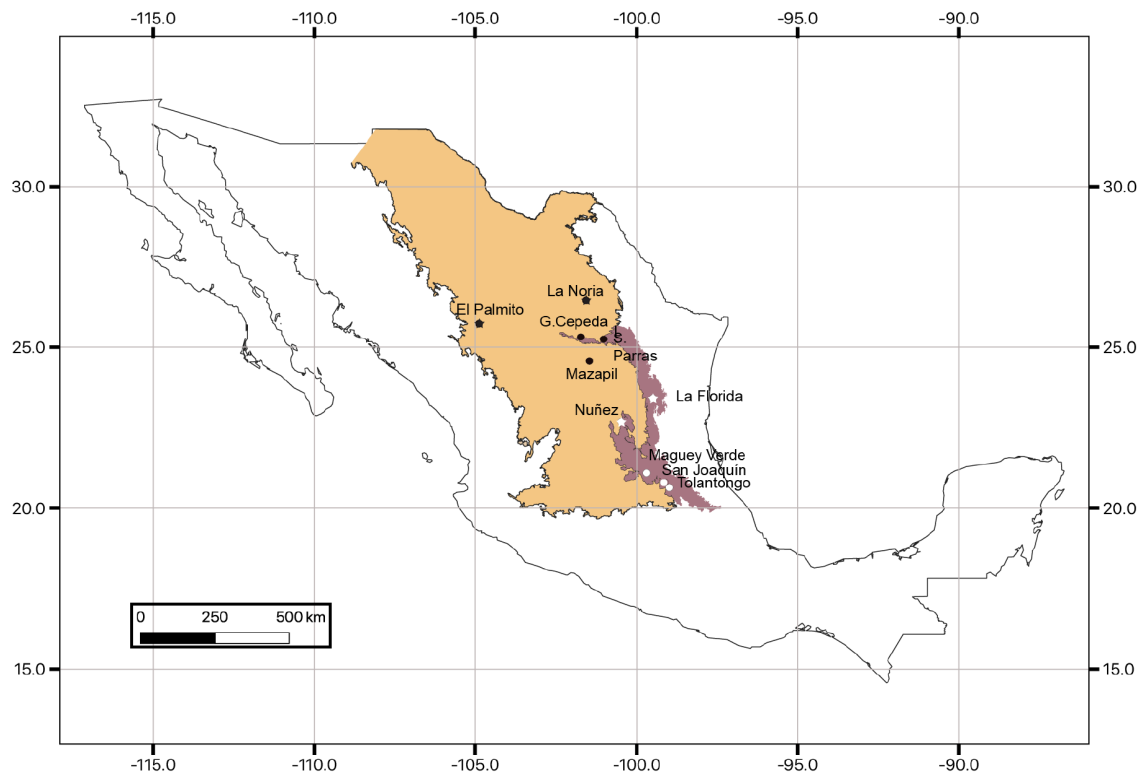
## Methods

### Sampling and genetic diversity characterization

Needles from a total of 90 healthy adult trees were arbitrarily chosen from ten populations distributed across the entire native range of *P. pinceana*. Figure 1 shows the regionalization in biogeographic provinces following Morrone et al. (2017). DNA was extracted using a modified CTAB protocol (Doyle & Doyle, 1987). Samples with a DNA concentration over 100 ng/ml were double-digested with the restriction enzymes *Pst*I / *Msp*I and fragments were single-end sequenced in 26 cycles with an Illumina Hi-Seq 2500 at the Wisconsin Biotechnology Center, Illinois.

The read adapters were removed using TRIMMOMATIC (v0.36; Bolger, 2014), and sequences with a minimum length of 80 bp in and a minimum Phred quality score of 30, were retained. The procedure was visualized with FASTQC (v0.11.7;

<https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>) and MULTIQC (v1.9; Ewels, 2016).



**Figure 1.** Geographic locations of 10 populations of *Pinus pinceana* involved in this study. The biogeographic province that encompasses the Chihuahuan Desert is located inside the sand color area, and the sampled populations from this province are indicated by a black dot, black stars correspond to Chw. The Sierra Madre Oriental is located inside the purple area and the sampled populations from this area are show with white dots, white stars correspond to SMO<sub>n</sub>.

High-quality reads were mapped to the *Pinus lambertiana* genome assembly (Stevens et al., 2016; [v1.5](#)) using the BWA-MEM algorithm (v0.7.10; Li et al., 2009). Unmapped reads were discarded. The MarkDuplicates feature from the Picard package (<http://broadinstitute.github.io/picard/>) was used to correct for artifacts of PCR duplication. Finally, we used HaplotypeCaller and VariantFiltration tools from the Genome Analysis Toolkit (GATK v3.7; McKenna et al., 2010) to filter SNPs by quality-by-depth (QD 2.0), mapping quality (MQ 40.0), Fisher strand bias test (to determine differences in the number



of sequences that support the reference and alternate alleles on each strand;  $FS > 60.0$ ), rank sum test for evaluation of the mapping qualities supporting the reference or alternate alleles ( $MQRankSum < -12.5$ ), rank sum test for assessment of bias in the position of alleles within the sequences ( $ReadPosRankSum < -8.0$ ), and strand bias between forward and reverse strands by the symmetric odds ratio test ( $SOR > 4.0$ ).

We obtained a set of biallelic SNPs that were filtered prior to analysis using VCFtools (v0.1.16; Danecek et al., 2011). The filtering parameters to retain biallelic SNPs had a minimum read depth of 15, and a maximum missing data of 0.75. We expected to get a better representation of singletons and common alleles controlling the proportion of missing data to reduce the stochastic variation (mutation) incorporated during the library construction and the sequencing process to obtain more confidence in the accuracy of the variant call. Finally, to avoid putative paralogs in both arrays, we used HDplot (v0.5-7; McKinney et al., 2017) to filter polymorphisms with  $H$  (heterozygosity) larger than 0.6 and  $D$  (deviation from even read-ratios in heterozygous) outside the range of -10 and +10. These parameters were chosen according to the proportion of heterozygous individuals within a population and allelic ratios within heterozygous individuals. We retained a total of 4 141 SNPs.

We estimated the genetic diversity for the ten populations using observed heterozygosity ( $H_O$ ), gene diversity ( $H_E$ ), and the inbreeding coefficient ( $F_{IS}$ ) using the  $R$  package *Hierfstat* (Goudet, 2005). Additionally, DNAsp (v6; Rozas et al., 2019) was used to calculate allelic richness, per site nucleotide diversity ( $\pi$ ), and pairwise population differentiation ( $F_{ST}$ ; Weir & Cockerham, 1984).

### ***Phylogeographic and population structure***

The Admixture (v1.3.0; Alexander & Lange, 2011) was used to estimate the population structure among individuals for  $K$  values from 2 to 10 admixture proportions. Genotypes were converted to ordinary PLINK files (.ped) using PLINK (v1.9; Purcell et al., 2007).

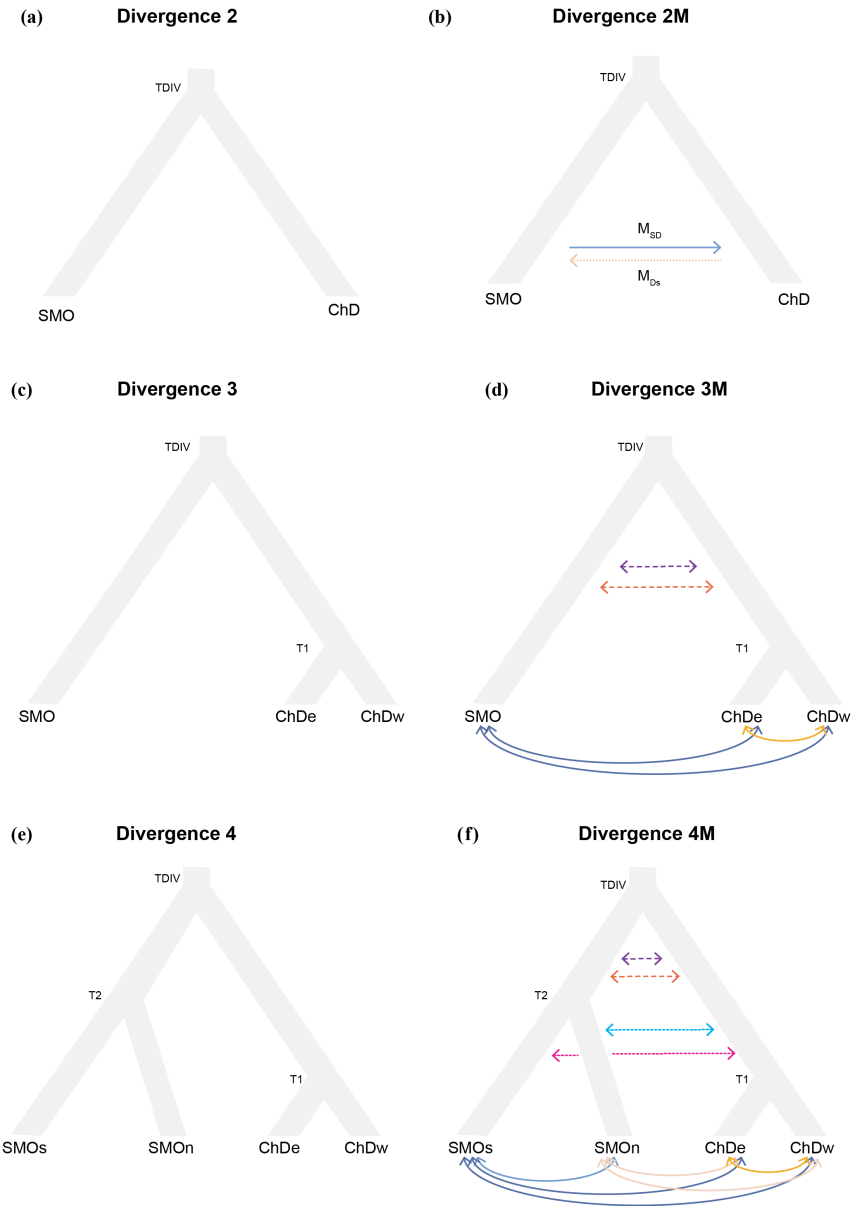
To estimate the clustering among samples without any prior for geographic provenance, we ran a discriminant analysis of principal components (DAPC) with the  $r$  package *Adegenet* (Jombart & Ahmed, 2011).

### ***Hypotheses of demographic history***

Demographic history of *P. pinceana* was inferred by testing six different scenarios via the composite likelihood approach of fastsimcoal (v2.6; Excoffier & Foll, 2011; Excoffier et al., 2013) using the site (allele) frequency spectrum (SFS) of 4 141 SNPs filtered without bias of frequency computed by *easySFS.py* (Figure S1; <https://github.com/isaacovercast/easySFS>) to infer the divergence times between and within biogeographic regions (Figure 2).

Following the clustering of DAPC our six scenarios modeled were: Divergence 1, to estimate divergence times between ChD and SMO populations ( $T_{DIV}$ ), and effective population sizes ( $N_e$ ). Divergence 2, to estimate the divergence times between the ChD ( $T_1$ ) in the northwest El Palmito and La Noria populations (ChDw), followed by the divergence between ChD and SMO populations ( $T_{DIV}$ ). Finally, Divergence 3 followed the clustering of DAPC to estimate the divergence time between and within regions as colonization or fragmentation events, within the ChD ( $T_1$ ) in the northwest El Palmito and La Noria populations (ChDw) and within SMO ( $T_2$ ) in the northern Nuñez and La Florida populations (SMOn), after the divergence event between ChD and SMO populations ( $T_{DIV}$ ). These three scenarios were redrawn with the same parameters ranges, adding asymmetrical migration rates among groups to construct the migration versions 1M, 2M, and 3M.

For each scenario, we estimated divergence times converted from generations to years using a generation time of 100 years. This generation time represents an approximation of the average age of reproductive individuals in a population, or the generation time for overlapping generations for this species (Franco & Silvertown, 2004). The mutation rate was set at  $7.28 \times 10^{-10}$  per base pair per generation as reported for *Pinus* (De la Torre et al., 2015). Thus, settings considered that individuals of *P. pinceana* have extended lifespans and a delayed reproduction (Pavek, 1994; Villanueva-Díaz et al., 2010).



**Figure 2.** Modeled demographic histories for *Pinus pinceana*. (a) Divergence 1, model draw for the divergence between SMO and ChD. (b) Divergence 1M including asymmetrical and bidirectional migration among regions. (c) Divergence 2, model draw for the divergence within the ChD followed by divergence between SMO and ChD. (d) Divergence 2M including asymmetrical and bidirectional migration among regions. (e) Divergence 3 model draw including fragmentation events in ChD to the NW and SMO to the SMO<sub>n</sub>. (f) Divergence 3M including asymmetrical and bidirectional migration among every region.

Divergence 1 with no-migration model was set using search ranges for the effective population sizes ( $N_e$ ) from 100 to 10 000 individuals in the SMO and ChD lineages, and a  $T_{DIV}$  between 5 00 and 9 000 generations ago. Divergence 2 was set using search ranges for the effective population sizes ( $N_e$ ) the SMO and ChD from 100 to 10 000 individuals, and from 70 to 1 000 individuals for ChDw.  $T_1$  was set to date the divergence event within the ChD at between 100 and 1 000 generations ago and  $T_2$  was set to date the divergence event between ChD and SMO with the range of 5 000 and 10 000 generations ago. Divergence 3 no-migration  $N_e$  of the northwestern Chihuahuan Desert (NW) and the SMO region was set from 10 to 900 individuals and between 1 000 and 10 000 individuals for the ChDe and SMOs. For date  $T_1$  the divergence event within the ChD was set from 50 to 1 000 generations, for  $T_2$  the divergence event within the SMO from 1 000 to 2 000 generations ago, and for  $T_3$  date the divergence event between ChD and SMO from 5 000 to 10 000 generations ago. Finally, for migration models, we used the same range adding between  $1 \times 10^{-9}$  to  $1 \times 10^{-4}$ , per base per generation, drawn from a log uniform prior distribution.

For each of the six models, we simulated 100 replicates, 150 000 iterations, and 30 ECM cycles. To select the best scenario, we estimated the Akaike Information Criterion AIC value (Akaike, 1974) using the maximum composite likelihood estimated from the best replicate and compared the models using the weighted Akaike information criterion (wAIC). Finally, we calculated confidence intervals of parameter estimates from 100 parametric bootstrap replicates by simulating SFS from the maximum composite likelihood estimates and re-estimating the parameters each time (Excoffier et al., 2013).

### ***Ecological niche modeling and refugia hypotheses***

We compiled 37 records verified from previously published results and from specimen records in the National Herbarium of Mexico (MEXU). Each presence point was separated by at least 5 km from other presence points, to avoid spatial autocorrelation. We extracted the values for 19 bioclimatic variables with a resolution of 30 arc-seconds from WorldClim (Fick et al., 2017) using QGIS (v3.10; <https://qgis.org/es/site/>) and performed an outlier analysis using Mahalanobis distances using the package JMP (v9; SAS Institute Inc., Cary, NC, 2010).

To reduce model overfitting, we selected independent bioclimatic variables before constructing the ENMs with a Principal Component Analysis using JMP (v9; SAS Institute Inc., Cary, NC, 2010). We retained Annual Mean Temperature (Bio1), Isothermality (Bio3), Temperature Seasonality (Bio4), Annual Precipitation (Bio12), and Precipitation in the Driest Month (Bio14) from PC1; Precipitation Seasonality (Bio15), and Precipitation of Warmest Quarter (Bio18) from PC2; and Minimum Temperature of Coldest Month (Bio6) from PC3.

To construct a potential niche model, we used Maxent (v3.3.3e; Phillips & Dudík, 2008) with linear and quadratic features, a regularization multiplier of 1, 30% of random test percentage, clamping, and 10 replicates. We performed projections to three points in the past: Mid-Holocene (MH; 6 kya), Last Glacial Maximum (LGM, 22 kya), and Last Interglacial (120-140 kya; Otto-Bliesner et al., 2008). For the Mid-Holocene (MH) and LGM, we used CCSM4 and MIROC-ESM World climate models. We processed the resulting average models by removing the lower 10<sup>th</sup> percentile of probability values to eliminate areas of over prediction. All raster processing was performed with the package *raster* in RStudio (Hijmans & van Etten, 2012). Areas were estimated by counting pixels with the package *raster*, where each pixel corresponds to approximately 1 km<sup>2</sup>.

## Results

### *Sequencing and genotype characterization*

The single-end sequences produced an average of 2.8 M for each individual, which was reduced to 2.54 M after quality control. The datasets obtained with GATK produced 613 651 variants. VCFtools was used to obtain the final processed data with 4 141 biallelic SNPs, with a moderate amount of missing data (mean=9.89%), and a mean depth per site of 61.7×.

Genetic diversity was estimated for the ten populations (Table 1). Globally, the observed heterozygosity ( $H_O$ ) was 0.19, the expected heterozygosity ( $H_E$ ) was 0.1231, and the inbreeding coefficient ( $F_{IS}$ ) was -0.5653. Levels of diversity ( $H_E$ ) were similar among populations and ranged from 0.103 (Maguey Verde, SMO) to 0.1354 (General Cepeda; ChD). Levels of  $H_O$  were highest for the ChD location, General Cepeda (0.2166), and the lowest for Maguey Verde (0.16) in SMO. Estimates for the inbreeding coefficient  $F_{IS}$  were negative for all populations, ranging from -0.5412 (Maguey Verde; SMO) to -0.6058 (La

Florida; SMO), while the highest nucleotide diversity was found in General Cepeda (0.0116), and the lowest in Maguey Verde (0.0083).

**Table 1.** Measures of genetic diversity for 90 individuals of *P. pinceana* from ten populations calculated from 4 141 single nucleotide polymorphism loci.

	Population	$H_E$	$H_o$	$F_{IS}$	$\pi$	Tajima D	$\theta_w$
	Tolantongo	0.1127	0.1790	-0.5456	0.0104	-0.0550 <sub>NS</sub>	0.1333
	San Joaquín	0.1111	0.1811	-0.5865	0.0108	0.0782 <sub>NS</sub>	0.1400
	Maguey Verde	0.1033	0.1640	-0.5412	0.0083	-0.0345 <sub>NS</sub>	0.1140
SMO	Nuñez	0.1166	0.1907	-0.5947	0.0113	0.0923 <sub>NS</sub>	0.1448
	La Florida	0.1130	0.1843	-0.6058	0.0099	0.1307 <sub>NS</sub>	0.1445
	El Palmito	0.1166	0.1849	-0.5927	0.0097	0.0247 <sub>NS</sub>	0.1408
	Mazapil	0.1269	0.2024	-0.5475	0.0101	0.0303 <sub>NS</sub>	0.1566
	La Noria	0.1291	0.2069	-0.5551	0.0109	0.0861 <sub>NS</sub>	0.1615
	G. Cepeda	0.1354	0.2166	-0.5522	0.0116	0.1444 <sub>NS</sub>	0.1686
ChD	Sierra Parras	0.1324	0.2144	-0.5759	0.0115	0.1746 <sub>NS</sub>	0.1702
	<b>Global</b>	0.1231	0.1900	-0.5653	0.0074	-0.5203 <sub>NS</sub>	0.1535

$H_o$ , observed heterozygosity;  $H_E$ , expected heterozygosity within populations;  $F_{IS}$ , inbreeding coefficient;  $\pi$ , nucleotide diversity

The average  $F_{IS}$  by locus was -0.195 while the median was -0.03. This difference is due to a biased average due to high frequency of loci with a high proportion of heterozygous SNPs. Table S1 shows the calculation for the mean and median values on  $F_{IS}$  for different sets after deleting the extreme values ( $F_{IS} < -0.799$ ). This changes the average  $F_{IS}$  estimates (using the remaining 3 762 SNPs) to an observed heterozygosity ( $H_o$ ) of 0.093, an expected heterozygosity ( $H_E$ ) equal to 0.116, and an average inbreeding coefficient ( $F_{IS}$ ) of -0.35. This effect was observed in all populations (Table S2).

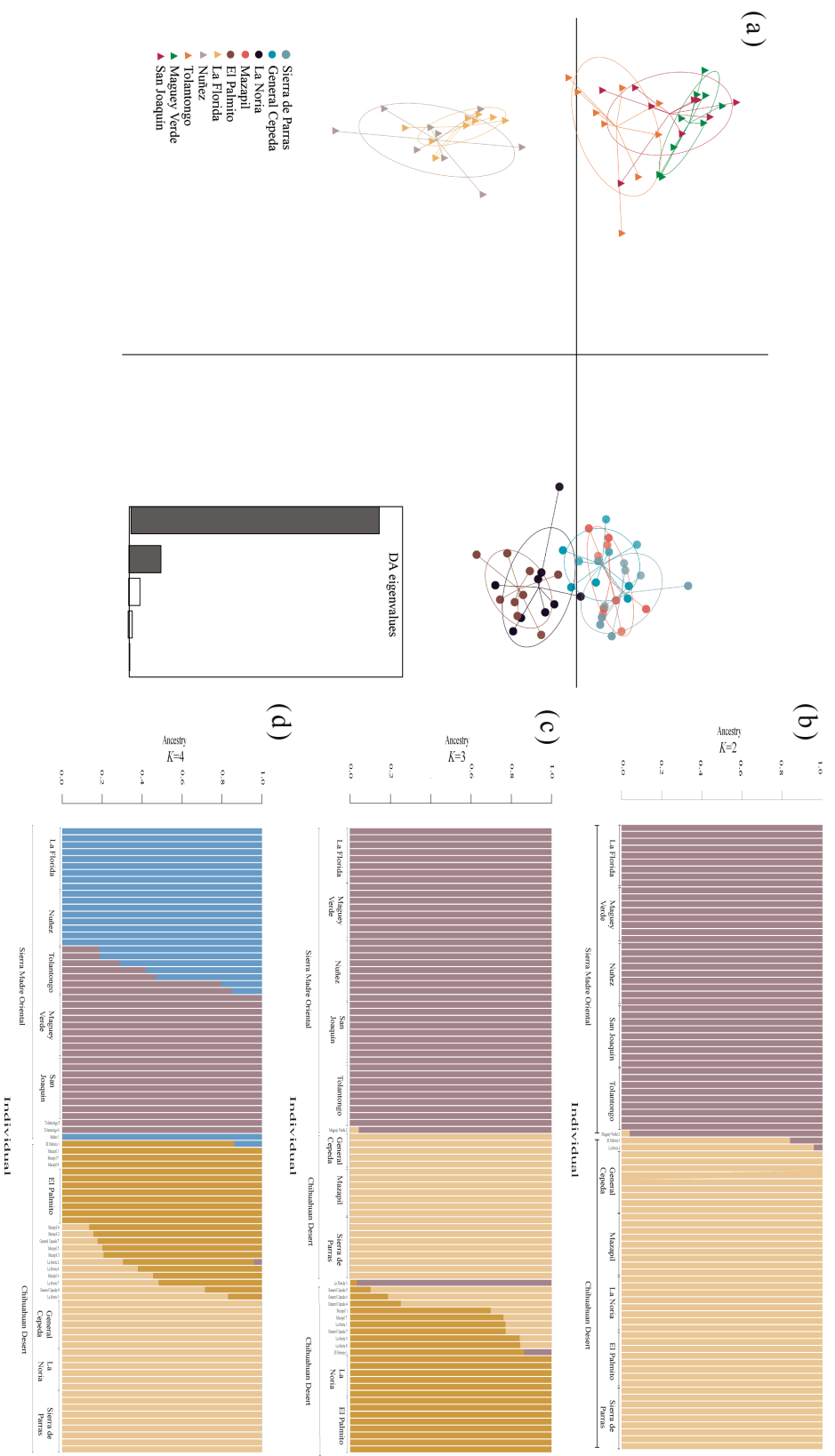
### ***Genetic structure and phylogeography pattern***

The global  $F_{ST}$  value was 0.0612 (Table 2). Within SMO, the populations had a mean  $F_{ST}$  = 0.012, compared to 0.010 within the ChD populations. Between biogeographic regions the mean value was 0.055.

The discriminant analysis of principal components (DAPC) showed two clear population clusters divided by the PC1 describing 14.9% of the total variance, and sub-clustering within the biogeographically regions showed by PC 2 that describe 4.09% of the total variance, where La Noria and El Palmito had the greatest divergence within the ChD populations and Nuñez and La Florida show the largest divergence within the SMO (Figure 3A). Admixture analysis also showed that  $K=2$  supports the clustering in two biogeographic areas. For  $K=2$ , 42 of the 44 samples from SMO were classified 100% in one cluster (Figure 3B) and 43 samples from ChD in the second cluster. Three samples were found with a level of admixture between both groups composed of individuals from Maguey Verde and northwest populations El Palmito and La Noria. The second-best supported clustering was found for  $K=3$  denoting a longitudinal gradient of admixture in the ChD samples from the northwest to the center populations (Figure 3C). Finally, for  $K=4$  (Figure 3D) the two biogeographical regions showed substructure between the NW localities El Palmito and La Noria and the rest of the ChD and between the SMO with Nuñez and La Florida and the south of the SMO.

**Table 2.** Pairwise differentiation ( $F_{ST}$ ) among the ten populations sampled of *P. pinceana* with 4 141 SNPs

SMO	Tolantongo									
	San Joaquín	-0.0012								
	Maguey Verde	-0.0004	-0.0006							
	Nuñez	0.0022	0.0084	0.0101						
	La Florida	0.0018	0.0093	0.0104	0.0008					
ChD	El Palmito	0.0331	0.0373	0.0362	0.0351	0.0358				
	Mazapil	0.0377	0.0388	0.0445	0.0424	0.0418	0.0074			
	La Noria	0.0402	0.0418	0.0469	0.0440	0.0445	-0.0007	0.0007		
	G. Cepeda	0.0410	0.0451	0.0496	0.0477	0.0459	0.0087	-0.0012	-0.0040	
	Sierra Parras	0.0469	0.0458	0.0527	0.0516	0.0501	0.0101	-0.0025	0.0016	-0.0057



**Figure 3.** Population structure and nucleotide diversity based on 4 141 genome-wide SNPs in *Pinus pineana*. (a) Scatterplots resulting from discriminant analysis of principal components (DAPC) for all 90 samples, and the eigenvalues of discriminant functions portrayed in the bottom right. (b) Admixture for  $K=2$ , (c) for  $K=3$  and (d) for  $K=4$ .



### ***Demographic history***

We used the entire 4 141 SNP dataset although it could include paralogs since there was no filter by frequency for heterozygous alleles. The population divergence time hypotheses from the demographic simulations for the six scenarios resulted in divergence time estimated during the Middle Pleistocene (Table 3). The best-supported model was the Divergence 1M according to the  $\Delta$ AIC and considering the split of the two genetic clusters. The dated divergence ( $T_{DIV}$ ) between them was estimated in the Middle Pleistocene (627 kya; 95% CI: 584.52 - 633.37; Table 4). Effective population sizes ( $N_e$ ) for groups were similar between groups, SMO with 6 711 individuals (95% CI: 6 477.24 – 6 763.9), and ChD with 6 639 individuals (95% CI: 6 422 – 6 689). Migration rates per generation ( $m$ ) were estimated to be significantly higher  $1.03 \times 10^{-5}$  (95% CI:  $9.09 \times 10^{-6}$  -  $1.07 \times 10^{-5}$ ) from SMO to ChD, than in the opposite direction ( $1.61 \times 10^{-6}$ ; 95% CI:  $1.04 \times 10^{-6}$  -  $1.83 \times 10^{-6}$ ; Table 5). The sub-fragmentation according to the Divergence 3 and 3M occurred during the interglacial cycles and the Middle Pleistocene (~127.7 - ~539.2 kya).

**Table 3.** Comparison of demographic models analyzed with fastsimcoal

<b>Model</b>	<b><i>k</i></b>	<b>AIC</b>	<b><math>\Delta</math>AIC</b>
Divergence 1	3	40 347.47	1074.42
Divergence 1M	5	39 273.05	0
Divergence 2	5	45 847.82	6 574.77
Divergence 2M	13	39 768.40	495.35
Divergence 3	7	41 405.14	2 132.09
Divergence 3M	20	39 983.50	710.44

**Table 4.** Parameters inferred for time divergence and effective population sizes from coalescent simulations with fastsimcoal under the demographic models

	<i>SMO</i>		<i>ChD</i>		<i>T<sub>DIV</sub></i>		
<b>Divergence 1</b>	6 739		6 640		627 300		
<b>Divergence 1M</b>	6 711		6 639		627 000		
	<i>SMO</i>		<i>ChD<sub>e</sub></i>	<i>ChD<sub>w</sub></i>		<i>T<sub>1</sub></i>	<i>T<sub>DIV</sub></i>
<b>Divergence 2</b>	57 186		57 227	5 870		1 840 800	3 483 100
<b>Divergence 2M</b>	44 836		22 880	3 784		1 208 500	2 667 900
	<i>SMO<sub>s</sub></i>	<i>SMO<sub>n</sub></i>	<i>ChD<sub>e</sub></i>	<i>ChD<sub>w</sub></i>	<i>T<sub>1</sub></i>	<i>T<sub>2</sub></i>	<i>T<sub>DIV</sub></i>
<b>Divergence 3</b>	7 831	268	25 322	2 191	223 900	127 700	572 000
<b>Divergence 3M</b>	22 872	472	7 619	1 564	539 700	539 200	541 600

Population names refers to the  $N_e$  number of individuals in the different groups;  $T_1$ ,  $T_2$ , and  $T_{DIV}$  are in years; SMO: Sierra Madre Oriental, SMOs: south SMO Tolantongo, San Joaquín and Maguey Verde populations, SMO<sub>n</sub>: northern SMO Nuñez and La Florida populations, ChD: Chihuahuan Desert, ChD<sub>e</sub>: east ChD Mazapil, General Cepeda and Sierra de Parras, ChD<sub>w</sub>: northwest ChD El Palmito and La Noria populations.

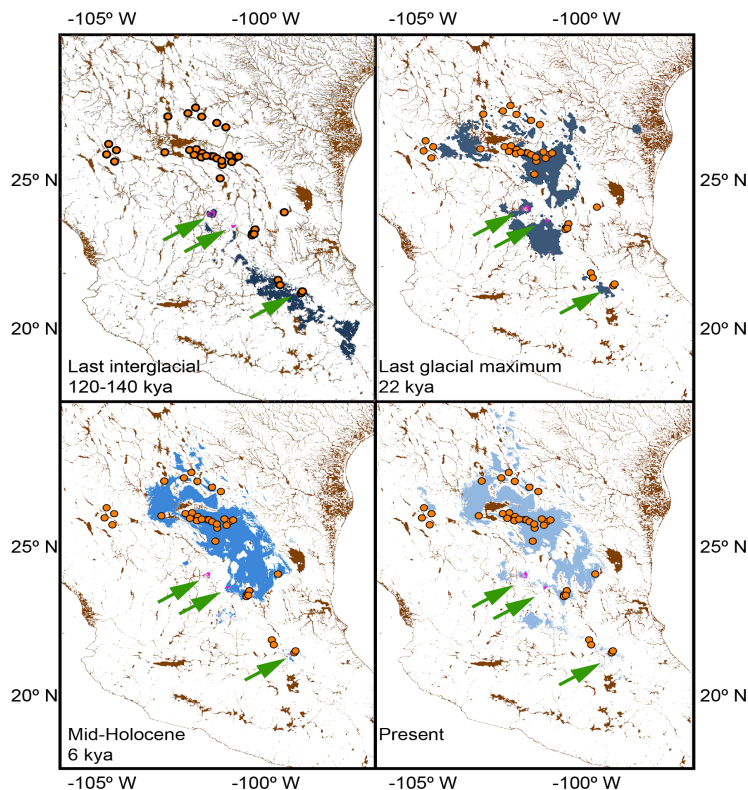
**Table 5.** Migration rates inferred from coalescent simulations with fastsimcoal under the demographic models

<b>Divergence 1M</b>		SMO	SMO	ChD		
		ChD		1.61 x10 <sup>-6</sup>		
<b>Divergence 2M</b>	<i>T<sub>1</sub></i>		SMO	ChDe	ChDw	
		SMO		3.44 x10 <sup>-7</sup>	1.69 x10 <sup>-8</sup>	
		ChDe	7.72 x10 <sup>-5</sup>		5.19 x10 <sup>-8</sup>	
		ChDw	1.58 x10 <sup>-5</sup>	1.91 x10 <sup>-4</sup>		
	<i>T<sub>2</sub></i>		SMO	ChD		
		SMO		7.35 x10 <sup>-8</sup>		
		ChD	1.45 x10 <sup>-8</sup>			
<b>Divergence 3M</b>	<i>T<sub>1</sub></i>		SMO <sub>s</sub>	SMO <sub>n</sub>	ChD <sub>w</sub>	ChD <sub>e</sub>
		SMO <sub>s</sub>		1.80 x10 <sup>-6</sup>	1.03 x10 <sup>-8</sup>	5.46 x10 <sup>-9</sup>
		SMO <sub>n</sub>	1.3x10 <sup>-8</sup>		2.52 x10 <sup>-7</sup>	2.90 x10 <sup>-7</sup>
		ChD <sub>w</sub>	7.74x10 <sup>-9</sup>	6.58 x10 <sup>-5</sup>		5.08 x10 <sup>-6</sup>
		ChD <sub>e</sub>	4.01x10 <sup>-9</sup>	1.24 x10 <sup>-4</sup>	3.47 x10 <sup>-8</sup>	
	<i>T<sub>2</sub></i>		SMO	ChD <sub>w</sub>	ChD <sub>e</sub>	
		SMO		1.93 x10 <sup>-8</sup>	2.83 x10 <sup>-9</sup>	
		ChD <sub>w</sub>	3.50 x10 <sup>-7</sup>		2.97 x10 <sup>-8</sup>	
		ChD <sub>e</sub>	9.22 x10 <sup>-8</sup>	2.72 x10 <sup>-9</sup>		
	<i>T<sub>3</sub></i>		SMO	ChD		
SMO			8.39 x10 <sup>-6</sup>			
	ChD	3 x10 <sup>-7</sup>				

*T<sub>1</sub>*, *T<sub>2</sub>*, and *T<sub>DIV</sub>* are in years; SMO: Sierra Madre Oriental, SMO<sub>s</sub>: south SMO Tolantongo, San Joaquín and Maguey Verde populations, SMO<sub>n</sub>: northern SMO Nuñez and La Florida populations, ChD: Chihuahuan Desert, ChDe: east ChD Mazapil, General Cepeda and Sierra de Parras, ChDw: northwest ChD El Palmito and La Noria populations.

### *Historical distribution projections*

The present and past projections cover a broader area than what is observed in the current populations (Figure 4). There was a significant latitudinal range shift, as well as an area increase from the Last Interglacial (LIG) to the present. The LIG distribution shifted to the southern limit of the ChD, bordering with the Rio Pánuco biogeographical province (Scheinvar et al., 2020), and decreased by ca. 70% from the present area (116 k pixels), with 34 k pixels. The Last Glacial Maximum (LGM; 89 k pixels) and mid-Holocene (MH; 114k pixels) predicted ranges occupy a smaller area (23% and 7% smaller than the current distribution, respectively), but close to areas in the current range. Flatlands and basins seem to be a barrier in the distribution of *P. pinceana*, as the areas identified as current or past water bodies have not been occupied by the species. The areas that remained stable, that is, with suitable conditions for *P. pinceana* since the LIG were quite small (2 k pixels) and mostly found in the southern ChD.



**Figure 4.** Ecological niche modeling predictions of *Pinus pinceana* current climate change scenarios (Last interglacial (LIG), Last glacial maximum (LGM), Mid-Holocene (MH), and present). Orange circles represent present localities, and blue indicates areas predicted for the species. Green arrows indicate areas that remained stable from the LIG to present.

## Discussion

### *Phylogeography*

We detected a clear geographic structure of genetic diversity of *P. pinceana* in the Chihuahuan Desert (Ortega-Ramirez et al., 2004; Castiglia & Fawcett, 2006; Czaja et al., 2014). This structure is related to climate dynamics during the Quaternary. These changes highlight the importance of two distinct processes: first, genetic drift due to reductions in effective population sizes in different areas; and second, fragmentation due to the aridification of the ChD.

The genetic diversity patterns were similar to the genetic diversity ( $H_0$ ) values described in conifer species with broad geographic ranges in Mexico like *Pinus patula*, *Pinus strobiformis*, and *Picea chihuahuana* (Peláez et al., 2020; Moreno-Letelier & Piñero, 2009; Jaramillo-Correa et al., 2006). The values previously reported for microsatellite (cpDNA) SSR loci in *P. pinceana* (Escalante, 2001; Figueroa-Corona, 2012) was substantially higher than the values found here. This is expected because microsatellites have higher mutation rates than biallelic SNPs located in functional genes (Hamblin et al., 2007). We found that four of the populations: La Noria in the northwest and Sierra de Parras, General Cepeda, and Mazapil in the ChD showed the highest levels of diversity (Table 1).

In accordance with previous reports in *P. pinceana* (Ledig et al., 2001; Molina-Freaner et al., 2001; Ledig et al., 1986) we found high observed heterozygous values. On the other hand, we surprisingly found negative  $F_{IS}$  averages. Similar situations with high heterozygosities and negative  $F_{IS}$  averages has been found in some conifers like *Cryptomeria japonica* var. *sinensis*, *Pinus patula* and *P. albicaulis* (Cai et al., 2020; Peláez et al., 2020; Liu et al., 2016).

We attribute these  $F_{IS}$  values to two artifacts, first the sub-estimation of linked loci between the SNPs giving that analyzing the most extreme values of  $F_{IS}$  (-0.8 to -1) in the 4 141 SNPs dataset we found 377 fixed heterozygous SNPs, establishing a distance based on Pavy (et al., 2012) of 100 bp at maximum between polymorphisms we obtained 276 of them linked loci in 96 linkage groups conformed by two to eleven SNPs. The second possibility is the presence of some paralogous SNPs that were not filtered with the approach of McKinney

(et al., 2017) as has been suggested in view of the complexities of conifer genomes (De la Torre et al., 2014; Leitch et al., 2019).

The differentiation of trees from the SMO and ChD was attributed to isolation by distance in a previous study based on cpDNA SSR loci (Escalante, 2001). Our results of genetic structure clearly identified two lineages, corresponding to the same two biogeographic regions. Furthermore, these lineages present relatively high levels of genetic differentiation between them ( $F_{ST} = 0.0612$ ), like that typically reported for conifer taxa with wide distribution ranges (Pelaez et al., 2020; Cobo-Simon et al., 2020; Jaramillo-Correa et al., 2020). Nevertheless, the discriminant components analysis shows that SMO and ChD groups have significant levels of admixture between them.

The structured biogeographic pattern between ChD and SMO along the Central Mexican Plateau has been reported for several desert scrub plant species from the ChD (*e.g.*, Sosa et al., 2009; Vasquez-Cruz & Sosa, 2016; Scheinvar et al., 2017). In all cases, a strong phylogeographic structure found within the SMO was correlated with geologic and paleoclimatic changes. As an example of this pattern, Vasquez-Cruz & Sosa (2016) identified a main phylogeographic pattern in species of Rosaceae where the ChD was the ancestral area, followed by contraction of suitable habitat during the Last Interglacial (~120 000 – 140 000 years), followed by an expansion during the LGM (~22 000 years) to the SMO and the Central Mexican Plateau and then recolonization of the ChD during the Mid-Holocene (~6 000 years). Loera et al. (2017) argued that this pattern between SMO and ChD regions in *Ephedra compacta* was reinforced by the lack of dispersal and changes in elevation due to biogeographic barriers within this region.

### ***Demographic history***

With a patchy but extensive geographic distribution, *P. pinceana* is not considered an endangered species by the IUCN, but it is considered endangered by Mexican law based on more detailed ecological, geographic, and genetic information (SEMARNAT 059-2010).

Demographic data can help us to apply historical context to the phylogeographic question. The current genetic differentiation patterns of the ChD xeric scrubs can be explained by an initial expansion followed by processes of isolation between lineages to the present very reduced zones of secondary contact and limited migration (Scheinvar et al.,

2017; Loera et al., 2017). The results of the six models give support consistent with changes in climate and the genetic structure founded in *P. pinceana* in the ChD during the Middle Pleistocene. We suggest that this allopatric fragmentation of two lineages, together with an expansion of the xeric scrubs and pine-oak forests, after the end of the glacial inter-cycles, was crucial in shaping the present distribution of the genetic diversity in *P. pinceana*.

Quaternary dynamics in the ChD have been described with geological and biogeographic evidence. Genetic evidence reveals different levels of diversities between the northern and southern ChD (Scheinvar et al., 2020). Genetic structure and demographic history during the Pleistocene have been found for the xeric scrubs *Agave lechuguilla*, *A. striata*, and *A. stricta* (Scheinvar et al., 2017; Trejo et al., 2016; Martínez-Ainsworth, 2013), the rodents *Thomomys umbrinus* and *Perognathus avus*, and the turtle *Kinosternon avescens* (Mathis et al., 2014; Neiswenter & Riddle, 2010; Serb et al., 2001). The pattern of differentiation between ChD and SMO has been found in *Ephedra compacta*, where the regionalization of the biogeographic zones in SMO and ChD matches with genetic structure (Loera et al., 2017).

The estimated migration rates show an asymmetric flux between regions (Table 5). This is probably due to the current distribution, which suggests that distance works as barrier for the extreme's population. Nevertheless, we still cannot establish a cause for the percentage of admixture in individuals from San Joaquín and El Palmito despite the low connectivity with nearby populations.

The separation of the northwestern ChD in the DAPC and the  $K=3$ , represents a later, more recent fragmentation from the ChD populations (Figure 3). The western populations are at the very edge of the distribution, which suggests that Nazas basin increase the isolation and restricted gene flow. This pattern of West/East divergence has been described for Chihuahuan Desert xeric scrubs like *Astrophytum* spp., *Berberis trifoliata*, *Agave victoria-reginae*, *Leucophyllum* spp., *Ephedra compacta* (Vázquez-Lobo et al., 2015; Angulo et al., 2017; Gándara & Sosa, 2014; Loera et al., 2017), and in the snake *Crotalus molossus* (Anderson & Greenbaum, 2012).

### ***Genetic diversity in a temporal context***

Phylogenetic divergence of *Pinus* subsection *Cembroides* occurred during the Miocene (11 Mya; Gernandt et al., 2008; Saladin et al., 2017; Jin et al., 2021). The fossil evidence for pinyon pines records large changes in the distribution range during the Pliocene along the Central Mexican Plateau (Lanner & Van Devender, 1981; Van Devender & Burgess, 1985). Conifers had a wider distribution during the Pleistocene in Mexico in the ChD, and a complex dynamic around the Holocene that diminished or extremely reduced their populations (Quiñonez-Perez et al., 2017; Jaramillo-Correa et al., 2006).

The high frequency of singletons and high diversity within populations of *P. pinceana* in this highly fragmented habitat has likely occurred recently. Their long generation time and overlap of demographic cohorts are also likely to contribute to these patterns. The prevalence of new polymorphisms in all populations is maintained by relatively large effective population sizes.

Morpho-physiological studies in *P. pinceana* detected morphological differentiation patterns, where the individuals from the ChD had thicker epicuticular waxes on the needles, faster growth, and larger primary roots, and greater volume of lateral roots in Chihuahuan Desert individuals than in individuals from the Sierra Madre Oriental, differences that suggest local morphological adaptation to aridity exposure (Martíñón-Martínez et al., 2010; Córdoba-Rodríguez et al., 2011). We could not ascribe any adaptive relevance to the differentiation of the genetic clusters, in part because of the low coverage ssequencing of the gene space regions in the *Pinus* genome, and the absence of specific candidate genes, but also because the strongly structured signatures in genetic diversity produced by changes in demographic history. An independent analysis using comparative transcriptomics for this species in both biogeographic regions detected a possible plastic response to biotic and abiotic factors present in the populations (Figueroa-Corona et al., 2021).

### ***Historical distribution***

Based on our analyses, we propose that *P. pinceana* differentiated in two lineages in the Middle Pleistocene (~627 kya; 95% CI: 584.52 - 633.37), followed by the division of both biogeographic regions (~127.7 - ~539.2 kya) during the interglacial cycles. Over time the climatic conditions in the habitat of *P. pinceana* became more common, in contrast to *Picea*



*chihuahuana*, which has become more restricted (Quiñonez-Perez et al., 2017; Jaramillo-Correa et al., 2006). The recent climatic changes of the ChD, in particular the increment in aridification, and the reduction of water bodies (Czaja et al., 2014) promoted the geographic expansion of the species, and the phylogeographic patterns obtained by our data, involving a gradual expansion over the ChD since the LGM. This pattern of expansion has been described in the ChD in other scrub species like *A. lechuguilla* and *E. compacta* (Scheinvar et al., 2017; Loera et al., 2017). Throughout the Mid-Holocene there was more connectivity across regions, while the Nazas Basin determined the division within the ChD, producing the separation between the northwestern and central ChD despite the lack of connectivity given the climatic conditions.

Mazapil constitutes one of the three most genetically diverse populations but also represents, according to the estimation of the habitat, the most stable region over the climate dynamics since the interglacial cycles, and thus we interpreted that this region has been a refuge for *P. pinceana* over time. This region has not been previously described as a refuge for the ChD (Vasquez-Cruz & Sosa; Loera et al., 2017; Scheinvar et al.; Gámez et al., 2017).

## **Conclusions**

The genetic diversity in *P. pinceana* was modified by dynamics during interglacial cycles in the Pleistocene. The demographic scenarios studied resulted in a ranking of models that were useful in gauging relative support for competing hypotheses. In particular, the best model involved the divergence of two lineages in the Middle Pleistocene (~627 kya; 95% CI: 633.37 – 584.52), which later colonized two biogeographic regions, the SMO and the ChD, occurring while interglacial cycles modified the Chihuahuan Desert as the aridification increased and paleolakes reduced their area. Both phylogenomic and population genomic coalescent-based analyses indicated that *P. pinceana* diverged into two lineages. This division probably occurred during the climatic changes of the Pleistocene and was related to the glacial-interglacial cycles. Thus, the phylogeographic history of *P. pinceana* is likely explained by climate dynamics that left perceptible marks in the patterns of genetic diversity and structure observed today in the species.

## Data availability statement

Raw reads are available in the SRA NCBI database (<https://www.ncbi.nlm.nih.gov/sra/>) deposited under the PRJNA719106 BioProject.

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### **Biosketch**

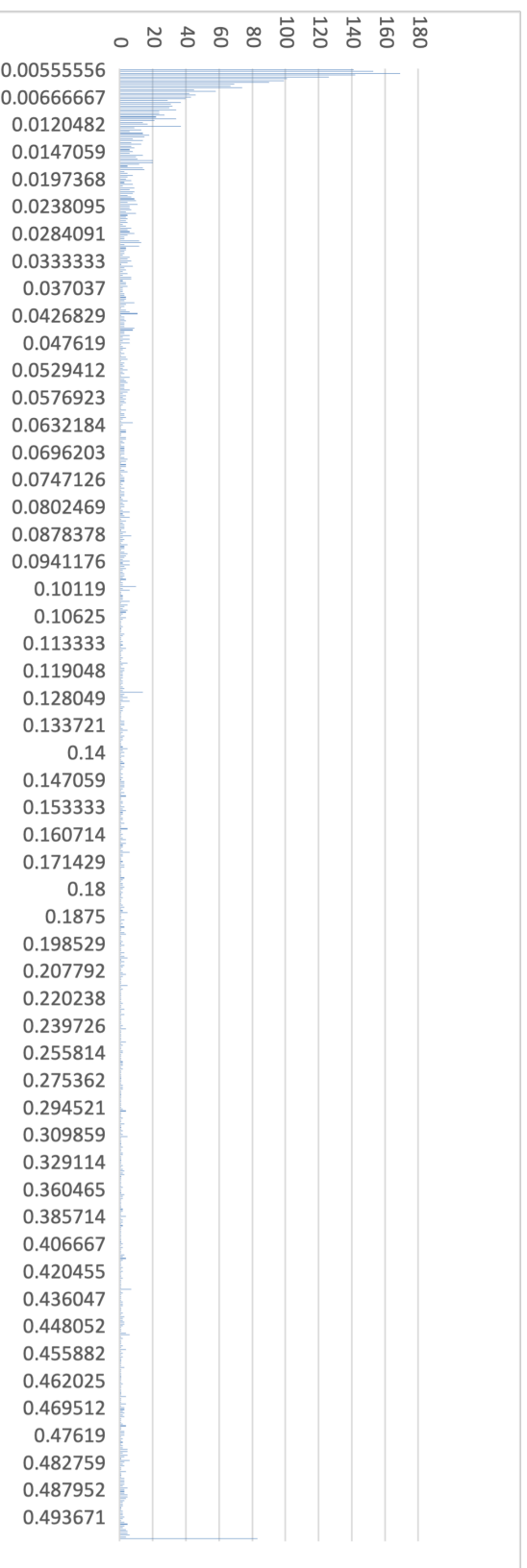
Author contributions: L.F.C., D.P. and D.S.G. formulated the project; A.M.L. analyzed the ecological niche modeling hypotheses, L.F.C. and P.P. conducted the bioinformatics processing, J.W. gave guidance on bioinformatics processing; D.O.D. gave guidance on bioinformatics and demographic history processing analyses. L.E.E. gave guidance on demographic history and the biogeographic patterns in the Chihuahuan desert. L.F.C. and D.P. led the writing. All authors commented on previous versions of the manuscript read and approved the final manuscript.

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## Supplementary materials



**Figure S1.** Site frequency spectrum for 4 141 SNPs

**Table S1.**  $F_{IS}$  calculation for extreme values reductions from 4 141 single nucleotide polymorphism loci

Removed	-	$\leq -0.949$	$\leq -0.899$	$\leq -0.849$	$\leq -0.799$	$\leq -0.749$	$\leq -0.699$	$\leq -0.649$	$\leq -0.59$	$\leq -0.549$	$\leq -0.499$
Average	-0.19	-0.15	-0.14	-0.13	-0.12	-0.12	-0.11	-0.1	-0.1	-0.09	-0.09
Median	-0.03	-0.01	-0.01	-0.01	-0.01	-0.01	-0.01	-0.01	-0.01	-0.01	-0.01

To establish the impact of putative paralogs, we provide the calculation for the mean and median  $F_{IS}$  values to show the effect of reducing the extreme values

**Table S2.** Measures of genetic diversity for 90 individuals of *P. pinceana* from ten geographical populations calculated from 3 762 single nucleotide polymorphism loci.

Population	$H_E$	$H_o$	$F_{IS}$	$\pi$	Tajima D	$\theta_w$	
Tolantongo	0.074	0.10	-0.31	0.0082	-0.42 <sub>NS</sub>	0.10	
San Joaquín	0.079	0.10	-0.35	0.0082	-0.31 <sub>NS</sub>	0.11	
Maguey Verde	0.099	0.14	-0.33	0.0065	-0.41 <sub>NS</sub>	0.09	
SMO	Nuñez	0.078	0.11	-0.37	0.0088	-0.30 <sub>NS</sub>	0.11
	La Florida	0.074	0.10	-0.35	0.0065	-0.30 <sub>NS</sub>	0.11
ChD	El Palmito	0.079	0.10	-0.37	0.0072	-0.37 <sub>NS</sub>	0.11
	Mazapil	0.090	0.12	-0.34	0.0072	-0.31 <sub>NS</sub>	0.12
	La Noria	0.078	0.11	-0.35	0.0087	-0.25 <sub>NS</sub>	0.13
	G. Cepeda	0.099	0.14	-0.36	0.0087	-0.18 <sub>NS</sub>	0.13
	Sierra Parras	0.095	0.13	-0.38	0.0090	-0.14 <sub>NS</sub>	0.13
	Global	0.116	0.0934	-0.35	0.0066	-0.63 <sub>NS</sub>	0.15

$H_o$ , observed heterozygosity;  $H_E$ , expected heterozygosity within populations;  $F_{IS}$ , inbreeding coefficient;  $\pi$ , nucleotide diversity

**Table S3.** Pairwise differentiation ( $F_{ST}$ ) among the ten populations sampled of *P. pinceana* with 3 762 SNPs

SMO	Tolantongo								
	San Joaquín	0.0061							
	Maguey Verde	0.0062	0.0069						
	Nuñez	0.0098	0.0173	0.0181					
	La Florida	0.0103	0.0195	0.0194	0.0104				
ChD	El Palmito	0.0451	0.0492	0.0468	0.0472	0.0487			
	Mazapil	0.0488	0.0506	0.0552	0.0545	0.0545	0.0166		
	La Noria	0.0521	0.0545	0.0586	0.0565	0.0581	0.009	0.0089	
	G. Cepeda	0.0525	0.058	0.0608	0.0603	0.0593	0.0184	0.0069	0.0043
	Sierra Parras	0.0596	0.0588	0.0645	0.0651	0.0648	0.02	0.0053	0.0103

### Capítulo 3

## **The introgression truly rules a part in the diversification of large cone pinyon pines (*Pinus* subsection *Cembroides* Engelm.) from North America?**

Laura Figueroa-Corona, José-Rubén Montes, Jill Wegrzyn, David S. Gernandt & Daniel Piñero

El siguiente capítulo está constituido por el artículo: The introgression truly rules a part in the diversification of large cone pinyon pines (*Pinus* subsection *Cembroides* Engelm.) from North America?, que está en proceso de elaboración y tiene el objetivo de probar la hipótesis planteada por Ledig et al. (2001) que sugiere que la diversidad en *P. pinceana* es resultado de la introgresión con *P. maximartinezii*. Para esto estudié la variación intraespecífica del genoma plastídico y SNPs en 858 genes nucleares de bajo número de copias, con estos dos sets de datos se reconstruyeron las relaciones filogenéticas y se evaluó si existieron eventos de introgresión o migración en tres especies de pinos piñoneros de conos grandes. Los resultados obtenidos del ensamblaje del genoma plastídico de 18 plastomas de *Pinus pinceana*, *P. rzedowskii* y *P. maximartinezii* y 112,893 SNPs muestran reconstrucciones filogenéticas que coinciden con estudios evolutivos previos indicando que la relación entre los linajes de pinos piñoneros de conos grandes y reflejan la ausencia de eventos de flujo genético o reticulación entre especies. Sin embargo, en ambas reconstrucciones se obtuvo en el linaje de *P. pinceana* una divergencia que refleja la estructura genética dentro del Desierto Chihuahuense y la Sierra Madre Oriental.

# The introgression truly rules a part in the diversification of large cone pinyon pines (*Pinus* subsection *Cembroides* Engelm.) from North America?

Laura Figueroa Corona<sup>1,2</sup>, José-Rubén Montes<sup>1,3</sup>, Jill Wegrzyn<sup>4</sup>, David S. Gernandt<sup>3</sup> and Daniel Piñero<sup>2</sup>

<sup>1</sup> Posgrado en Ciencias Biológicas, Universidad Nacional Autónoma de México, Ciudad de México, Mexico

<sup>2</sup> Departamento de Ecología Evolutiva, Instituto de Ecología, Universidad Nacional Autónoma de México, Ciudad de México, México

<sup>3</sup> Departamento de Botánica, Instituto de Biología, Universidad Nacional Autónoma de México, Ciudad de México 04510, México

<sup>4</sup> Department of Ecology and Evolutionary Biology, University of Connecticut, Storrs, CT 06269, USA

Email addresses:

LFC: laurafc25@gmail.com

JRM: ruben.montes@st.ib.unam.mx

JW: jill.wegrzyn@uconn.edu

DSG: dgernandt@ib.unam.mx

DP: pinero@unam.mx

## ABSTRACT

- *Premise of the study:* We addressed whether the lineages in the three pine species experienced introgression events or migration using the intraspecific nuclear variation and plastid genome, two different set of inheritance to reconstruct the phylogenetic divergence in the large cone pinyon pines (*Pinus* subsection *Cembroides* Engelm.)
- *Methods:* We inferred the lineages relationships under coalescent-based method and Bayesian inference. We used to infer a species tree with 112,893 polymorphisms in 858 protein-coding nuclear genes and an inference with 18 assembled plastomes from

*Pinus pinceana*, *P. rzedowskii*, and *P. maximartinezii*. Finally, we performed a reticulation analysis using Dsuite applied to 112,893 nuclear biallelic SNPs across quartets.

- *Results:* Phylogenetic and coalescent results coincide with previous evolutionary studies. For the three species of large cone pinyon pines show the same topology and the *D* test results did not detect events of gene flow or introgression within *Pinus pinceana*, *P. rzedowskii*, and *P. maximartinezii*. Within the *P. pinceana* show a lineage divided into two clades that reflects the genetic population structure within the Chihuahuan desert and the Sierra Madre Oriental regions.
- *Discussion:* Our results add evidence for divergences without introgression in the diversification inside the large cone pines. The high diversity that previously has been described as an introgression between species could be explained by the phylogeographic pattern and isolation from two genetic groups in *P. pinceana*.

**KEYWORDS:** large cones pinyon pines, plastomes lineages, *Pinus pinceana*, *Pinus maximartinezii*, *Pinus rzedowskii*.

## INTRODUCTION

*Pinus* subsection *Cembroides* Engelm. is a group of small to medium-sized trees and shrubs with economical and biological importance ([Richardson, 2000](#)). The North America pinyon pines have enlarged seeds with lipid-rich megagametophytes and an absent or vestigial wing. Pinyons present divergent morphology mainly in cones, seeds, and needles ([Malusa, 1992](#)). Pinyon pines have been divided into two groups, those with small cones (< 10 cm) and those with large cones (>10 cm; [Malusa, 1992](#)). The large cone group is monophyletic and comprises *Pinus maximartinezii* Rzed., *P. pinceana* Gordon, and *P. rzedowskii* Madrigal & M. Caball ([Montes et al., 2019](#)). [Ortiz-Medrano et al. \(2016\)](#) described a positive relationships between enlarged seeds and cones with aridity dated in the basal divergence of *P. maximartinezii* and *P. pinceana*. Phylogenetic studies suggest that wingless seeds evolved either once with a reversal to winged seeds in *P. rzedowskii*, or twice—once in the small cone clade and once in the ancestor of *P. pinceana* and *P. maximartinezii* ([Gernandt et al., 2005](#)).

A third alternative is that the MRCA of *Pinus* section *Parrya* had wingless seeds, and wings were regained in the ancestor of *Balfouriana* and in *P. rzedowskii*.

*Pinus maximartinezii* and *P. pinceana* are endemic to Mexico with high levels of genetic variation heterogeneously distributed among populations ([Ledig et al., 2001](#); [Molina-Freaner et al., 2001](#)). *Pinus pinceana* is distributed in the Sierra Madre Oriental (SMO) and the Chihuahuan desert (ChD), whereas *P. maximartinezii* is confined to two Sierra Madre Occidental (SMOc) forest localities. Based on monoterpene composition ([Zavarin & Snajberk, 1987](#)), and high diversity on the cpDNA SSRs frequencies [Ledig et al. \(2001\)](#), proposed the hypothesis that *P. pinceana* and *P. maximartinezii* have undergone subtle events of hybridization that produce the speciation of *P. maximartinezii*.

Using chloroplast and nuclear markers ([Gernandt et al., 2005](#); [Montes et al., 2019](#); [Willyard et al., 2007](#)) recovered a sister group relationship between *Pinus maximartinezii* and *P. pinceana* with high statistical support. Recently, genomic approaches discovered phylogeographic patterns and detected changes in genetic diversity of *P. pinceana* during the climatic dynamic on the interglacial cycles over the Pleistocene. The best supported model involved the isolation of two lineages, one distributed in the SMO, and the other extended over the ChD ([Figuroa-Corona et al., in prep](#)). Here, considering that the plastome is an independent evolutionary unit, paternally inherited may carry signals of reticulation or introgression events, we hypothesize that divergence in plastid and nuclear markers will be useful to reconstruct the evolutionary changes and migration events between populations of *P. maximartinezii* and *P. pinceana* populations.

## METHODS

### Sampling

We sampled needles of 17 individuals from 14 locations and one haploid megagametophyte seed from the entire distribution range of *P. pinceana*, three individuals of *P. maximartinezii*, and one sample from *P. rzedowskii* as an outgroup (Table S1; Figure S1). We extracted DNA using the CTAB method ([Doyle and Doyle, 1987](#)) for diploid leaf tissue and from the Qiagen DNeasy kit protocol for the haploid tissue *P. pinceana*. DNA concentration was measured

using a Qubit fluorometer and Qubit dsDNA HS assay kit (Life Technologies, Carlsbad, California, USA) and a Nanodrop spectrophotometer (Thermo Scientific, Waltham, Massachusetts, USA).

Samples with  $\geq 200$  ng of DNA and an A260/A280 between 2.0 and 2.2 were sent to Arbor Biosciences (Ann Arbor, MI, USA) for preparation of genomic libraries. For target enrichment (Gnirke et al., 2009), putative single copy nuclear genes were screened for probe design from *Pinus* species (*P. taeda* L., *P. pinaster* Aiton, and *P. sylvestris* L.; Neves et al. 2013; Willyard et al. 2007). A mixture of enriched and unenriched libraries were included in the sequencing runs to obtain both low-copy nuclear genes and high-copy number plastomes (Hyb-Seq; Weitemier et al., 2014). The samples were multiplexed and spread across eight Illumina paired-end sequencing runs (Table S2) that also included samples for other studies.

### Marker selection and gene selection alignment

Reads were trimmed using TRIMMOMATIC ver. 0.32 ([Bolger et al., 2014](#)), retaining sequences with an 80 bp minimum length and minimum quality score of 30. The procedure was executed with FASTQC ver. 0.11.7 (<https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>) and all the samples were condensed with MULTIQC ver. 1.9 ([Ewels et al., 2016](#)).

To identify SNPs, we used a total of 858 putative single-copy nuclear genes from the genome reference of *Pinus lambertiana* ver. 1.5 (available at <https://treegenesdb.org/jbrowse>). All high-quality reads were mapped using BWA-MEM ver. 0.7.10 ([Li et al., 2009](#)) with default parameters. The unmapped reads were discarded.

We used the Picard package (<http://broadinstitute.github.io/picard/>) Mark Duplicates to correct for artifacts of PCR duplication by only keeping one read or read-pair with the highest summed base quality among those of identical external coordinates and/or same insert lengths. Finally, we used Haplotypecaller and Variant Filtration tools from GATK ver. 3.7 ([McKenna et al. 2010](#)), SNP quality by depth (QD 2.0), mapping quality (MQ 40.0), Phred-scaled *P-value* using Fisher's exact test to detect strand bias (FS >60.0), Mapping



Quality Rank Sum Test (MQRankSum < -12.5), Read Pos Rank Sum Test (ReadPosRankSum < -8.0), and Strand Odds Ratio (SOR > 4.0) processing variants.

Finally, we made a filter prior to analysis using VCFTools ver. 0.1.16 ([Danecek et al., 2011](#)) to a list of biallelic SNPs and retained a total of 112 893 SNPs with a minimum mean depth > 6, Max missing by sample < 0.3, Max missing by site < 0.05, and a MAC of 1.

To qualify the structure within the samples we used Admixture ver. 1.3.0 ([Alexander & Lange 2011](#)) to estimate the population structure among individuals with admixture proportion from  $K = 3$  to 17. Genotypes were converted to ordinary PLINK files (.ped) using PLINK ver. 1.9 ([Purcell et al., 2007](#)).

### **Plastome assembly**

We used two short-read assemblers, one reference guided assembly conducted with Geneious ver. 9.2.6 ([Kearse et al., 2012](#)) and a *de novo* short read assembly conducted with SPAdes ver. 3.6.2 ([Bankevich et al., 2012](#)) to assemble plastomes. Trimmed reads were imported and processed in Geneious, were mapped to a reference plastome (*P. californiarum*; DSG1549) previously annotated from *P. monophylla* (MH612859; [Gernandt et al., 2018](#)), generating consensus sequences. Mapped reads were imported to SPAdes ver. 3.9 ([Bankevich et al., 2012](#)) in format FASTQ. The assembly in SPAdes resulted in several scaffolds per species which were imported again to Geneious. Scaffolds (<500 bp) were eliminated, and the rest were mapped to the previously generated consensus sequences. New consensus sequences were remapped to produce the final plastome sequences (see [Aguirre-Dugua & Gernandt, 2017](#)). Heterozygous sites were manually edited based on the quality of the sequences and comparison of the sites with a reference.

We eliminated plastomes with lower coverage values (<10×) prior to the phylogenetic analysis. Plastome sequences were aligned in MAFFT ver. 7.0 ([Katoh et al., 2002](#)) implemented in Geneious ver. 9.2.6. The alignment was imported to Gblocks ver. 0.91b for further editing ([Castresana, 2000](#)). To construct a haplotype network, we generated a median

joining network using PopART ver. 1.7 with the multiple alignment of plastid sequences (118.28 kb: [Leigh & Bryant, 2015](#)).

### **Plastome phylogenetic analyses**

Based on an alignment with the 18 individuals plastomes, nucleotide substitution models using the Akaike Information Criterion (AIC) were selected in jModelTest ver. 2.1.10 to choose the best model ([Darriba et al., 2012](#); [Miller et al., 2010](#)) to infer a phylogenetic tree under Bayesian inference with MrBayes ver. 3.2.6 ([Huelsenbeck and Ronquist 2001](#); [Miller et al., 2012](#)) with partition blocks for coding and non-coding regions. The analysis was conducted using the General Time Reversible model (GTR), allowing both rate and frequency variation combined with invariant sites (I+G). The analysis was run using four chains, three heated and one cold, and heating of 0.2. Two independent runs for 40,000,000 generations were performed with sampling every 1,000 generations, discarding 0.25 as a burn-in fraction. We used Tracer ver. 1.7.1 ([Rambaut et al., 2018](#)) to corroborate chain convergence. Tree topologies were summarized in a 50% majority rule tree. The consensus tree was imported into FigTree ver. 1.4.0 for further editing (Rambaut, 2012).

### **Coalescent-based analyses**

A species tree was inferred under a multispecies coalescent model using singular value decomposition scores for species quartets (SVDquartets) ([Chifman & Kubatko, 2014](#)) implemented in PAUP\* ver. 5.0 ([Swofford, 2002](#)). SVDquartets infers phylogenetic relationships with SNPs and accommodates incomplete lineage sorting (ILS). For this method, we used a concatenated alignment with 19 samples and 112,893 SNPs from 858 low-copy nuclear genes. We removed from the analysis the sample DSG440 source of a haploid megagametophyte to only analyze biallelic SNPs. We computed the quartet-based phylogeny under the QFM algorithm (Quartet Fiduccia and Mattheyses algorithm). A maximum of 1,000,000 randomly chosen quartets were estimated and we performed 500 replicates of bootstrap.

## Introgression analysis

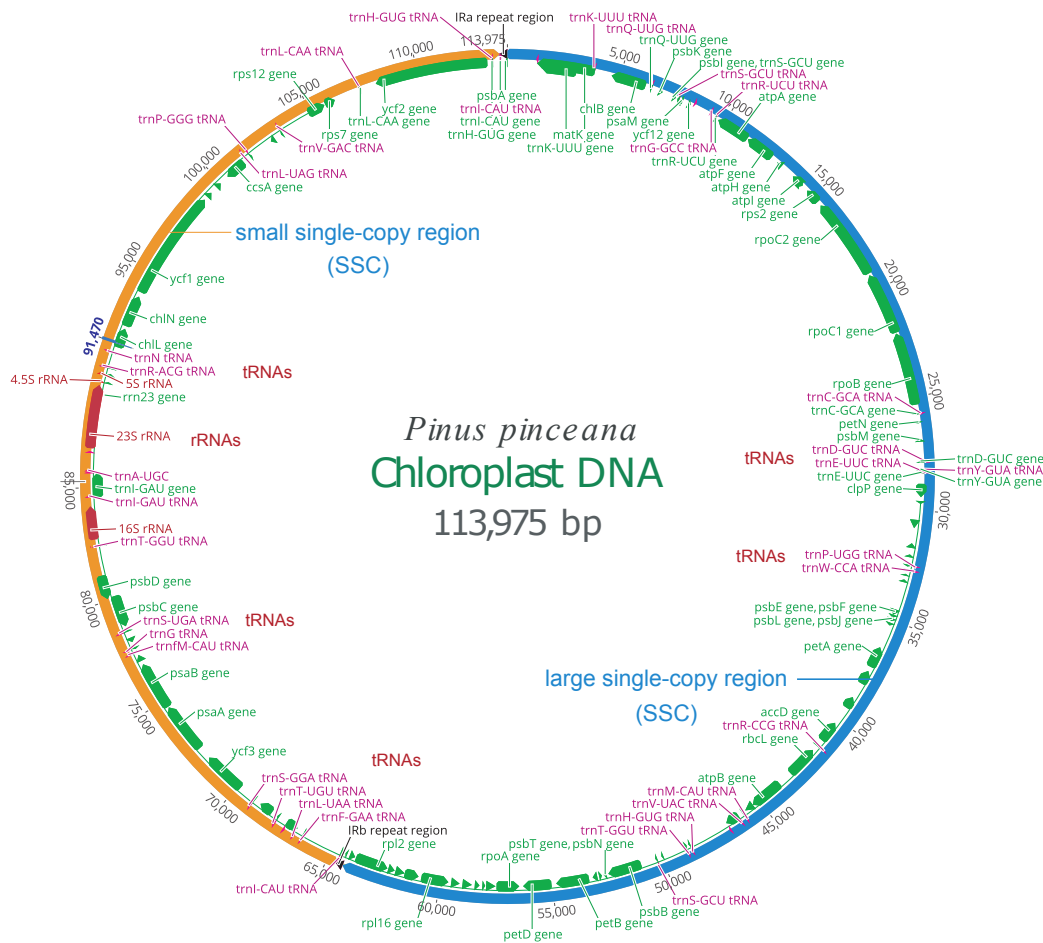
We used *Dsuite* ver. 1 ([Malinsky et al., 2020](#)) to estimate *D*-statistics (also known as the ABBA-BABA statistic) to detect gene flow or introgression between lineages dividing the populations of *P. pinceana* according to biogeographic regions. The test was applied to 112 893 biallelic SNPs across quartets (populations, taxa, species) on a typical tree topology (((P1, P2), P3), O), where O is the outgroup, *P. rzedowskii*, which served to define the ancestral allele (A), while the derived alleles were denoted by (B). Site patterns were ordered as follows: BBAA (P1 and P2 sharing the derived allele), ABBA (P2 and P3 sharing the derived allele), and BABA (P1 and P3 sharing the derived alleles). Under the null model (no gene flow), ABBA and BABA patterns were expected to occur with equal frequencies, while a significant deviation from that suggested possible introgression between P3 and either P1 or P2 ([Malinsky et al., 2020](#)). The *D* value was expected to be 0 if no gene flow had occurred and significantly different from 0 if gene flow (or introgression) existed between P2 and P3.

## RESULTS

### Plastome assembly statistics

The mean coverage was low in all plastomes assembled (11.7 - 36.69×). The only unusually high coverage was recorded in the unique haploid sample of *P. pinceana* from Hidalgo (440; 1908.5×) in the Sierra Madre Oriental. The atypical coverage value was produced by two probes for two cpDNA genes (UMN-6920 between *cemA* and *ycf4*, and 0-8850 near *psaB*) in the first version of the probes that were removed for the later versions (Table S2). In the *P. pinceana* plastome assemblies, we found a mean coverage of 18×, and a mean length of 113,975 pair bases (bp), but substantial variation in overall plastome size among samples was detected (Figure 1). Plastomes of *P. maximartinezii* had a length 103,141 bp and a mean coverage of 14×. Finally, the sample of *P. rzedowskii* had a length of 115,157 bp and a coverage of 36.69× (Figure S2; Table 1).

The overall difference of ~1,182 bp in plastome size reflected the substantial variation in a large single-copy region (LSC) between the inverted repeat region A (IRA) and inverted repeat region B (IRB). The average GC (%) content in CDS for both species was 38.6%. Gene density (total number of genes per kb) was 0.09 for *P. maximartinezii* and *P. pinceana*, and 0.097 for *P. rzedowskii*.



**Figure 1.** The plastid genome of *Pinus pinceana* (sample 440). Small and large single-copy regions (SSC, LSC) inverted repeat regions (IRA, IRB), tRNA sequences (purple), rRNA sequences (red) and gene sequences and gene clusters are shown as series in green.

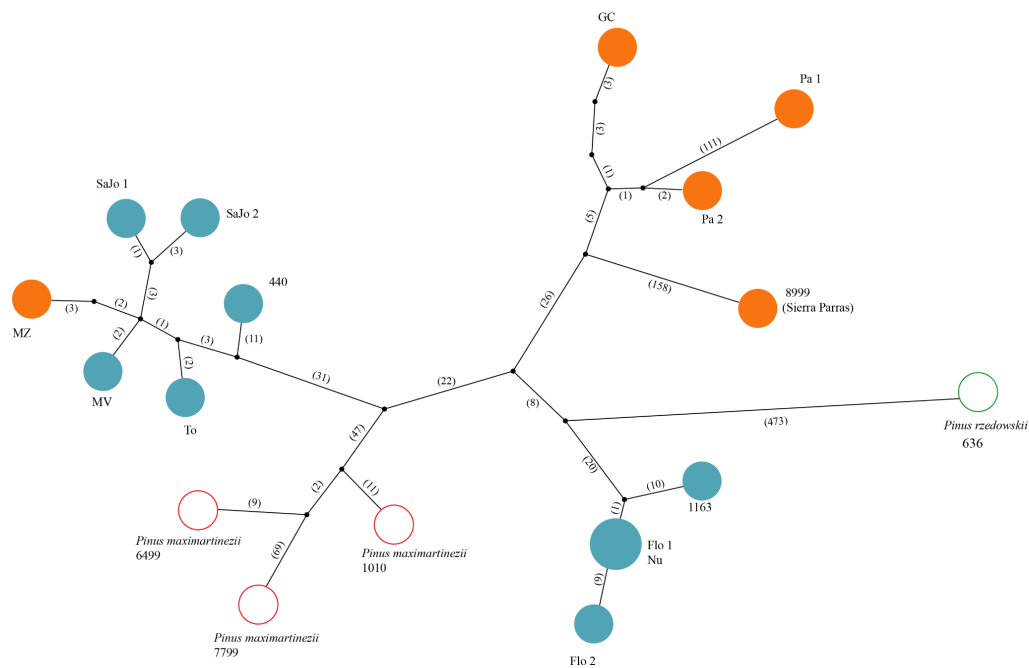
**Table 1.** General features of the assembly of complete plastomes of large cone pinyon pines.

Species			Total length	# contigs	N50	N75	Coverage	
<i>P. maximartinezii</i>		6499	113,141	13	13,116	5,821	11.98	
		1010	114,990	7	30,308	20,725	14.73	
		7799	109,503	37	4,071	2,736	7.59	
<i>P. rzedowskii</i>			636	115,157	7	19,355	17,948	36.69
<i>P. pinceana</i>	San Joaquin 1	SaJo1	116,296	3	73,913	41,575	25.43	
	San Joaquín 2	SaJo2	116,221	3	73,736	41,677	23.54	
	Tolantongo	To	116,342	6	21,711	20,868	22.95	
	Florida 1	Flo1	115,997	5	75,017	5,821	17.83	
	Florida 2	Flo2	116,313	5	42,111	42,111	16.7	
	Núñez	Nu	116,212	7	31,814	20,424	24.38	
	Maguey Verde	MV	115,470	5	41,161	41,161	35.1	
			1163	112,036	21	7,550	4,696	10.8
			440	113,975	13	13,971	9,377	1,908.52
			8999	110,365	45	3,886	1,765	7.32
	Mazapil General	MZ	115,878	5	41,684	41,684	22.29	
	Cepeda	GC	116,002	6	22,015	9,377	30.6	
	Palmito 2	Pa2	116,313	6	59,386	20,302	19.11	
Palmito 1	Pa1	116,455	6	22,047	20,496	33.95		

N50: median of lengths at 50; N75: median of lengths at 75; SMO: Sierra Madre Oriental; ChD: Chihuahuan Desert

No gene losses were detected across species. The most variable region between species was the LSC region, which had a length of 65,271 bp in *P. maximartinezii*, 65,207 bp in *P. rzedowskii* and 64,283 to 65,357 bp in *P. pinceana*. Length variation was not associated with provenance or phylogenetic relationship. Also, the *ycf1* gene presents insertions across species, ranging in length from 5,949 to 6,067 bp in *P. maximartinezii*, 5,004 to 6,217 bp in *P. pinceana* and 6,060 pb in *P. rzedowskii*. The *ycf2* region varied in length from 6,261 to 6,310 bp in *P. maximartinezii*, 4,978 to 6,294 bp in *P. pinceana* and 6,222 bp in *P. rzedowskii*. The *petB* gene length was 1,443 bp in *P. maximartinezii* and *P. rzedowskii* ranged from 1,438 to 1,443 bp in *P. pinceana* from the SMO samples and 1,443 to 1,461 bp from the ChD samples.

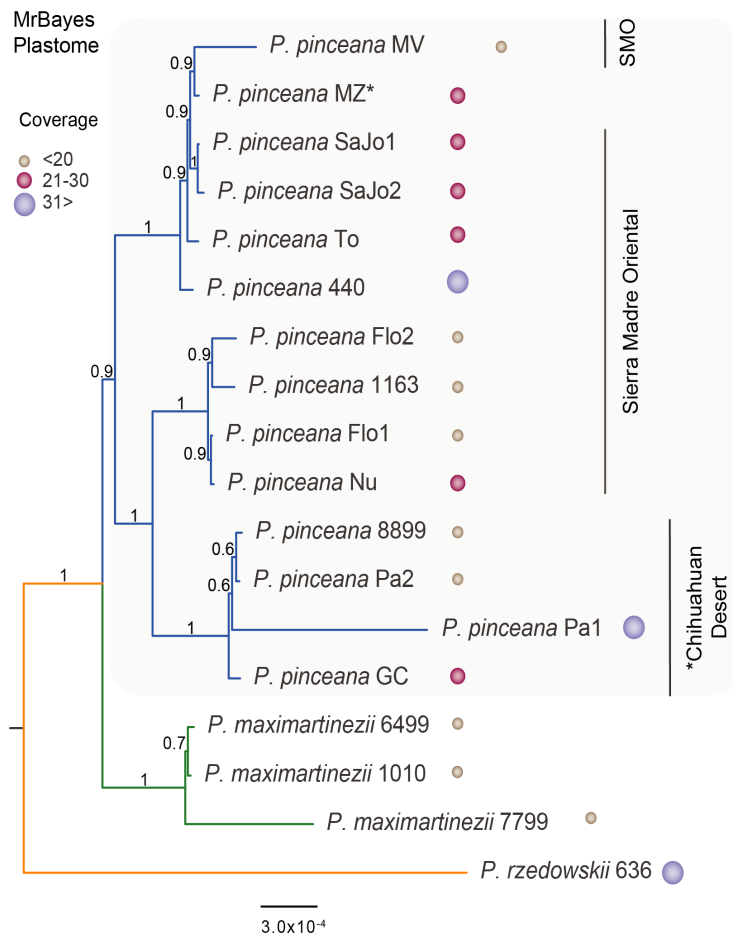
The haplotype network was constructed with 17 haplotypes identified from all the samples (Figure 2). Two samples from SMO locations Flo 1 and Nu share the same haplotype. Smaller distances in *P. pinceana* were observed in the SMO samples with 80 changes, similar to the distance between the closest haplotypes of *P. pinceana* and *P. maximartinezii*. A Mazapil sample (MZ) from the ChD had fewer differences with extreme south samples from the SMO than with other samples from the ChD. The greatest number of differences were found between the *P. rzedowskii* and the other haplotypes. Thus, the relationships among haplotypes corresponded to both biogeography and phylogeny.



**Figure 2.** Haplotype network of large cone pinyon pines constructed by the complete plastome genome. The size of the dots represents the number of individuals with a given haplotype weighted by the total polymorphism shared. On blue circles shows the SMO locations on orange circles the ChD locations of *P. pinceana*, for *P. maximartinezii* the red outline circles, and the green outline circle corresponds to *P. rzedowskii*.

### Phylogenetic and coalescence-based results

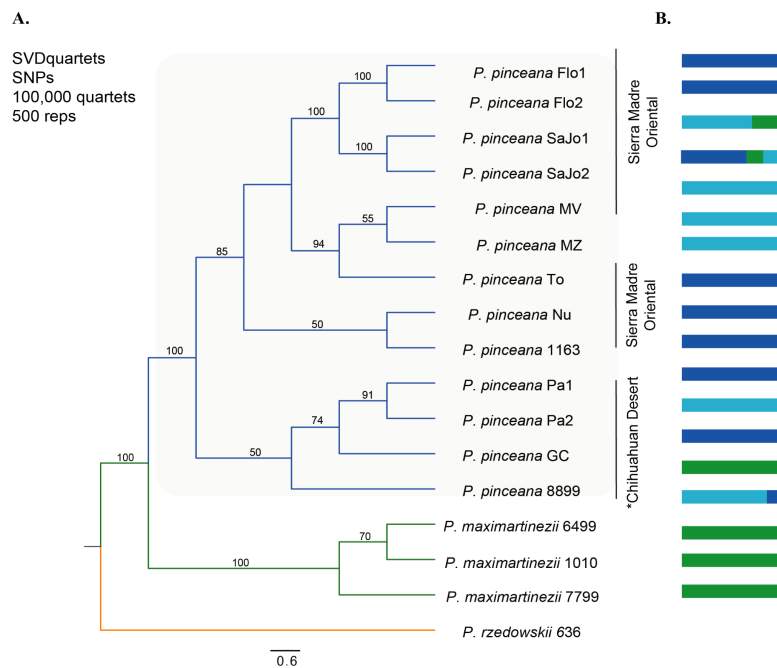
The plastome alignment had 18 terminals and 206 parsimony-informative characters (Figure 3). The Bayesian majority-rule tree showed that each species constitutes a well-supported (>0.9 posterior probability) monophyletic lineage. All relationships among lineages were well supported. The *P. pinceana* lineage is divided into two main clades that correspond to the biogeographic provenance of the samples, except for the Mazapil (MZ) sample. Most branches in the Bayesian-tree are short, except for *P. pinceana* PA1 and *P. maximartinezii* 7,799, that indicated few differences between taxa.



**Figure 3.** Consensus tree by the Bayesian inference based on 18 plastome sequences. On the right side, the sequencing coverage is shown proportional to the size of the circles. The values on branches represent the posterior probabilities.

For the nuclear reconstruction was used 12,521 parsimony informative SNPs from the 858 low copy nuclear genes. The bootstrap values for most branches in the phylogeny were up to 60%. The *P. pinceana* individuals were divided into two sister clades corresponding to the two biogeographic regions, Sierra Madre Oriental and Chihuahuan desert (Figure 4). To compare both reconstructions we assembled a tanglegram with the consensus tree from the plastome Bayesian inference and the ultrametric coalescence reconstruction of SVDquartets using the default parameters in Dendroscope (Huson & Scornavacca, 2012; Figure S3).

The admixture analysis shows results for  $K = 3$ , the best supported number of clusters. Only SaJo2 shows admixture with the 3 groups. The *P. maximartinezii* samples share complete similarity with *P. pinceana* GC sample and partially with SaJo samples. The remaining samples of *P. pinceana* samples have 100% similarity with two groups, without a geographical sense except for the 8899 sample, that shares admixture between these two groups (Figure 4B).



**Figure 4.** Ultrametric tree constructed by SVDquartets using 12,521 SNPs for nuclear genes concatenated from 858 lowcopy genes for 17 samples of large cone pinyon pines **A.** Maximum likelihood tree inferred. **B.** shows the admixture composition for the best supported clustering ( $K = 3$ ).



## Reticulation test

*D*-tests between lineages or populations based on 12,521 SNPs produced extremely low *D*-values ( $D = 0.00142$  on average) and a non-significant *p*-value between lineages (Table 2), rejecting the hypothesis of introgression between *P. pinceana* and *P. maximartinezii*. Identical values were obtained among *P. pinceana* and *P. maximartinezii*, which can be attributed to the equal number of ancestral polymorphisms shared between the two lineages of *P. pinceana* and *P. maximartinezii*.

**Table 2.** *D*-statistics for large cone pinyon pine clade lineages and genetic cluster using 12 521 polymorphisms from 858 low copy nuclear genes. Using a topology (((P1, P2), P3), O), where O is the outgroup, *P. rzedowskii*, which served to define the ancestral allele (A), while the derived alleles were denoted by (B).

P1	P2	P3	<i>D</i> statistic	<i>p</i> -value
<i>P. pinceana</i> SMO	<i>P. pinceana</i> ChD	<i>P. maximartinezii</i>	0.00127	0.240213
<i>P. pinceana</i> ChD	<i>P. pinceana</i> SMO	<i>P. maximartinezii</i>	0.00127	0.240213
<i>P. maximartinezii</i>	<i>P. pinceana</i> SMO	<i>P. pinceana</i> ChD	0.00173	0.117542

## DISCUSSION

### Plastome assembly statistics

Evolutionary studies using whole plastids in pines has increased in the last decade ([Parks et al., 2012](#); [Gernandt et al., 2018](#); Montes et al., *in prep*). The quality of sequences, total reads, and percent of coverage in pines depends on the library preparation and the sequencing platform ([Cronn et al., 2008](#); [Parks et al., 2012](#); [Gernandt et al., 2018](#)) and the assembly quality depends on data and assembler methods ([Abbas et al., 2014](#)). Using the Hyb-Seq method, we recovered 18 plastomes of 3 species with a mean length of  $114,664 \pm 2,145.5$  pb.

In contrast, the mean length for the recovered by [Parks et al. \(2012\)](#) using the Illumina platform was  $117,157 \pm 3,634$  bp in *P. thunbergii*, 2,493 bp more than reported here. Similarly, the total length of the plastome alignment after filtering was slightly shorter than previous studies (see [Gernandt et al. 2018](#); Montes et al. *in prep*) but substantially shorter than the genus-wide alignment of [Parks et al. \(2012\)](#), which had a total length of 141,265 bp. Since the species studied here are closely related, the informative sites were less than reported in others previous studies of pines (15,151 infs: [Parks et al., 2012](#); 1,833 infs: [Gernandt et al., 2018](#); 11,281 infs: Montes et al., *in prep*). However, our results add more information at population level.

### **Intraspecific vs interspecific differences**

The sequence of plastidic genome within the large cone pinyon pines shows low divergence; this outcome has been reported in other whole chloroplast assemblies ([Whitall et al., 2010](#)) as a result of the reported low substitution rate in *Pinus* ( $7.28 \times 10^{-10}$  substitution per site per year; [De la Torre et al., 2017](#)), combined with rapid diversification events during the Last Miocene / Pleistocene (Montes et al., *in prep*). Nevertheless, the topology found in the nuclear and plastid is congruent between species phylogenies. The absence of a signal of hybridization in this clade of pines indicates that reticulate evolution is not a main cause in the large cone pinyon pines.

One of the results to emerge from our analysis of nearly plastidic genome is low within these pine species. The most variable region was the large single-copy region, which play a vital role in regulation of expression, and the intron or gene losses that have been found in other chloroplast genomes ([Xu et al., 2020](#); [Liu et al., 2018](#); [Shrestha et al., 2019](#)).

The separation of *P. pinceana* into two lineages with both nuclear and plastid DNA reflects the genetic structuring within the biogeographic regions. This result was reported in an earlier study using cpDNA SSR loci ([Escalante, 2001](#)). More recently it has been associated with demographic events during the Quaternary, when the interglacial dynamics increased aridity producing changes over the distribution of *P. pinceana* distribution ([Figueroa-Corona et al., in prep](#)). The Chihuahuan Desert and Sierra Madre Oriental

populations are distinct and fully differentiated in both plastomes and nuclear genomes, with topologies that are fully congruent with other phylogenetic studies of subsection *Cembroides* ([Gernandt et al., 2005](#); [Whitall et al., 2010](#); [Montes et al., 2019](#)). This is shown graphically as a tanglegram constructed with default parameters in Dendroscope ([Huson & Scornavacca, 2012](#); Figure S3).

Our results cannot distinguish introgression events within the large cone pinyon pines. The admixture analysis indicates a degree of recombination within and between species. However, the support in the phylogenetic reconstruction shows a robust and clear divergence. Within the *P. pinceana* samples, the divergence between two different clades shows divergence in accordance with the biogeographical regions, with rare migration events between the two groups. The Mazapil sample (MZ) shows reticulation between the phylogenetic reconstruction between plastomes and nuclear markers, thus could be explained by two hypotheses, one, because it is one of the most genetically diverse populations, and projections along the Pleistocene indicated that this is the most stable region for *P. pinceana* constituting a Pleistocene refugia along the interglacial cycles working as reservoir the most ancient genetical diversity ([Figueroa-Corona et al., in prep](#)), and second, could be evidence of plastome capture produced by the transit of pollen across population from the different biogeographical regions.

Given the results obtained and the high agreement with other phylogenetic reconstructions obtained with different markers, we find no evidence to support the introgression hypothesis proposed by [Ledig et al., \(2001\)](#), that the diversity inside *P. pinceana* was a reflection of past hybridization events with the sister species *P. maximartinezii*.

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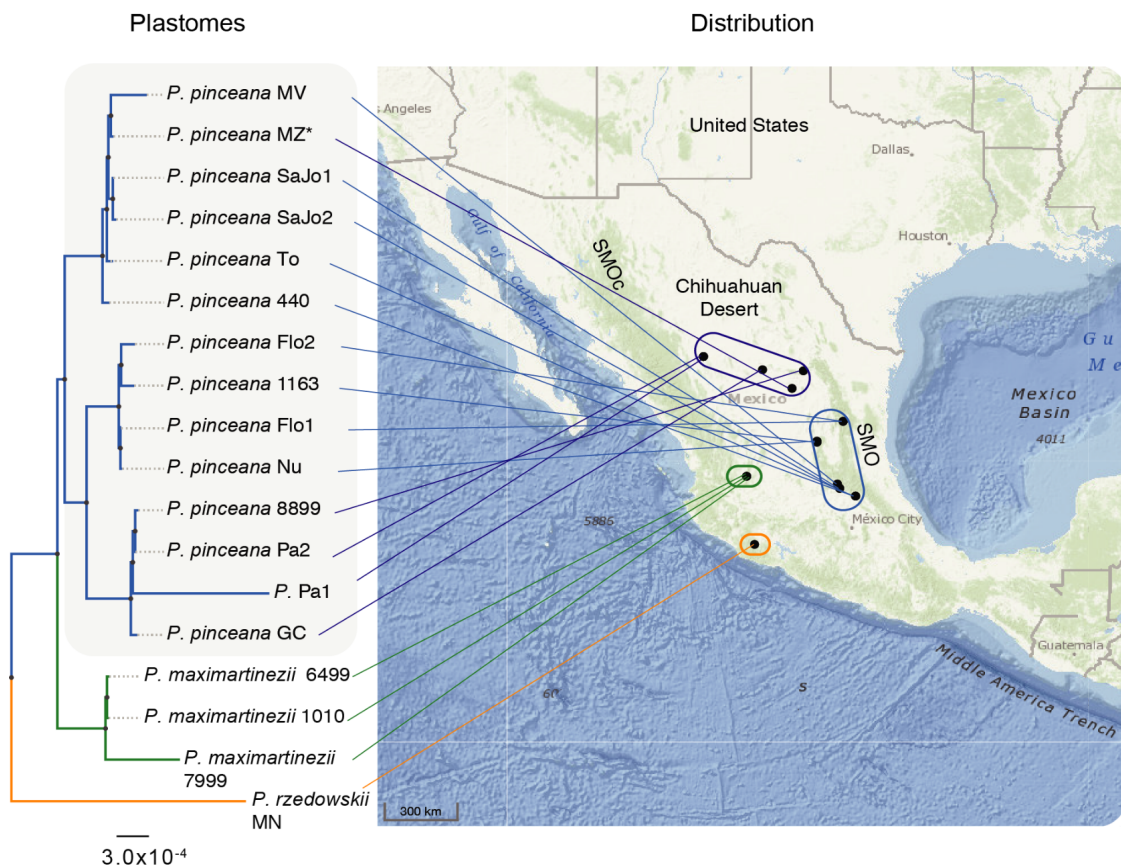
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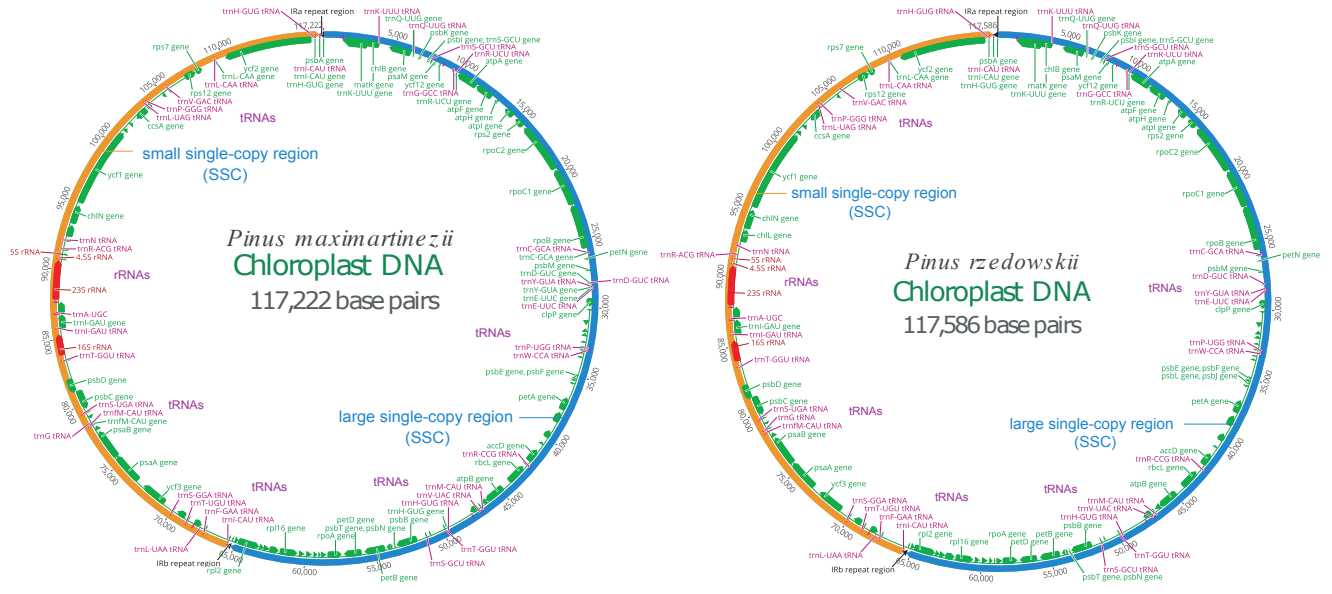
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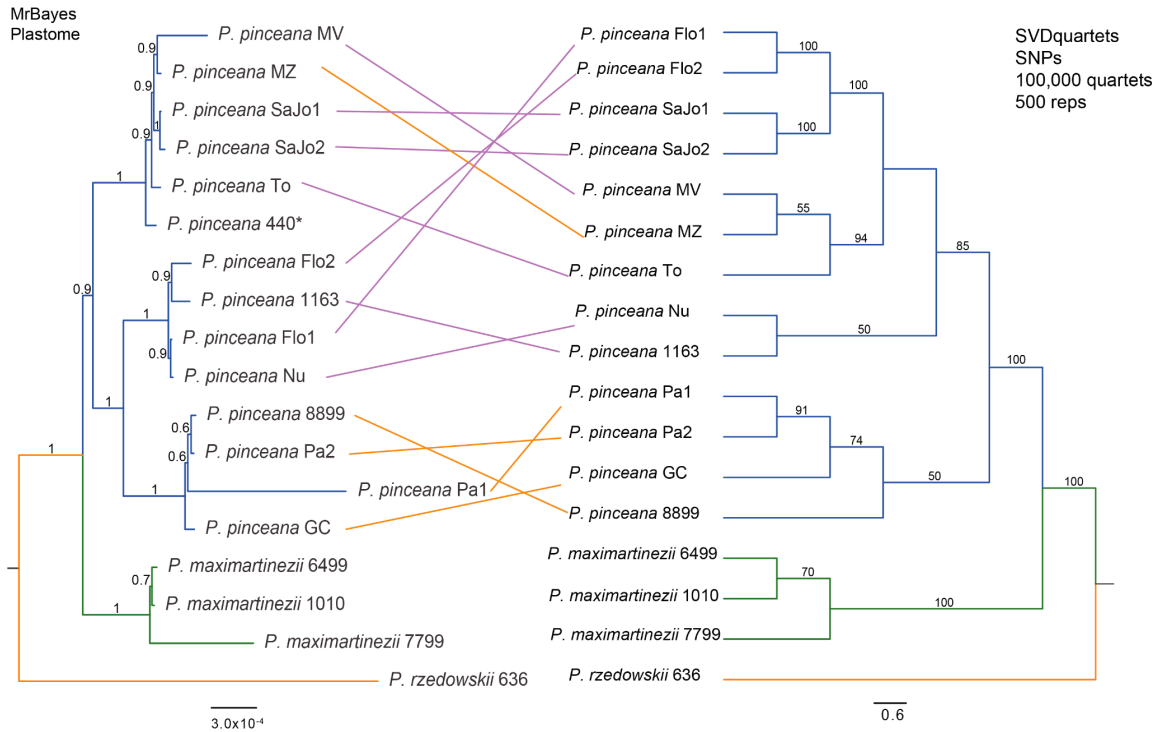
## Supplementary material



**Figure S1.** Geographic distribution of samples tracking the phylogenetic reconstruction with the chloroplast genome.



**Figure S2.** The complete chloroplast genome of *Pinus maximartinezii* and *P. rzedowskii*



**Figure S3.** Tanglegram of Bayesian majority-rule consensus tree and SVDquartets lineages tree. A. Maximum likelihood tree based on concatenated alignment with 858 nuclear genes. B. Coalescent-based tree based on 1 000 000 quartets.



**Table 1S.** Sampled sites

<b>Location</b>		<b>Condition</b>	<b>Latitude (N)</b>	<b>Longitude (W)</b>
<b>Tolantongo</b>	To	SMO	20.63	98.99
<b>Magüey Verde</b>	MV	SMO	21.09	99.695
<b>Nuñez</b>	Nu	SMO	22.671	100.483
<b>San Joaquín</b>	SaJo	SMO	20.91	99.633
<b>DSG 440</b>		SMO	20.635	98.99
<b>DSG 1163</b>		SMO	22.618	100.488
<b>Florida</b>	Flo	SMO	23.4	99.503
<b>Mazapil</b>	MZ	ChD	24.59	101.472
<b>DSG 8999</b>		ChD	25.255	101.02
<b>General Cepeda</b>	GC	ChD	25.301	102.607
<b>Palmito</b>	Pal	ChD	25.741	104.881
<b><i>P. maximartinezii</i></b>				
<b>DSG6499</b>			19.493	98.848
<b>DSG7799</b>			21.35	103.225
<b>DSG1010</b>			19.317	99.195
<b><i>P. rzedowskii</i></b>				
<b>DSG636</b>			18.821	102.926

**Table S2.** Conditions and concentrations of the parallel sequencing per sample. Ratio of unenriched (unenriched:enriched).

Lane	Probe version	ID collector	Sample	DNA	Samples/lane	sequencing depth	Illumina Sequence Lengths
GP02	V1	DSG440	<i>Pinus pinceana</i>	Genomic, 1n from megagametophyte	48	20:80	100
GP03	V2	DSG1163	<i>Pinus pinceana</i>	Genomic, 2n from needle	48	30:70	100
GP03	V2	DSG8999	<i>Pinus pinceana</i>	Genomic, 2n from needle	48	30:70	100
GP03	V2	DSG07799	<i>Pinus maximartinezii</i>	Genomic, 2n from needle	48	30:70	100
GP03	V2	DSG1010	<i>Pinus maximartinezii</i>	Genomic, 2n from needle	48	30:70	100
GP03	V2	DSG6499	<i>Pinus maximartinezii</i>	Genomic, 2n from needle	48	30:70	100
GP03	V2	DSG636	<i>Pinus rzedowskii</i>	Genomic, 2n from needle	48	30:70	100
GP06	V2	Sierra de Parras	<i>Pinus pinceana</i>	Genomic, 2n from needle	48	40:60	125
GP06	V2	Mazapil	<i>Pinus pinceana</i>	Genomic, 2n from needle	48	40:60	125
GP06	V2	Nuñez	<i>Pinus pinceana</i>	Genomic, 2n from needle	48	40:60	125
GP06	V2	Palmito1	<i>Pinus pinceana</i>	Genomic, 2n from needle	48	40:60	125
GP06	V2	Palmito2	<i>Pinus pinceana</i>	Genomic, 2n from needle	48	40:60	125
GP06	V2	Maguey Verde	<i>Pinus pinceana</i>	Genomic, 2n from needle	48	40:60	125
GP06	V2	General Cepeda	<i>Pinus pinceana</i>	Genomic, 2n from needle	48	40:60	125
GP06	V2	Tolantongo	<i>Pinus pinceana</i>	Genomic, 2n from needle	48	40:60	125
GP07	V3	Florida 1	<i>Pinus pinceana</i>	Genomic, 2n from needle	96	40:60	150
GP07	V3	Florida 2	<i>Pinus pinceana</i>	Genomic, 2n from needle	96	40:60	150
GP07	V3	San Joaquín 1	<i>Pinus pinceana</i>	Genomic, 2n from needle	96	40:60	150
GP07	V3	San Joaquín 2	<i>Pinus pinceana</i>	Genomic, 2n from needle	96	40:60	150

## Discusión general

La arquitectura genética, es decir, las contribuciones genéticas a un fenotipo son variables en tiempo y espacio, influenciadas por la mutación, la migración, la recombinación, la deriva genética y la selección (Timpson et al., 2018). La variación genética se fundamenta como el sujeto de la evolución y conforma a largo plazo un efecto en el fenotipo. Esta variación genética comprende el número de polimorfismos genéticos que influyen en el rasgo, las frecuencias de los polimorfismos en la población, la magnitud de sus efectos, además de sus interacciones entre genes y con el ambiente (Savolainen et al., 2007; Timpson et al., 2018).

Este proyecto fue planteado con tres aproximaciones genómicas en una de las coníferas más ampliamente distribuidas en condiciones de extrema aridez, *Pinus pinceana*, con el objetivo de combinar estrategias de secuenciación de alto rendimiento para estudiar tanto en regiones neutrales como en genes altamente conservados por el efecto intenso de la selección purificadora y la actividad transcripcional. Con tres diferentes aproximaciones genéticas, analicé 1) la expresión transcripcional en *P. pinceana*, y cómo responde fisiológicamente al medio ambiente, dada sus interacciones a factores bióticos y abióticos; 2) el efecto de su historia demográfica y los patrones filogeográficos en su diversidad genética; y 3) los patrones de divergencia dentro de los linajes de *P. pinceana* y sus especies hermanas.

### 1. Transcriptómica en *Pinus pinceana*

La aproximación funcional en este proyecto, planteada a partir de la identificación la respuesta a la aridez por medio de los cambios en la expresión genética, la describo en el Primer Capítulo, donde planteé entender la relación fenotipo-ambiente utilizando siete individuos de localidades geográficas con distintos patrones de precipitación y temperatura en el año. El ensamble *de novo* del transcriptoma de *P. pinceana* se caracterizó entre 27,217 y 42,590 transcritos (CDS) en los individuos analizados. La varianza transcripcional entre localidades responde esencialmente a dos factores: 1) la naturaleza de los estudios de RNA-seq, como una muestra puntual, local y específica a las condiciones del organismo en el

momento de la colecta; 2) la variación entre las localidades y el diseño experimental usado no estuvieron sujetas a condiciones controladas u homogéneas, ya que se priorizó coleccionar individuos en toda el área de distribución sin utilizar réplicas locales que permitieran una mejor representatividad local. Consecuentemente, esta varianza transcripcional repercute en las rutas metabólicas caracterizadas, influye en los niveles de expresión cuantificados y evidencia la diversidad de relaciones ecológicas locales con organismos parásitos y endófitos.

A pesar de la alta heterogeneidad, el número de transcritos caracterizados está de acuerdo con lo reportado en estudios previos de transcriptomas de acículas de coníferas y del número estimado de genes en los genomas de coníferas. Por ejemplo, el transcriptoma de *Sequoiadendron giganteum* con 35,183 CDS; *P. cembroides* con 20,656 CDS; o en *P. sylvestris* con 24,780 CDS (Scott et al., 2020; Webster et al., *in prep.*; Ojeda et al., 2019).

La variabilidad detectada en los perfiles de expresión genética podría reflejar la respuesta a factores bióticos y abióticos, a pesar de que en el estudio no pudimos atribuir ninguna relevancia adaptativa, en parte debido a la cobertura relativamente baja en la comparación entre muestras, y a la ausencia de genes candidatos específicos. Las rutas metabólicas caracterizadas en el transcriptoma de *P. pinceana* ayudaron a identificar la respuesta transcripcional en las localidades naturales reguladas por la expresión génica de rutas metabólicas de respuesta al estrés biótico o abiótico, que involucran la tolerancia al estrés e incluyen mecanismos que limitan y reparan el daño a los componentes celulares.

Las respuestas fisiológicas de respuesta al efecto ambiental y las adaptaciones a la aridez están influenciadas por mecanismos internos y factores ambientales externos y la interacción entre ambos. Las respuestas fisiológicas son variables en duración e intensidad, afecta la síntesis, concentración, transporte y almacenamiento de metabolitos primarios y secundarios (Brodrigg et al., 2005). Los ajustes ocurren en múltiples vías metabólicas para restaurar los desequilibrios químicos y energéticos.

A partir de los resultados obtenidos en el Primer Capítulo, caractericé los transcritos presentes en 7 árboles de su distribución natural, y evalué el perfil de expresión génica, comparándolos entre regiones ambientales y con dos árboles fuera de rango altitudinal y de

precipitación en el área natural de distribución. Con ambas comparaciones, describí la participación de genes que actúan en protección estructural relacionados con las condiciones de aridez y estrés, como osmoprotectores, acuaporinas, chaperonas, proteínas de embriogénesis tardía y proteínas de choque térmico; factores que promueven la eliminación de especies reactivas de oxígeno (ROS); y reguladores del metabolismo de la pared celular y de la protección a las estructuras celulares. A nivel hormonal, encontré reguladores en la síntesis del ácido salicílico (SA), ácido jasmónico (JA), y genes *pr-1* ortólogos identificados previamente en brasinoesteroides, implicados en las respuestas al estrés por aridez, congelación, salinidad y osmótico (Akbulak et al., 2020; Jirage et al., 1999). Corcuera et al. (2012a, b) correlacionaron en *P. pinaster* los niveles de JA con la humedad relativa del aire y con la presencia de patógenos, puesto que también son inductores de la producción de JA en lesiones locales (Rojo et al., 2003; Koo & Howe, 2009).

La presencia de intermediarios en la regulación por ABA resulta igualmente interrelacionada con la respuesta al estrés abiótico. Para las coníferas el transporte a distancia de reguladores de ABA puede no resultar eficiente a fin de provocar el cierre de los estomas a tiempo y evitar déficits de agua en los tejidos (Brodrribb et al., 2014). Perks et al. (2002) encontraron que en *P. sylvestris* que el cierre de los estomas precedió al aumento de la concentración de ABA en respuesta a un aumento en el déficit de agua del suelo. Es importante resaltar que los factores reguladores de SA encontrados ha sido descritos como inductores de la síntesis de osmoprotectores, como la prolina bajo estímulos de estrés hídrico (Umebese et al., 2009; Yang & Miao, 2010; Xiao et al., 2008, 2009). Li et al. (2014) describieron en *Torreya grandis* que la acumulación de SA y prolina inducen actividad de la cadena respiratoria, la síntesis de clorofila y la reducción de ROS, además de prevenir daño en membrana durante el estrés por salinidad.

La regulación hacia arriba de transcritos reguladores de la respuesta al estrés oxidativo que se han estudiado a partir del análisis de fenotipos tolerantes. Sánchez-Gómez et al. (2017) han descrito que la respuesta al déficit hídrico provoca una reducción de la tasa fotosintética, en la conductancia estomática y en la eficiencia de las cadenas respiratorias. De María et al. (2020) describieron en *P. pinaster* respuestas diferenciales para la tolerancia al

estrés hídrico y la regulación del estrés oxidativo con proteínas (RING-H2), que de acuerdo con los resultados del Primer Capítulo se encuentran reguladas positivamente en individuos de la región más árida del área de distribución de la especie.

La regulación hacia arriba de reguladores de la respuesta al estrés abiótico no fue exclusiva de las regiones de mayor aridez. También encontramos una mayor acumulación de transcritos que codifican para MYB4 y varias proteínas de choque térmico en SMO, factores de transcripción reportados en la tolerancia al estrés por calor, el equilibrio hídrico de las hojas y la respuesta a patógenos virales o fúngicos (Wang et al., 2019). MYB4 es un factor de transcripción que participa en la regulación de la síntesis de skimato y flavonoides, osmoprotectores, ABA, jasmonatos y brassinosteroides (Wang et al., 2020).

De manera conspicua, también encontramos transcritos relacionados directamente con la respuesta al estrés biótico. Entre las familias ortólogas génicas mejor representadas están las proteínas NBS-LRR, relacionadas con la inmunidad innata a través del reconocimiento de patógenos, que incluyen bacterias, virus, hongos, nemátodos, insectos y oomicetes (Meyers et al., 2002; Eitias et al., 2008; McHale et al., 2006). La presencia de estas proteínas concuerda con la incidencia de secuencias encontradas de insectos patógenos, de levaduras asociadas con ellos y de hongos patógenos y endófitos.

## 2. La diversidad genética e historia demográfica de *Pinus pinceana*

Más allá de la plasticidad y de la posible respuesta adaptativa en la regionalización ambiental actual de *P. pinceana*, exploré la relación genotipo-ambiente en el Segundo Capítulo de esta tesis, en el que se caractericé su diversidad genética en un contexto histórico.

Trabajos previos habían estudiado a *P. pinceana* usando aloenzimas y cpDNA SSRs, reportaron una alta variación genética al compararla con otras coníferas de amplia distribución. Estos estudios detectaron un patrón filogeográfico que divide a las poblaciones en dos linajes sin evidencia de migración, aparentemente aislados por la distancia entre las poblaciones (Ledig et al., 2001; Molina-Freaner et al., 2001; Escalante, 2001; Figueroa-Corona, 2012). A diferencia de estos trabajos, en esta tesis utilicé polimorfismos en el

genoma completo para la caracterización de la diversidad genética, con el objetivo de muestrear polimorfismos en regiones codificantes, reguladoras y neutrales.

Encontré una alta representación de alelos en baja frecuencia, y una alta diversidad dentro de las poblaciones en su hábitat fragmentado. La prevalencia de nuevos polimorfismos mantenidos por tamaños poblacionales efectivos relativamente grandes podría explicarse por dos escenarios diferentes: 1) la retención de nuevas variantes genéticas y, 2) la retención de parálogos y pseudogenes.

La diferenciación biogeográfica entre ChD y SMO ha sido reportado en varias especies de la comunidad biótica del Desierto Chihuahuense y, de manera similar, ha sido correlacionada con los cambios geológicos y paleoclimáticos. Otros organismos estudiados incluyen plantas como *Agave lechuguilla*, *A. striata*, y *A. stricta*, roedores como *Thomomys umbrinus*, *Perognathus avus*, y la tortuga *Kinosternon avescens* (Mathis et al., 2014; Neiswenter & Riddle, 2010; Serb et al., 2001; Scheinvar et al., 2017; Trejo et al., 2016; Martínez-Ainsworth, 2013; Scheinvar et al., 2020).

Para *Pinus pinceana*, los procesos de aislamiento y la baja abundancia de las condiciones climáticas favorables han mantenido la distribución a una zona muy reducida, por lo que queda limitado el contacto secundario y la migración. La separación de El Palmito, con una ubicación noroeste, aparentemente representa una fragmentación más reciente de las poblaciones del Desierto Chihuahuense. Este patrón de divergencia oeste/este en el Desierto Chihuahuense que también se ha descrito en organismos de matorrales xéricos como *Astrophytum* spp., *Berberis trifoliata*, *Agave victoria-reginae*, *Leucophyllum* spp. y *Ephedra compacta* (Vázquez-Lobo et al., 2015; Angulo et al., 2017; Gándara & Sosa, 2014; Loera et al., 2017), y en la serpiente *Crotalus molossus* (Anderson & Greenbaum, 2012). Esta regionalización de la diversidad genética en *P. pinceana* concuerda con sus patrones históricos en su demografía y con la dinámica climática modelada durante el Cuaternario. La separación entre sus dos linajes ocurrió durante el Pleistoceno medio durante los ciclos interglaciares (~627 kya; 95% CI: 584.52 - 633.37).

Los procesos de aridificación durante el Pleistoceno redujeron los cuerpos de agua en la región, y es posible que la cuenca de Nazas determinara la división dentro del ChD

produciendo la separación entre el noroeste y el centro. Con el incremento de la aridificación, las condiciones climáticas favorables para *P. pinceana* se volvieron más comunes en contraste con lo que se han planteado en la región para *Picea chihuahuana*, que actualmente las condiciones climáticas favorables para esta especie son escasas (Quiñonez-Pérez et al., 2017; Jaramillo-Correa et al., 2006).

De acuerdo con las proyecciones climáticas a lo largo del tiempo, la región con mayor estabilidad es Mazapil, población que en concordancia con los resultados del segundo capítulo es una de las tres poblaciones más diversas genéticamente. Así, con estas dos evidencias interpretamos que esta región ha funcionado como un refugio para *P. pinceana*. Sin embargo, esta región no ha sido descrita previamente como refugio pleistocénico en otros estudios filogeográficos en el Desierto Chihuahuense (e.g., Vasquez-Cruz & Sosa et al., 2016; Loera et al., 2017; Scheinvar et al., 2017; Gámez et al., 2017).

Los cambios climáticos que han afectado la distribución de *P. pinceana* han ocurrido desde los ciclos interglaciares, y han generado una expansión gradual sobre el Desierto Chihuahuense. Actualmente, las diferencias del clima entre ChD y SMO se dan por cambios en la precipitación y la temperatura. Siendo las poblaciones en ChD suelen ser mucho más secas, con un menor índice de contenido de agua, por lo que muestran una menor acumulación de agua en el suelo durante el año, en comparación con las poblaciones en SMO.

Con los resultados hasta ahora mencionados, la heterogeneidad y regionalización en *P. pinceana* se puede apreciar tanto a nivel de expresión genética, y como a nivel de polimorfismos genéticos en el genoma completo. La divergencia entre los dos linajes se debe a su historia demográfica que ha modelado el espectro de frecuencias, que evidencian esta diferenciación entre las poblaciones. Abonando a esta divergencia, estudios morfofisiológicos han detectado que los individuos del Desierto Chihuahuense tienen una cubierta cerosa más gruesa en las acículas, y un crecimiento más rápido en la raíz primaria y mayor volumen en raíces laterales que individuos de la Sierra Madre Oriental (Martíñón-Martínez et al., 2010; Córdoba-Rodríguez et al., 2011).

La respuesta a la aridez constituye un rasgo poligénico y combina estrategias ecofisiológicas plásticas y adaptativas (Bewley, 1979). El estudio de la adaptación y la



plasticidad conforman uno de los desafíos actuales más interesantes de la biología evolutiva (Seehausen et al., 2014). Por ello, elaborar planteamientos genómicos es importante para entender la organización y efecto en regiones funcionales, codificantes o neutrales. Los estudios de los efectos de la selección pocas veces se han detectado por la fijación de alelos únicos que contribuyan con un efecto mayor, y más frecuentemente se han descrito a través de cambios en las frecuencias de variantes relacionadas funcionalmente (Barrett & Schluter, 2008; Rockman, 2012; Berg & Coop, 2014; Wellenreuther & Hansson, 2016).

La respuesta transcripcional, conjunta y ortóloga, hacia factores bióticos y abióticos en el ambiente natural representa un reto para reconocer *per se* la tolerancia a la aridez. Sin embargo, reconocer patrones de respuesta en árboles adultos, con una longevidad por encima de los 80 años, ya establecidos y bajo las relaciones ecológicas locales naturales, me permitió establecer una base para entender la diversidad y heterogeneidad que concurre en la distribución natural de *P. pinceana*.

### 3. Variación intraespecífica de los pinos piñoneros de conos grandes

Las diferencias genéticas y morfológicas descritas en *P. pinceana* fueron el fundamento de una hipótesis de divergencia que señala que existe un patrón de hibridación y divergencia entre *P. pinceana* y *P. maximartinezii* (Ledig et al., 2001). Con una perspectiva filogenética, en el Tercer Capítulo analicé la divergencia en el clado ancestral de los pinos piñoneros, utilizando comparaciones entre genes altamente conservados y la reconstrucción del genoma del cloroplasto completo. De acuerdo con las reconstrucciones filogenéticas, la divergencia de *Pinus* subsección *Cembroides* ocurrió durante el Mioceno (hace 11 Ma; Gernandt et al., 2008; Saladin et al., 2017; Wei-Tao et al., 2021). La evidencia fósil de pinos piñoneros registra grandes cambios en el rango de distribución durante el Plioceno en el Altiplano Mexicano (Lanner & Van Devender, 1981; Van Devender & Burgess, 1985). Las coníferas tuvieron una distribución más amplia durante el Pleistoceno en México en el Desierto Chihuahuense (Quiñonez-Pérez et al., 2017; Jaramillo-Correa et al., 2006).

Los genomas plastídicos de las especies reportados en el Tercer Capítulo también serán útiles en futuros estudios filogenéticos, y para poder analizar finamente la

diversificación de los pinos piñoneros de la subsección *Cembroides*. Las secciones del genoma plastídico en la región IRB tendrán que compararse con las regiones correspondientes en los genomas plastídicos de otras especies dentro de la subsección.

Para *P. pinceana*, la divergencia entre las poblaciones del Desierto Chihuahuense y la Sierra Madre Oriental es clara, tanto los polimorfismos en el DNA plastídico como los genes nucleares altamente conservados que muestran una divergencia entre los individuos de estas regiones biogeográficas. A nivel de especie, las reconstrucciones filogenéticas por SNPs nucleares concatenados y el genoma plastídico mostraron concordancias con las topologías construidas con otros estudios de la subsección *Cembroides* (Gernandt et al., 2005; Whittall et al., 2010; Montes et al., 2019). Nuestros resultados no pueden distinguir entre el flujo de genes, eventos de captura de cloroplasto o los eventos de introgresión dentro de las muestras de *P. pinceana* y *P. maximartinezii*.

La divergencia entre los dos clados obtenidos tuvo un buen soporte y solo se observó para la muestra de Mazapil un evento de reticulación, que puede ser explicado a partir de las evidencias en las reconstrucciones climáticas durante el Cuaternario, que describen la zona como estable en la distribución de la especie en este intervalo de tiempo, además de ser una de las poblaciones con mayor diversidad genética.

De esta manera, la hipótesis de la introgresión propuesta por Ledig et al., (2001), de acuerdo con nuestros diferentes análisis filogenéticos ha sido descartada. Sin embargo, esta tesis demuestra que existe una variación intraespecífica en *P. pinceana*, a lo largo de su distribución natural. Estos dos grupos, como planteé en el Segundo Capítulo, se han mantenido aislados, que se mantiene por una baja migración entre ellos, y no es un reflejo de eventos de hibridación pasados con su especie hermana *P. maximartinezii*. Hasta el momento no existe aislamiento reproductivo entre los dos linajes de *P. pinceana*, y aunque el flujo genético parece ser limitado, no existe evidencia de la acción de algún mecanismo de especiación, o de un posible efecto de un proceso que potencialmente produzca un cambio adaptativo entre los dos linajes ligado a las diferentes condiciones ambientales presentes en la distribución.

## Conclusiones y perspectivas generales

De acuerdo con la evidencia reportada en los cambios de nivel de expresión génica, es posible concluir que la expresión diferencial a los factores abióticos y bióticos en *P. pinceana* es muy heterogénea en su ambiente natural. Particularmente, la vía de regulación por fitohormonas ABA y JA, y la regulación de ROS fueron las vías de respuesta mejor caracterizadas para el control y regulación del estrés. Sin embargo, las interrelaciones de factores estresantes no hacen posible distinguir propiamente la respuesta exclusivamente en los cambios en la respuesta a la aridez.

Si bien hay una respuesta diferencial a los cambios en el ambiente, a nivel transcripcional parece ser un mecanismo plástico que cubre un espectro ante numerosos factores de estrés. Determinar si esta respuesta es adaptativa requerirá que realizar el análisis bajo condiciones homogéneas, junto con una comparación cruzada de condiciones, además de evaluar la heredabilidad de los fenotipos variables.

A partir de la diversidad genética, identifique dos grupos genéticos considerando regiones aleatorias del genoma, genes altamente conservados y el plastoma completo. La divergencia puede explicarse por el efecto de procesos filogeográficos históricos que han dejado huella en el patrón de diversidad, y es posible que esta diferenciación no incluya procesos adaptativos.

La diversidad genética en *P. pinceana* fue afectada por la dinámica interglaciar, que dividió a los clústers genéticos durante los cambios climáticos del Pleistoceno medio relacionados con los ciclos glaciario-interglaciario (~627 kya), seguido de la subfragmentación del Desierto Chihuahuense y de la Sierra Madre Oriental (~127.7 - ~539.2 kya), mientras que la aridificación aumentaba y los paleolagos reducían su área.

Aparentemente, la población de Mazapil tuvo una alta estabilidad de hábitat durante el Pleistoceno medio y de acuerdo con la diversidad genética identificada, interpreto que esta región representó un refugio durante los ciclos interglaciares. Esta hipótesis puede también explicar la reticulación entre las construcciones filogenéticas de los dos clústers genéticos entre las regiones biogeográficas.

Las relaciones filogenéticas a partir del plastoma y la concatenación de SNPs en genes altamente conservados recapturan reconstrucciones previas de la divergencia de los piñoneros de conos grandes. Sin embargo, esta tesis logró incorporar la divergencia de los grupos genéticos, descartando en cualquiera de ellos eventos de introgresión con *P. maximartinezii* como previamente se había sugerido.

Entre los dos clústers genéticos no existen evidencias de aislamiento reproductivo, ya fuera como una respuesta transcripcional o por diferencias genéticas debidas a la selección natural o a algún mecanismo adaptativo local, o bien, un proceso de especiación. Los grupos genéticos parecen ser resultado de la historia demográfica y de los cambios climáticos durante el Pleistoceno medio, lo que ha generado la diferenciación de las poblaciones.

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